Inhibition of autophagy limits vertical transmission of Zika virus in pregnant mice

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Zika virus (ZIKV) infection during pregnancy leads to devastating fetal outcomes, including intrauterine growth restriction and microcephaly. Greater understanding of mechanisms underlying ZIKV maternal-fetal transmission is needed to develop new therapeutic interventions. Here, we define an important role for the autophagy pathway in ZIKV vertical transmission. ZIKV infection induced autophagic activity in human trophoblasts and pharmacological inhibition limited ZIKV infectivity. Furthermore, deficiency in an essential autophagy gene, Atg16l1, in mice limited ZIKV vertical transmission and placental and fetal damage and overall improved placental and fetal outcomes. This protection was due to a placental trophoblast cell-autonomous effect of autophagic activity, not to alterations in systemic maternal ZIKV infection. Finally, an autophagy inhibitor, hydroxychloroquine, approved for use in pregnant women, attenuated placental and fetal ZIKV infection and ameliorated adverse placental and fetal outcomes. Our study reveals new insights into the mechanism of ZIKV vertical transmission and suggests that an autophagy-based therapeutic warrants possible evaluation in humans to diminish the risks of ZIKV maternal-fetal transmission.

INTRODUCTION
Zika virus (ZIKV) is an emerging mosquito-transmitted flavivirus and global public health threat. Hitherto, ZIKV was known to cause a self-limiting febrile illness characterized by rash, myalgia, conjunctivitis, and headache. However, recent epidemics have linked ZIKV infection to Guillain-Barré syndrome in adults (Cao-Lormeau et al., 2016) and placental insufficiency, fetal demise, microcephaly, and other congenital malformations in fetuses and newborn infants in the setting of maternal infection during pregnancy (Chibueze et al., 2017). In the human placenta, ZIKV has a broad cell tropism, including villous placental trophoblasts, endothelial cells, fibroblasts, and fetal Hofbauer macrophages in the intervillous space (El Costa et al., 2016; Jurado et al., 2016; Miner et al., 2016; Quicke et al., 2016; Tabata et al., 2016; Aagaard et al., 2017). This evidence suggests that ZIKV can disseminate into the intrauterine space and infect the fetus through a transplacental route.

Recent studies have demonstrated that inoculation of ZIKV through a subcutaneous or intravaginal route in pregnant mice with compromised type I interferon signaling results in severe placental and fetal damage, including intrauterine growth restriction and fetal demise (Miner et al., 2016; Yockey et al., 2016), which recapitulates many features of congenital ZIKV syndrome in humans (Mysorekar and Diamond, 2016; Cao et al., 2017). Furthermore, in situ hybridization (ISH) and viral antigen staining has established the presence of ZIKV infection in multiple mouse and human trophoblast cells (Miner et al., 2016; Tabata et al., 2016; Cao et al., 2017; Vermillion et al., 2017). The preferential replication of ZIKV in trophoblasts and fetal endothelial cells in the placenta suggests that ZIKV enters into the fetal circulation in mice in part by compromising the placental barrier. Bayer et al. (2016) showed that type III interferon (IFN-λ) secreted by primary cultured human trophoblast cells was able to protect trophoblasts from ZIKV infection in an autocrine or paracrine manner. TIM1, a member of the T cell immunoglobulin and mucin domain protein family, has been shown to play a critical role in ZIKV infection in primary trophoblasts (Tabata et al., 2016). However, the pathogenesis and underlying mechanisms of ZIKV-induced maternal-fetal transmission in vivo across the placental trophoblast barrier remain unknown.

