Lymphoid tissue phospholipase A2 group IID resolves contact hypersensitivity by driving antiinflammatory lipid mediators

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Resolution of inflammation is an active process that is mediated in part by antiinflammatory lipid mediators. Although phospholipase A2 (PLA2) enzymes have been implicated in the promotion of inflammation through mobilizing lipid mediators, the molecular entity of PLA2 subtypes acting upstream of antiinflammatory lipid mediators remains unknown. Herein, we show that secreted PLA2 group IID (PLA2G2D) is preferentially expressed in lymphoid tissue and surface MHC class II expression in LN cells or DCs. Altogether, our results highlight PLA2G2D in the LNs contributed to mobilization of a pool of polyunsaturated fatty acids that could serve as precursors for antiinflammatory/pro-resolving lipid mediators such as resolvin D1 and 15-deoxy-Δ12,14-prostaglandin J2, which reduced Th1 cytokine production. In hapten-induced contact dermatitis, resolution, not propagation, of inflammation was compromised in skin and LNs of PLA2G2D-deficient mice (Pla2g2d−/−), in which the immune balance was shifted toward a proinflammatory state over an antiinflammatory state. Bone marrow–derived DCs from Pla2g2d−/− mice were hyperactivated and elicited skin inflammation after intravenous transfer into mice. Lipidomics analysis revealed that PLA2G2D in the LNs contributed to mobilization of a pool of polyunsaturated fatty acids that could serve as precursors for antiinflammatory/pro-resolving lipid mediators such as resolvin D1 and 15-deoxy-Δ12,14-prostaglandin J2, which reduced Th1 cytokine production and surface MHC class II expression in LN cells or DCs. Altogether, our results highlight PLA2G2D as a "resolving sPLA2" that ameliorates inflammation through mobilizing pro-resolving lipid mediators and points to a potential use of this enzyme for treatment of inflammatory disorders.

In many acute and chronic disorders, unresolved inflammation is a major mechanism of disease pathogenesis (Libby, 2002). Inflammation is a protective host response to foreign challenge or tissue injury that, if unopposed, could lead to loss of tissue structure and function. Many inflammatory processes have self-limiting and self-resolving systems, which are regulated by antiinflammatory or pro-resolving mediators during the course of inflammation (Serhan et al., 2008). Uncontrolled balance between pro- and anti-inflammatory systems is a hallmark of chronic inflammatory disorders.
Figure 1. Expression of PLA2G2D in CD11c+ DCs. (A) Quantitative RT-PCR of Pla2g2d relative to Gapdh in various tissues of C57BL/6 mice (mean ± SEM from four mice, pooled from two experiments). (B) Quantitative RT-PCR of sPLA2 mRNAs relative to Gapdh in cells from mouse spleen, thymus, and LNs (mean ± SEM from five mice, pooled from two experiments). (C) Immunohistochemistry of mouse spleen with anti–mouse PLA2G2D or control antibody. A boxed area is magnified. (D) Quantitative RT-PCR of Pla2g2d and Pla2g5 relative to Rn18s in several splenic cell populations (mean ± SEM from...
antiinflammatory mediators is now considered to be a central feature in many cases of acute and chronic inflammation.

Lipid mediators derived from polyunsaturated fatty acids (PUFAs) have recently attracted much attention as critical regulators of inflammatory responses (Serhan, 2007). Eicosanoids, such as prostaglandins and leukotrienes derived from ω-6 arachidonic acid (AA; C20:4), are generally known as proinflammatory lipid mediators (Samuelsson et al., 1987). However, some AA metabolites such as 15-deoxy-Δ12,14-prostaglandin J₂ (15d-PGJ₂) and lipoxins have antiinflammatory capacities (Rossi et al., 2000; Straus et al., 2000). Furthermore, depending on tissue-specific contexts and receptor subtype occupancy, prostaglandin E₂ (PGE₂), and D₂ (PGD₂) often suppress the recruitment or activation of neutrophils, macrophages, or DCs (Gomez et al., 2005; Trivedi et al., 2006; Rajakariar et al., 2007; Honda et al., 2009). Recently, novel families of lipid mediators derived from ω-3 eicosapentaenoic acid (EPA; C20:5) and docosahexaenoic acid (DHA; C22:6), such as resolvins and protectins, have been shown to possess potent pro-resolving activities and thereby participate in the resolution of inflammation (Serhan et al., 2002; Mukherjee et al., 2004). PUFAs can also drive alternative antiinflammatory programs by acting on PUFA-sensing plasma membrane (e.g., GPR120) or nuclear (e.g., PPARγ) receptors (Diep et al., 2000; Oh et al., 2010), or by attenuating ER stress (Ariyama et al., 2010).