The placenta normally uses physical and immunological strategies to protect the fetus from maternal-fetal transmission of pathogens. One important mechanism the placenta uses in its defense is autophagy (Delorme-Axford et al., 2013; Cao et al., 2016), which targets intracellular components for lyosomal degradation and is important for host defense against many pathogens (Levine et al., 2011). Notwithstanding this, some pathogens have evolved mechanisms to evade, inhibit, or even hijack the host autophagy machinery to facilitate infection and survival (Cemma and Brumell, 2012). Recent studies have indicated that autophagy has an important function in placental defense against microbial agents (Delorme-Axford et
and provide a foundation for developing therapeutics to re-
central damage and fetal death. Our findings demonstrate that
(HCQ) reduced ZIKV vertical transmission and limited pla-
with these findings, we demonstrate that inhibiting auto-
deficient mice reduced ZIKV infection in placentas and fe-
cells and in the mouse placenta. Inhibition of the autophagy
infection activates autophagic activity in human trophoblast
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phagy on vertical transmission of ZIKV. We show that ZIKV
infection activates autophagic activity in human trophoblast
cells and in the mouse placenta. Inhibition of the autophagy
pathway in both human trophoblast and autophagy gene-
deficient mice reduced ZIKV infection in placentas and fe-
tuses and resulted in improved fetal outcomes. Consistent
with these findings, we demonstrate that inhibiting au-
phagy by treating pregnant mice with hydroxychloroquine
(HCQ) reduced ZIKV vertical transmission and limited pla-
cental damage and fetal death. Our findings demonstrate that
autophagy promotes ZIKV pathogenesis during pregnancy
and provide a foundation for developing therapeutics to re-
duce maternal-fetal transmission of ZIKV.

RESULTS
ZIKV infection induces autophagic activity in human trophoblasts

Autophagy is a vital part of the host response to many mi-
crobial infections. Thus, we reasoned that ZIKV may regulate
autophagic activity in trophoblasts cells to facilitate its repli-
cation in the placenta. We infected a human cytotrophoblast
(CTB) cell line, JEG-3, with a Brazilian strain of ZIKV (Para-
ibas 2015) at a multiplicity of infection of 0.1 and collected
samples 6, 12, 24, or 48 h later for virus titration as described
previously (Miner et al., 2016). We examined the level of au-
ophagy markers in CTBs after ZIKV infection by Western
blot for the microtubule-binding protein light chain 3 (LC3)
protein, which converts from the soluble form LC3-I to the
lipidated form LC3-II and serves as an indicator of au-
thapic activity or flux. We found that LC3-II was significantly
increased at 6 and 12 h postinfection (hpi; Fig. 1 A and Fig.
S1 A). Because this could indicate either increased autophagy or
inhibition of autophagosome maturation, we next treated
cells with bafilomycin (Baf) A1, which inhibits autophagosom-
al and lysosomal fusion (Klionsky et al., 2016). In this case,
LC3-II further accumulated in ZIKV-infected cells, indicat-
ing that ZIKV activated autophagy (Fig. 1 A and Fig. S1 A).

ZIKV infection has been associated with LC3-II–positive
autophagosome formation in skin fibroblasts (Hamel et al.,
2015) and neural stem cells (Liang et al., 2016). To identify
whether LC3+ autophagosome formation was affected in
trophoblasts infected with ZIKV, we transfected CTBs with a
plasmid carrying EGFP-LC3 and subsequently exposed them
to ZIKV. Immunofluorescence staining for GFP revealed
diffuse LC3 staining in uninfected CTBs and puncta-
ting in ZIKV-infected CTBs (Fig. 1 B). The number of
EGFP-LC3+ punctae remained higher in ZIKV-infected
than in uninfected cells upon Baf A1 treatment (Fig. 1, B and
C), indicating that the increase in LC3 punctae was due to
enhanced autophagosomal formation. These results demon-
strate that ZIKV infection induces canonical autophagy re-
sponse in human trophoblasts.

Suppression of autophagic activity impairs ZIKV infection in human trophoblasts

Manipulation of autophagy pathway activity can alter mi-
crobial infection and pathogenesis in other cells and tissues
(Levine et al., 2011). We next determined whether increas-
ing or decreasing baseline autophagy in trophoblasts altered
ZIKV persistence in these placental cells. We infected JEG-3
CTBs with ZIKV and then treated infected and control cells
with chemical modulators of autophagy pathways: rapamy-
cin and Torin 1 induce autophagy, whereas 3-methyladenine
(3-MA), chloroquine (CQ), and Baf A1 inhibit autophagy at
initiation (3-MA and CQ) and autophagolysosome forma-
tion (Baf A1) stages. Importantly, these autophagy modulators
did not induce changes in CTB viability, with the exception
of Torin 1 treatment, which was associated with a small re-
duction in cell viability (Fig. S1 B). Treatment of CTBs with
the autophagy inhibitors 3-MA, CQ, and Baf A1 resulted in a
significant decrease in ZIKV replication at 48 hpi (Fig. 1 D).
Reciprocally, administration of the autophagy inducers rapa-
ymycin and Torin 1 resulted in an increase of viral infection in
CTBs (Fig. 1 D). Immunofluorescence staining in infected
trophoblasts confirmed fewer ZIKV antigen–positive cells
(Fig. 1, E and F) after treatment with autophagy inhibitors
but greater numbers of antigen–positive cells after incubation
with autophagy inducers (Fig. 1, E and F). Collectively, these
data show that inhibition of autophagy limits ZIKV infection
in trophoblasts and suggests that modulation of this pathway
can be harnessed to combat this infection.

ZIKV infection induces autophagy in vivo, and loss of ATG16L1 expression impairs in utero transmission of ZIKV

To investigate the physiological role for autophagy in the
context of an in vivo model of maternal-fetal ZIKV trans-
mision, we assessed whether ZIKV induced autophagy in the
placenta. Similar to the human trophoblast response to ZIKV
infection, immunoblotting for LC3-II in ZIKV-infected
whole placentas (5 d postinfection) showed increased levels
compared with uninfected placentas (Fig. S2 A). We also de-
tected a concomitant reduction in levels of p62 (also called
sequestome-1 [Sqstm1]), a substrate degraded by autophagy
pathway (Klionsky et al., 2016; Fig. S2 A). Immunohistochem-
atical staining revealed that the ZIKV infection–associated reduction in p62 levels, thereby supporting autophagy pathway activation, was localized primarily to the trophoblast-rich region of the placenta (Fig. S2 B). Thus, ZIKV infection in vivo can activate autophagic activity in the placenta.

On the basis of these findings, we hypothesized that genetic manipulation of autophagic activity would alter ZIKV pathogenesis in pregnancy. To test this hypothesis, we used mice hypomorphic (HM) for a key autophagy gene, Atg16L1 (Atg16l1HM), because (1) ATG16L1 has a key role in autophagosome maturation as part of the protein complex that directs LC3 to autophagosomes en route to fusion with lysosomes, (2) Atg16l1HM mice exhibit stalled autophagosome formation and reduced autophagy in multiple tissues, and (3) Atg16l1HM mice exhibit normal fertility, and their placentas are histologically indistinguishable from wild-type (WT) mice in the absence of infection (Cao et al., 2016). We inoculated pregnant WT and Atg16l1 HM mice (Atg16l1HM) with 10^3 focus-forming units (FFU) of ZIKV (Paraiba 2015) through a subcutaneous route at day E9.5, 1 d after pretreatment with a single dose of anti-Ifnar1 to facilitate dissemination (Fig. 2 A), as reported previously (Miner et al., 2016). Pregnant dams were followed longitudinally for morbidity, levels of maternal viremia, and viral burden in the placenta and fetal heads at E14.5. Notably, ZIKV infection in Atg16l1-deficient placentas was ~10-fold lower compared with WT controls (Fig. 2 B). ISH revealed that the overall abundance of ZIKV RNA–positive placental trophoblasts was decreased in Atg16l1HM placentas (Fig. 2 C). The reduced viral burden in Atg16l1HM placentas was associated with decreased placental damage: pathological phenotypes observed in ZIKV-infected WT placentas, such as apoptotic trophoblasts and increased number of nucleated fetal erythrocytes, were reduced in Atg16l1HM placentas (Fig. 2 D). Further histological analysis showed that ZIKV-infected Atg16l1HM placentas had larger and thicker placental layers than WT placentas (Fig. 2 E), and the labyrinth zone thickness was greater as well (Fig. 2 F).

Finally, ZIKV titer in Atg16l1-deficient fetus was significantly lower (~15-fold) compared with WT controls (Fig. 3 A). Moreover, fetuses in the Atg16l1HM group at E14.5 were larger than WT counterparts, indicating that the fetal growth restriction induced by ZIKV infection was limited by the HM Atg16l1 allele (Fig. 3 B). These data together indicate that a deficiency in Atg16l1 function results in reduced placental and fetal infection of ZIKV, thus suggesting that...
ATG16L1 plays an important role in governing placental susceptibility to ZIKV infection and maternal-fetal transmission.