Bioynthesis of PUFA-derived lipid mediators is initiated by hydrolysis of phospholipids by phospholipase A₂ (PLA₂; Murakami et al., 2011). Of more than 30 members of the PLA₂ family, group IVA cytosolic PLA₂α (cPLA₂α; Bonventre et al., 1997; Uozumi et al., 1997) and, in certain cases, group VIA Ca²⁺-independent PLA₂ (iPLA₂; Moon et al., 2008) play major roles in stimulus-coupled AA release for eicosanoid generation. However, the molecular entity of PLA₂ enzymes that can trigger antiinflammatory lipid mediator programs still remains obscure. The secreted PLA₂ (sPLA₂) family, which consists of 11 isoforms, has long been thought to play proinflammatory roles in stimulus-coupled AA release for eicosanoid generation, as PLA2G2A protects from infection by degrading bacterial membranes (Wenrauch et al., 1996), PLA2G5 facilitates phagocytotic clearance of harmful materials by macrophages (Balestrieri et al., 2009; Boilard et al., 2010), and PLA2G10, when overexpressed in macrophages, can prime antiinflammatory, rather than proinflammatory, responses through production of PGE₂ and 15d-PGJ₂ (Curfs et al., 2008). Although sPLA₂s can liberate various fatty acids, including ω-3 PUFAs, in vitro, it remains unknown whether they could participate in some inflammation resolution processes by driving antiinflammatory lipid mediators in vivo.

Group IID sPLA₂ (PLA2G2D) was initially identified as an isoform structurally related to PLA2G2A and is abundantly expressed in lymphoid organs (Ishizaki et al., 1999; Shakhov et al., 2000; Valentin et al., 1999). Although PLA2G2D-transfected cells exhibited augmented AA release (Murakami et al., 2001) and infusion of an artificial PLA2G2D-Fc fusion protein into mice attenuated autoimmune diseases (von Allmen et al., 2009), its precise functions and mechanistic actions in vivo remain obscure. Herein, using mice lacking PLA2G2D (Pla2g2d<sup>−/−</sup>), we show that PLA2G2D is a lymphoid tissue-associated "resolving sPLA₂" that participates in the resolution of contact hypersensitivity (CHS). This is the first demonstration of a particular sPLA₂ subtype that lies upstream of the pro-resolving lipid mediators in a tissue-specific context.

RESULTS
PLA2G2D is preferentially expressed in lymphoid tissue-resident DCs and macrophages

We first aimed to determine the sites where PLA2G2D is intrinsically expressed. Previous studies have reported that PLA2G2D is expressed in secondary lymphoid organs (Ishizaki et al., 1999; Valentin et al., 1999; Shakhov et al., 2000). Consistently, quantitative RT-PCR for Pla2g2d in various tissues of C57BL/6 mice revealed its preferential expression in secondary lymphoid organs, including inguinal and mesenteric LNs, Peyer’s patch, and spleen (Fig. 1 A). Among the sPLA₂s, PLA2G2D is a major isoform expressed in cells from lymphoid organs (Fig. 1 B). In mouse spleen, cells with adhesive morphology and large cytoplasm in the red pulp and marginal zone, as well as focally in the white pulp, were stained with anti-PLA2G2D but not control antibody, whereas PLA2G2D staining was scarcely seen in lymphocytes (judging from round cell shape and small cytoplasm; Fig. 1 C).

three mice, a representative of three experiments). The separation procedure of individual cells is illustrated in Fig. S1. (E) Confocal immunofluorescence microscopy of PLA2G2D, CD11c, or F4/80 in spleen and LNs from Pla2g2d<sup>+/+</sup> and Pla2g2d<sup>−/−</sup> mice. (F) The AutoMACS-sorted splenic CD11c<sup>+</sup> cells were stained for MHC class II by FACS (right), and then analyzed for expression of Pla2g2d relative to Rn18s (mean ± SEM from four mice, a representative of two experiments; left). (G) Immunohistochemistry of human LNs with anti-human PLA2G2D or control antibody. A boxed area is magnified. Arrows indicate PLA2G2D<sup>+</sup> cells. (H) Confocal immunofluorescence microscopy of PLA2G2D and CD11c in human LNs. (I and J) AutoMACS-sorted CD11c<sup>+</sup> cells from mouse spleenocytes were further FACS-sorted for CD11b (I) or CD4 and CD8 (J; right), and then examined for expression of Pla2g2d relative to Rn18s (mean ± SEM from four mice, representative results from three experiments; left). (K) Expression of Pla2g2d relative to Rn18s in CD4<sup>+</sup>, CD8<sup>+</sup>, or CD4<sup>+</sup>CD25<sup>+</sup> T cells and CD11c<sup>+</sup> cells (a representative of two experiments). FACS profiles are representative of two (F) or three (I and J) experiments. Images are representative of two (H) or three (C, E, and G) experiments.
To accurately determine the cellular type(s) in which PLA2G2D is expressed, mouse splenocytes were sorted into CD11c+ and CD11c− cells, and the CD11c− population was further sorted for CD3ε (a T cell marker), CD45R (a B cell marker), and CD11b (a macrophage marker; Fig. S1). Quantitative RT-PCR revealed that Pla2g2d was enriched in the CD11c+ fraction (Fig. 1 D), suggesting that PLA2G2D is expressed in lymphoid tissue-resident CD11c+ cells of the mononuclear phagocyte lineage, including DCs and a subset of macrophages. In comparison, Pla2g5, which was detected in mouse spleen at a lower level, was distributed in CD11b+CD11c− macrophages, whereas Pla2g1b, Pla2g2a (absent in C57BL/6 mice because of a natural mutation), Pla2g2e, Pla2g2f, and Pla2g10 were hardly detected in these cells (Fig. 1, B and D). When mouse spleen and LNs were subjected to confocal immunofluorescence microscopy, PLA2G2D was largely colocalized with CD11c and only partially with F4/80, a genuine macrophage marker (Fig. 1 E), indicating that it is expressed in CD11c+ DCs and a subpopulation of CD11c+ F4/80+ macrophages. Further sorting of the AutoMACS-sorted CD11c+ cells revealed the distribution of Pla2g2d in MHC class II+ cells in preference to MHC class II− cells (Fig. 1 F), suggesting that PLA2G2D is mainly expressed in DCs of low activation state and is down-regulated in DCs undergoing activation (also see below). Staining of PLA2G2D was also found in human LNs, where it was distributed in cells within the paracortical area and was colocalized with CD11c (Fig. 1 G, H).

In mouse spleen, lymphoid-resident conventional DCs are subdivided into three major subsets, including CD11b+CD4+CD8α− DCs (CD4+ DCs), CD11b+CD8α− DCs (CD8α− DCs), and CD11b−CD4−CD8α− DCs (CD4−CD8α− DCs; Vremec et al., 2000; Tamura et al., 2005). When the AutoMACS-sorted CD11c+ cells were further FACS-sorted for CD11b, Pla2g2d mRNA was enriched in the CD11b+ population (Fig. 1 I). FACS sorting of another pool of CD11c+ cells for CD4 and CD8α showed more abundant expression of Pla2g2d in CD4+ DCs than in CD8α− or CD4−CD8α− DCs (Fig. 1 J). Thus, among the conventional DCs, PLA2G2D is preferentially expressed in the CD11b+CD4+CD8α− DC subset. Because PLA2G2D is expressed in T reg cells (von Allmen et al., 2009), we compared the expression of Pla2g2d in CD11c+ cells and CD4+CD25− T reg cells in mouse spleen. Among T cells, Pla2g2d was enriched in CD4+CD25− T reg cells relative to total CD4+ and CD8α− T cells, as previously reported (von Allmen et al., 2009). However, its expression was markedly higher in CD11c+ cells than in T reg cells (Fig. 1 K).

Generation of Pla2g2d−/− mice

To investigate the roles of PLA2G2D in vivo, we generated Pla2g2d−/− mice by gene targeting. Construction of the Pla2g2d targeting vector is illustrated in Fig. 2 A. Appropriately targeted ES cell clones were used to obtain chimeric mice that transmitted the targeted locus through the germ line. Male mice carrying a targeted Pla2g2d gene were crossed with female CAG-Cre transgenic mice, which allowed deletion of exons 2 and 3 from the Pla2g2d gene in all tissues. Heterozygous mice carrying a mutated Pla2g2d allele (Pla2g2d+/−) were backcrossed onto the C57BL/6 background (n > 12). Successful ablation of the Pla2g2d gene was confirmed by PCR genotyping from tail biopsy (Fig. 2 B) and by the absence of its mRNA (Fig. 2 C) and protein (Fig. 2 D) in the LNs and spleen of Pla2g2d−/− mice as demonstrated by quantitative RT-PCR and immunoblotting, respectively. Also, PLA2G2D staining was absent in the LNs of Pla2g2d−/− mice (Fig. 1 E), confirming the specificity of the anti-PLA2G2D antibody used here.