**Atg16l1 deficiency in trophoblasts limits ZIKV infection in the placenta**

To assess whether the observed virological phenotype in Atg16l1^Hm^ mice was due in part to changes in maternal infection, we measured viral burden in maternal serum and spleen but observed no significant differences in between the pregnant Atg16l1^Hm^ and WT mice (Fig. 4, A and B). Moreover, the HM Atg16l1 allele had no effect on ZIKV RNA localization or abundance within the maternal decidua (Fig. 4 C). Consistent with these data, no difference in viral titers were noted in nonpregnant Atg16l1^Hm^ mice infected with ZIKV (Fig. 4, D and E) or in infected male Atg16l1^Hm^ mice (not depicted). These findings indicate that a loss of Atg16l1 function has no little or no effect on systemic or maternal decidual infection but preferentially affects infection of the fetal-derived placental compartments.

To confirm these findings, we assessed whether loss of Atg16l1 exclusively in cells of the trophoblast lineage was sufficient to limit ZIKV maternal-fetal transmission. We generated mice that lack Atg16l1 only in trophoblast lineage cells by crossing the Cyp19-promoter-driven Cre^-^recombinase mice (Wenzel and Leone, 2007) with Atg16L1^fl/fl^ mice. Pregnant Cyp 19 Cre^+^ Atg16L1^fl/fl^ mice and Cre^-^ control mice were treated with anti-Ifnar1 as described above and subsequently inoculated subcutaneously at E9.5 with 10^3^ FFU of ZIKV (Paraiba 2015; Fig. 5 A). Analysis of the placentas from Cyp 19 Cre^+^ Atg16L1^fl/fl^ mice revealed that loss of this gene in trophoblasts resulted in reduced ZIKV infection in the placenta, similar to that seen with the Atg16l1^Hm^ mice (Fig. 5, B and C). Unexpectedly, these differences in placental infection did not translate into reductions in fetal head titers.
(Fig. 5 D) or increases in fetus size (not depicted), suggesting that ATG16L1 may be important for viral survival or replication in other cell types (e.g., fetal brain neurons, Hofbauer cells at the maternal-fetal interface).

Pharmacological inhibition of autophagy results in decreased ZIKV infection at the maternal-fetal interface

Given that a reduction of autophagic activity in human trophoblasts and genetic loss of function of Atg16l1 in mice limited ZIKV infection, we reasoned that an existing drug that inhibited autophagy and could be administered in pregnancy might have immediate therapeutic utility. HCQ is a Food and Drug Administration–approved class C drug that is used clinically to treat pregnant patients with malaria or autoimmune diseases (Kaplan et al., 2016). As systemic administration of HCQ has been successfully used to dampen autophagic activity in vivo (Rosenfeldt et al., 2013), we tested its efficacy against ZIKV infection during pregnancy. WT pregnant mice were administered HCQ (40 mg/kg/d) via intraperitoneal injection beginning at day +1 (E10.5) after ZIKV infection (Fig. 6 A); this dosage has known in vivo inhibitory effects on autophagy (Rosenfeldt et al., 2013) and was not associated with any differences in litter size compared with PBS-treated mice (Fig. S3 A). HCQ treatment significantly increased the levels of p62 (indicative of reduced autophagic activity) in trophoblasts, which validates the inhibitory effects of HCQ on autophagy in the placenta (Fig. 6 B). We next evaluated the effect of HCQ treatment during ZIKV infection in pregnancy. Notably, HCQ-treated mouse placentas sustained lower levels of ZIKV infection compared with PBS-treated controls (Fig. 6 C).

Although several groups have observed that treatment with HCQ or the related CQ can reduce flavivirus infection in cell culture, possibly through effects on viral fusion or maturation through inhibition of acidification of intracellular vesicles (Farias et al., 2014; Delvecchio et al., 2016), its function in vivo is less clear, as a randomized clinical trial of CQ treatment of Dengue virus infection showed no benefit in adults from Vietnam (Savarino et al., 2003; Tricou et al., 2010). To ensure that the reduction in viral burden in the placenta was not due to reduced infection titers in the peripheral maternal tissues, we analyzed viral titers in the pregnant dams. No differences in viral burden in the serum, spleen, or maternal decidua were observed between the HCQ- and PBS–treated groups (Fig. 6 D and E; and Fig. S3 B). However, ISH staining for ZIKV RNA showed a marked reduction of ZIKV RNA in the placentas of HCQ-treated mice compared with controls (Fig. 6 F). Thus, the reduction in ZIKV infection by HCQ treatment in the context of pregnancy was specific to the placenta.