The ratio of the genotypes of heterozygous male and female offspring exhibited Mendelian proportions, and homozygous null mice were indistinguishable from WT littermates in terms of survival rates, appearance, fertility, and gross behavior, suggesting that the lack of PLA2G2D did not affect embryonic and postnatal development in normal states. The serum levels of biomedical markers and the proportions of blood cells were similar between the genotypes (Tables S1 and S2). DC, T cell, B cell, and macrophage populations in spleen, inguinal, and mesenteric LNs, Peyer’s patch, and thymus were indistinguishable between Pla2g2d−/− and Pla2g2d+/− mice, except that the splenic proportion of CD4+ T cells was slightly greater in Pla2g2d−/− mice (Fig. 2 E–G). Thus, PLA2G2D ablation did not profoundly affect hematopoiesis or lymphopoiesis under normal conditions.

Impaired resolution of hapten-induced CHS in Pla2g2d−/− mice

To assess the immunoregulatory role of PLA2G2D in vivo, Pla2g2d−/− mice and WT littermates were subjected to hapten-induced CHS. In this well-known model for Th1 response, application of the hapten antigen dinitrofluorobenzene (DNFB) onto abdominal skin (sensitization), followed by the second application of the same antigen onto ear skin (elicitation) induced ear swelling. In normal skin, there are resident CD11c+ populations including epidermal Langerhans cells and dermal DCs, which capture antigen and migrate into draining LNs (Heath and Carbone, 2009). Although Pla2g2d expression was low in nonlymphoid tissues (Fig. 1 A), confocal microscopy of the ear revealed that scattered PLA2G2D signal was colocalized in dermal and to a lesser extent in epidermal CD11c+ cells in WT mice, while it was absent in Pla2g2d−/− skin (Fig. 3 A). Inflammation recruits monocytes that are differentiated into CD11c+ inflammatory DCs or macrophages, which act as a peripheral stimulator of T cells (Heath and Carbone, 2009). Remarkably, cells expressing both PLA2G2D and CD11c were increased in the dermis and even epidermis after DNFB challenge for 24 h (Fig. 3 A), indicating that PLA2G2D is also expressed in many, if not all, inflammatory DCs or macrophages migrating into the inflamed skin.
We then compared the process of CHS between Pla2g2d<sup>+/+</sup> and Pla2g2d<sup>−/−</sup> mice. In WT mice, ear swelling peaked by day 3 and declined to nearly the basal level by day 6 after DNFB challenge (Fig. 3 B). At lower doses of DNFB, milder edema peaked on days 2–3, and then shifted to the declining phase (Fig. 3 C), confirming that the onset and resolution were not affected by the intensity of the stimulus. The initial process of ear swelling by day 3 occurred normally in Pla2g2d<sup>−/−</sup> mice, with no significant difference from Pla2g2d<sup>+/+</sup> mice. After reaching the peak, ear edema in Pla2g2d<sup>−/−</sup> mice continued to maintain the maximally exaggerated state by day 5, and then decayed at a slower rate by day 10, at which time the ear was still thicker than in WT mice (Fig. 3 B), implying a delayed resolution in the null mice. In accordance, histological
Figure 3. Impaired resolution of skin inflammation in Pla2g2d<sup>−/−</sup> mice in a DNFB-induced CHS model. (A) Confocal immunofluorescence microscopy of PLA2G2D (IID-Alexa647; red) and CD11c (CD11c-PE; green) in ear skin from Pla2g2d<sup>+/+</sup> (+/+) and Pla2g2d<sup>−/−</sup> (−/−) mice before and 24 h after elicitation of DNFB-induced CHS. Green signal in the cartilage is nonspecific. Images are representative of two experiments. (B) Monitoring of ear thickness over the elicitation period of CHS in Pla2g2d<sup>+/+</sup> and Pla2g2d<sup>−/−</sup> mice (left). Result is a representative of three experiments (mean ± SEM from 7
examination of the ears showed more severe ear swelling, epidermal hyperplasia, and inflammatory cell infiltration in Pla2g2d−/− mice than in Pla2g2d+/− mice on day 5 (Fig. 3 D).