Histological analysis of HCQ-treated ZIKV-infected mice showed improvement in placental areas with reduced placental damage, as indicated by decreased numbers of apoptotic trophoblasts and immature fetal erythrocytes (Fig. 6, G and H). Similar to the results noted in Atg16l1HM mice,
HCQ treatment reversed the reduction in area of the labyrinth (Fig. 6 I). Thus, HCQ treatment rescues the placental insufficiency that occurs after ZIKV infection. Consistent with a reduction in placental infection and improved tissue disease, HCQ treatment reduced ZIKV infection in the fetal head, which was associated with larger fetal body size (Fig. 6, J and K). Finally, we sought to confirm that this improvement in fetal status was due to inhibition of autophagy by treating ZIKV-infected Atg16l1<sup>fl/fl</sup> mice with HCQ. As expected, there was no difference in placental or fetal ZIKV titer in Atg16l1<sup>fl/fl</sup> mice with or without HCQ treatment (Fig. S3, C and D). Thus, our findings indicate that HCQ administration in pregnant dams can reverse the ZIKV associated autophagy induction in the placenta and reduce placental insufficiency, thus limiting vertical transmission of ZIKV infection and fetal growth defects.

**DISCUSSION**

The placenta acts as barrier to protect the developing fetus from the invading pathogens. In previous studies, we showed that autophagy protects against bacterial infection of the placenta (Cao et al., 2016). Here, in the context of ZIKV, we demonstrate the opposite: blocking the autophagy pathway in the placenta restricted ZIKV infection in the placenta and fetus during pregnancy. Most important, we demonstrated that placental trophoblast specific knockout of a key autophagy gene, Atg16l1, was sufficient to limit ZIKV placental infectivity. To our knowledge, our work is the first to show a placental cell–derived mechanism governing susceptibility to maternal–fetal transmission of ZIKV. These findings enhance our understanding of the cellular mechanisms of ZIKV maternal–fetal transmission and a possible therapy for enhancing the placental barrier function and controlling ZIKV infection.

Flaviviruses have been known to modulate autophagy in the context of infection (Pirooz et al., 2014). Different flaviviruses appear to have evolved distinct abilities to avoid or use autophagy to promote survival or infection at different stages of the viral life cycle, including viral replication, assembly, and release (Chiramel et al., 2013). For example, West Nile virus infection does not induce LC3 lipidation, and depletion of ATG5, a binding partner of ATG16L1, does not affect its replication (Vandergaast and Fredericksen, 2012; Martin-Acebes et al., 2015). However, administration of Tat-beclin-1, an autophagy-inducing peptide, reduces neuronal infection, cell death, and mortality associated with West Nile virus intracranial inoculation (Shoji-Kawata et al., 2013). In contrast, a deficiency in ATG5 in mouse neuroblastoma cells resulted in higher viral replication of the Japanese encephalitis virus (Sharma et al., 2014), suggesting that autophagy restricts infection. Others have shown that inducers of autophagy can lead to increased Dengue virus burden and pathogenicity in mice (Mateo et al., 2013), with a requirement of autophagy for optimal viral RNA replication and virion maturation (Mateo et al., 2013).

In vitro evidence has suggested that the autophagy pathway can modulate ZIKV infection: ZIKV was shown to induce autophagy in skin fibroblasts (Hamel et al., 2015) and human fetal neural stem cells (Liang et al., 2016), and this was associated with enhanced replication. However, in vivo evidence for such effects, especially on maternal–fetal transmission of ZIKV, has been lacking. We show that a genetic or pharmacologically induced deficiency in autophagy impairs
ZIKV infection in the placenta and fetus without affecting systemic infection of other maternal organs. Thus, inhibition of autophagy is beneficial for the host to limit ZIKV maternal-fetal infection, which contrasts with its host defense roles in other placental infection models (Delorme-Axford et al., 2013; Cao et al., 2016). This finding suggests that ZIKV has
evolved strategies to usurp autophagy pathway for its own replicative advantage, but that this effect is restricted to certain cell types. It remains unclear why the impact of autophagy on ZIKV infection is more apparent in placental cells compared with other cell types. Our current work provides in vivo evidence that decreased autophagic activity is beneficial for the host in fighting ZIKV maternal-fetal transmission and indicates why ZIKV would induce autophagy. Recent studies have addressed the mechanisms of how ZIKV may have this effect. A study investigating ZIKV pathogenesis in human neurospheres showed that two nonstructural proteins of ZIKV, NS4A and NS4B, induced autophagy by suppressing the Akt-mTOR pathway (Liang et al., 2016). Moreover, ZIKV NS3 protease can cleave FAM134, an ER-localized protein required for reticulophagy, a selective form of autophagy that leads to ER degradation (Lennemann and Coyne, 2017).