Quantitative RT-PCR of ear skin revealed that the expression levels of inflammatory markers, including Cd11c, Cd11b, and Cd68 (surface markers of inflammatory DCs or macrophages), Ifng (IFN-γ; a Th1 cytokine), Il1b, Il6, and Tnf (IL-1β, IL-6, and TNF; proinflammatory cytokines), Cd2, and Ccr2 (a monocyte-attracting chemokine and its receptor), and S100a9 (a marker of epidermal hyperplasia) peaked by day 3, and then declined by day 5 in Pla2g2d+/− mice, whereas they were dramatically increased on day 5 in Pla2g2d−/− mice (Fig. 3 E). A robust increase of Ifng in Pla2g2d−/− mice over Pla2g2d+/− mice became obvious on day 3 before that of other proinflammatory markers, implying that the exacerbated inflammation in the null mice stemmed from the augmented Th1 response (Lu et al., 1998). In contrast, expression levels of Il10 (IL-10), an antiinflammatory cytokine (Fickenscher et al., 2002), and of Foxp3, a marker of T reg cells (Shevach, 2009), in Pla2g2d−/− skin did not differ significantly from those in Pla2g2d+/− skin throughout the experimental periods (Fig. 3 E), suggesting that T reg cell migration or proliferation in the inflamed skin was unaffected by PLA2G2D deficiency. Thus, during the resolution process of CHS, a cytokine milieu in the affected skin is skewed toward a proinflammatory profile rather than an antiinflammatory profile in Pla2g2d−/− mice.

We also examined the inflammatory status in the draining LNs during the elicitation phase of CHS. After DNFB challenge to the ears, Pla2g2d expression in the LNs of WT mice was increased on days 1 to 5 (Fig. 4 A), reaching a much higher level than in the skin (Fig. 4 B). The modest increase of Pla2g2d was accompanied by a concomitant increase of Cd11c and a robust increase of Ifng on days 1–3 (Fig. 4 A). Subsequently, the expressions of Cd11c and Ifng were declined to their basal levels by day 5 in Pla2g2d+/− mice, underscoring the resolution of LN inflammation. In Pla2g2d−/− LNs, however, the expression levels of Cd11c and Ifng still remained at their highest levels on day 5 (Fig. 4 A). On day 5, Cd11c+MHC class II+ activated DCs (Fig. 4 C) and CD3e+ CD44+CD62L− effector T cells (Fig. 4 D) were greater in Pla2g2d−/− LNs than in Pla2g2d+/− LNs. Thus, as in the affected skin, inflammation failed to resolve in the draining LNs of DNFB-challenged Pla2g2d−/− mice relative to Pla2g2d+/− mice.

LN cells were harvested from DNFB-sensitized Pla2g2d−/− and Pla2g2d+/− mice and cultured for 24 h ex vivo in the presence of dinitrobenzene sulfonic acid (DNBS; a water-soluble form of DNFB). Expression levels of the Th1 cytokines Ifng and Il12a were markedly higher in Pla2g2d−/− LN cells than in WT cells, whereas those of the antiinflammatory cytokines Tgfβ and Il10, as well as the T reg cell marker Foxp3, were unaffected by the absence of PLA2G2D (Fig. 4 E). In agreement, the elevated secretion of IFN-γ, but not IL-10, in the culture of Pla2g2d−/− LN cells relative to Pla2g2d+/+ LN cells over time was confirmed by ELISA (Fig. 4 F), and the unaltered Cd4+Foxp3+ T reg cell population in Pla2g2d−/− LNs was confirmed by flow cytometry (Fig. 4 G). These results indicate again that the proinflammatory signal exceeds the antiinflammatory signal in Pla2g2d−/− LNs compared with Pla2g2d+/+ LNs. However, our results do not support the proposal that PLA2G2D facilitates the expansion and immunosuppressive function of T reg cells (von Allmen et al., 2009), at least in the CHS model used here.