Devastating fetal and neonatal outcomes caused by the recent ZIKV epidemic highlight the importance of studying mechanisms underlying maternal-fetal transmission of ZIKV. Placental trophoblast barriers contribute to defense responses to ZIKV vertical transmission. We showed that HCQ can successfully reduce viral burden both in human trophoblast cells and in mouse placentas. Whereas HCQ treatment leads to an increase in pH in intracellular vesicles, including lysosomes, and autophagosomes, and CQ has been shown to have antiviral activity in cell culture by inhibiting viruses including ZIKV at different stages of the viral life cycle (Savarino et al., 2003; Delvecchio et al., 2016), our p62 expression data suggest that the anti-ZIKV activity after HCQ treatment in vivo is due to modulation of autophagy. Consistent with this idea, other autophagy inhibitors, (e.g., 3-MA), which target different stages of autophagy, showed anti-ZIKV activity, whereas treatment with the autophagy activators rapamycin and Torin 1 resulted in increased viral infection in cultured trophoblasts. Moreover, HCQ treatment in mice phenocopied the reduced viral burden phenotype observed in mice deficient in Atg16L1, with decreased infection in the placenta and fetus but not in maternal tissues, which might be expected by a drug that was affecting a key stage in the viral life cycle. In addition, HCQ-induced inhibition of autophagy had no discernible impact on ZIKV infection of Atg16L1-deficient mice. These findings suggest that inhibition of autophagy specifically impairs ZIKV infection of the placenta, which results in reduced infection and disease in the fetuses.

Given the epidemic nature of ZIKV infection and its potential for congenital malformations, immediate interventions to prevent or treat ZIKV infection, especially at the maternal–fetal interface, are urgently required. Although several studies have shown promising anti-ZIKV activity with clinically approved drugs (Barrows et al., 2016; Delvecchio et al., 2016; Elifyk, 2016; Eyer et al., 2016; Xu et al., 2016; Zmurko et al., 2016; Sacramento et al., 2017), most of these experiments were performed in cell culture, and of the few showing in vivo efficacy, none of the studies were performed in the setting of pregnancy. Although mouse placentas are anatomically different from human and nonhuman primate placentas (Rossant and Cross, 2001), the placental trophoblast barrier functions analogously. Our experiments on ZIKV infection inhibition in a mouse model of pregnancy are applicable to the understanding of ZIKV infection pathogenesis and may provide the foundation for experiments in nonhuman primates and clinical trials in humans to further define the therapeutic effects of modulating autophagic activity on ZIKV congenital disease.

Additionally, a limitation of many of the drugs (e.g., PHA-690509, a cyclin-dependent kinase inhibitor) is that they may not be suitable for administration during pregnancy because of their own potential teratogenic effects. HCQ is a Food and Drug Administration–approved class C drug that has been used to treat malaria and autoimmune diseases during pregnancy without apparent injury to the fetus, although it has not been evaluated in a randomized controlled trial. The current recommendation is that the potential benefits may warrant HCQ use in pregnant women despite potential risks, depending on the target disease. Our results suggest that a pharmacological inhibitor of autophagy warrants possible evaluation in nonhuman primates and humans during pregnancy to diminish the risks of ZIKV infection and disease in developing fetuses.

MATERIALS AND METHODS

Ethics statement

All animal procedures were reviewed and approved by the Institutional Animal Care and Use Committee at the Washington University School of Medicine. Inoculations and dissections were performed under anesthesia to minimize animal suffering.

Viruses and titration

The Brazilian strain of ZIKV (Paraiba 2015) was provided by S. Whitehead (Bethesda, MD) and originally obtained from P.E.C. Vasconcelos (Instituto Evandro Chagas, Levilândia, Brazil). Virus stocks were propagated in Vero cells. The titers of ZIKV stocks were determined by focus-forming assay (FF on Vero cells as described previously; Miner et al., 2016). Studies with ZIKV were conducted under biosafety level 2 and animal biosafety level 3 containment.

Cell culture and infection

JEG-3 cells were obtained from ATCC (HB-36) and cultured in F12/DMEM media supplemented with 10% FBS (Thermo Fisher Scientific) at 37°C with 5% CO2. JEG-3 cells were infected with ZIKV at a multiplicity of infection of 0.1 for 2 h, washed twice with warm PBS, and cultured in fresh medium with indicated treatments. At indicated time points, supernatants were harvested for virus titration or fixed for immunofluorescence staining of ZIKV using a ZIKV-specific mAb, ZIKV-2, as previously described (Miner et al., 2016). For monitoring autophagic flux, JEG-3 cells were transiently transfected with an EGFP-LC3 plasmid (Addgene;
deparaffinized tissues were quenched with 3% H2O2, blocked, and stained with hematoxylin and eosin to assess morphology. Histologic images were captured by use of a Nikon Eclipse microscope equipped with an Olympus DP71 color camera under 2×, 20×, and 40× objectives. Measurement of size and thickness of different placental layers were performed using ImageJ (National Institutes of Health).

For immunohistochemical staining of mouse placentas, deparaffinized tissues were quenched with 3% H2O2, blocked for 2 h, and incubated with primary antibodies anti-p62 (1:500; ab56416; Abcam) overnight. The M.O.M. kit (BMB-2202; Vector Laboratories) was applied for blocking and secondary antibody incubation according to the manufacturer’s instructions. The sections were then incubated with the ABC reagent from the Vectastain universal kit (PK-7200; Vector Laboratories) and developed using the DAB substrate kit (SK-4100; Vector Laboratories). Tissues were counterstained with hematoxylin. A no–primary antibody staining was included as a negative control.

Mouse experiments

Atg16l1Hm and Atg16l1flox/flox mice were provided by H. Virgin (Washington University, St. Louis, MO; Cadwell et al., 2008). Cyp19-Cre transgenic mice were a kind gift from Gustavo Leone (The Ohio State University, Columbus, OH) and were generated as previously described (Wenzel and Leone, 2007). All mice were on a C57BL6 background and housed under a 12-h light/12-h dark cycle in a specific pathogen–free mouse breeding facility. For mating, 8- to 10-wk-old male and nulliparous female mice were housed from 5 p.m. to 8 a.m., and day 0.5 of pregnancy was defined as the first observation of a vaginal plug. Pregnant mice were inoculated by subcutaneous route in the footpad with 105 FFU of ZIKV in a volume of 50 µl. One day before infection, mice were treated with a single dose of 2 mg of an Ifnar1-blocking mouse antibody (MAR1-5A3, purchased from Leinco, Inc.) by intraperitoneal injection. Fetal size were measured by crown-rump length and occipital-frontal diameter at 4 d postfixation.

Measurement of viral burden

ZIKV-infected pregnant mice were euthanized on E14.5, and fetoplacental units were dissected. Placentas and fetal heads were weighed and homogenized in 250 or 500 µl PBS. All homogenized tissues from infected animals were stored at −80°C until virus titration. RNA from tissue samples was extracted with the RNeasy Mini kit (QIAGEN). Serum from ZIKV-infected mice and culture medium from ZIKV-infected JEG-3 cells were extracted with Viral RNA Mini kit (QIAGEN). ZIKV RNA copy number was determined by one-step quantitative RT-PCR on an ABI 7500 Fast Instrument using standard cycling conditions as described previously (Miner et al., 2016). The following primers and probe targeting ZIKV were used: 5′-CCACATGTCTCTGCA GACATATG-3′; reverse: 5′-TTGACGAGCGTGCACAC AG-3′; and probes: 5′-56-FAM/AGCCTCCCTTGACAA GCAGTGC3IABkFQ-3′ (Integrated DNA Technologies).

Histology and immunohistochemistry staining

Placentas and fetuses were dissected from cesarean section on E14.5. Harvested samples were fixed in 10% neutral buffered formalin (Thermo Fisher Scientific) at room temperature and embedded in paraffin. At least five placentas from different dams with the indicated genotypes or treatments were sectioned and stained with hematoxylin and eosin to assess morphology. Histologic images were captured by use of a Nikon Eclipse microscope equipped with an Olympus DP71 color camera under 2×, 20×, and 40× objectives. Measurement of size and thickness of different placental layers were performed using ImageJ (National Institutes of Health).

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RNA ISH

RNA ISH was performed with an RNAscope 2.5 (Advanced Cell Diagnostics) according to the manufacturer’s instructions. In brief, formalin-fixed paraffin-embedded tissue sections were deparaffinized and incubated at 60°C for 1 h. H2O2 was applied to quench endogenous peroxidases for 10 min at room temperature. Slides were then boiled in RNAscope Target Retrieval Reagents for 15 min and incubated for 30 min in RNAscope Protease Plus. The ZIKV probe (catalog #467871), positive probe (the plr2a probe; catalog #312471) and negative probe (targeting bacterial gene dapB; catalog #310043) were designed and synthesized by Advanced Cell Diagnostics. Tissues were counterstained with hematoxylin and mount. Images were captured with standard bright-field microscopy under 2× and 20× objectives.

Western blotting

Total protein was extracted from frozen mouse placenta samples and homogenized in RIPA buffer (Cell Signaling Technology) with protease inhibitor cocktails (Sigma-Aldrich). For cell cultures, cells were lysed directly in RIPA buffer. Equivalent amounts of total protein, determined by BCA assays (Thermo Fisher Scientific), were separated on 4%-20% Mini-PROTEAN precast gels (Bio-Rad Laboratories) and transferred to polyvinylidene fluoride membranes. Membranes were blocked in 5% milk in PBS with Tween 20 for 2 h at room temperature and incubated with indicated antibodies in 5% BSA at 4°C overnight. The following primary antibodies and dilutions were used: anti-LC3B (1:2,000; Cell Signaling Technology), anti-p62 (1:1,000; ab56416; Abcam), and anti-GAPDH (1:2,000; 3700s; Cell Signaling Technology). ImageJ was used for densitometry of Western blots.

Statistical analysis

GraphPad Prism 5.0 was used for all analyses. The analyses of virologic or histopathological data were conducted using a Mann–Whitney test or ANOVA with a multiple comparison test. P-values < 0.05 were considered to indicate statistical significance.

Online supplemental material

Supplemental material associated with this study provides supporting data for additional time points of ZIKV infection of human trophoblasts, increased autophagic activity in
mouse placenta, and that HCQ treatment of Atg16l1HM mice does not alter ZIKV infection. Fig S1 shows ZIKV infection in human trophoblasts. Fig S2 shows that ZIKV infection induces autophagic activity in mouse placenta. Fig. S3 shows that HCQ treatment reduces placental and fetal ZIKV infection specifically via autophagy.

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REFERENCES


SUPPLEMENTAL MATERIAL

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Figure S1. **ZIKV infection in human trophoblasts.** (A) Western blots for LC3-I/II show enhanced autophagy flux upon ZIKV infection at 6 hpi. ZIKV–infected JEG-3 (multiplicity of infection 0.1, 2 h) were cultured for 6 h and harvested. Baf A1 was applied for 30 min before harvesting to monitor autophagic flux. Images represent data from four independent experiments. (B) Cell viability assays of uninfected JEG-3 cells cultured in medium supplemented with indicated autophagy modulators (at same dosage as described in Fig. 1 F) for 48 h; n = 4 for each group. Data depict mean ± SEM. *, P < 0.05; ns, not significant (ANOVA with a Dunn's multiple-comparison test). Rap, rapamycin.

Figure S2. **ZIKV infection induces autophagic activity in mouse placentas.** (A) Immunoblot and quantification of LC3 and p62 show increased autophagic activity in mouse placentas (at E14.5) infected with ZIKV compared with uninfected controls. GAPDH, a loading control; n = 3–5. Data depict mean ± SEM. *, P < 0.05, Mann-Whitney test. (B) Representative immunohistochemical staining of p62 in WT placentas with or without ZIKV infection at E14.5. Images represent data from five independent dams. Bars, 100 µm.
Figure S3. **HCQ treatment reduces placental and fetal ZIKV infection specifically via autophagy.** (A) Litter size at E14.5 with or without HCQ treatment; n = 6 per group. Results represent mean ± SEM, three independent experiments. ns, not significant, Mann-Whitney test. (B) Representative ISH images of ZIKV RNA in indicated maternal uterine decidua. Bars, 100 µm. (C and D) Pregnant HM female mice were treated with HCQ (40 mg/kg/day via intraperitoneal route) or DMSO as a mock control from day +1 post–ZIKV infection (E10.5) to E14.5. Viral burden of ZIKV in placentas (C) and fetal head (D) were measured by quantitative RT-PCR; n = 8 from three independent dams. ns, not significant, Mann-Whitney test.