To verify whether PLA2G2D deficiency did not affect the initial process of CHS, we examined the migration of skin-resident DCs into the draining LNs during the sensitization phase. After capture of a foreign antigen, skin-resident DCs are activated and migrate into the regional LNs, where they present the antigen to T cells (Heath and Carbone, 2009). The fluorescent hapten antigen FITC was topically applied onto the skin of Pla2g2d−/− and Pla2g2d+/− mice, and then the migration of FITC-captured skin DCs into the draining LNs was monitored by flow cytometry. The proportion of FITC+CD11c+ or FITC+MHC class II+ DCs was similar between Pla2g2d−/− and Pla2g2d+/− LNs (Fig. 4, H and I), implying that the lack of PLA2G2D does not affect antigen uptake, recruitment, and activation of skin DCs migrating into the LNs. Notably, many of the FITC+CD11c+ cells in the LNs did not display PLA2G2D signal (Fig. 4 J), suggesting that PLA2G2D expression was decreased after activation and LN recruitment of skin DCs. Thus, the apparent effect of PLA2G2D deficiency is restricted to the late period of CHS.

Pla2g2d−/− DCs are hyperactivated

To assess whether the increased immune responses in Pla2g2d−/− mice is attributable to some abnormalities in DCs, we analyzed GM-CSF–driven BM-derived DCs (CD11b+CD11c+ BMDCs; McGeeachy, 2011). Although the expression of Pla2g2d was minimal in BM cells, it was robustly induced in GM-CSF-driven BMDCs (Fig. 5 A). Compared with BMDCs, Pla2g2d expression in M-CSF–driven BM-derived macrophages (CD11b+ CD11c− BMDMs) was low (Fig. 5 B), agreeing with the preferential expression of PLA2G2D in CD11c+ DCs in vivo. When BMDCs or even BMDMs were treated for 24 h with LPS, Pla2g2d expression was reduced markedly (Fig. 5, A and B).
Figure 4. Impaired resolution of LN inflammation in Pla2g2d<sup>-/-</sup> mice in a DNFB-induced CHS model. (A) Quantitative RT-PCR of Pla2g2d, Cd11c, and Ifng relative to Gapdh in the LNs of Pla2g2d<sup>+/+</sup> (+/+) and Pla2g2d<sup>-/-</sup> (−/−) mice during the elicitation phase of CHS (mean ± SEM from four mice per group). (B) Quantitative RT-PCR of Pla2g2d relative to Gapdh in the ear skin and LNs of Pla2g2d<sup>+/+</sup> and Pla2g2d<sup>-/-</sup> mice on day 5 with or without DNFB challenge (mean ± SEM from four mice per group). (C and D) FACS for activated DCs (CD11c<sup>+</sup>MHC class II<sup>+</sup>; C) and effector T cells (CD3ε<sup>+</sup>) on April 6, 2017 Downloaded from Published May 20, 2013
a finding reminiscent of its down-regulation by LPS in mouse tissues (Suzuki et al., 2000; von Allmen et al., 2009) and human macrophages (Lindbom et al., 2005). Thus, corroborating with the data shown in Figs. 1 F and 4 J, PLA2G2D expression inversely correlates with DC activation.

Even without stimulation, Pla2g2d+/− BMDCs had greater surface expression of MHC class II, CD80, and CD86 than Pla2g2d+/+ BMDCs (Fig. 5 C). Further incubation with a suboptimal concentration of LPS increased more CD11b+ MHC class II+ (Fig. 5 D) and CD80+CD86+ BMDCs from Pla2g2d−/− mice than from Pla2g2d+/+ mice. There were substantial increases in IL-6 and TNF secretion from Pla2g2d−/− BMDCs over Pla2g2d+/+ BMDCs in response to suboptimal LPS (Fig. 5 F). Moreover, when C57BL/6 mice were intravenously transferred with Pla2g2d+/+ or Pla2g2d−/− BMDCs that had been sensitized with DNBS, and then subjected to DNFB challenge, mice transferred with Pla2g2d−/− BMDCs exhibited greater edema than those transferred with Pla2g2d+/+ BMDCs (Fig. 5 G). Thus, PLA2G2D deficiency allows BMDCs to be hyperactivated, leading to more severe inflammation.

**Altered lipid profiles in the LNs of Pla2g2d−/− mice**

Because PLA2G2D is a lipolytic enzyme, we speculated that the impaired resolution of CHS in Pla2g2d−/− mice might result from some perturbed lipid metabolisms. We therefore performed electrospray ionization mass spectrometry (ESI-MS) to comprehensively monitor the levels of free PUFAs and their oxygenated metabolites in the draining LNs and skin of Pla2g2d−/− mice in comparison with Pla2g2d+/+ mice on day 5 after DNFB challenge, a time point when Pla2g2d−/− mice exhibited an impaired resolution of inflammation.

Bioactive metabolomes for AA-, EPA-, and DHA-derived products revealed that DNFB challenge led to substantial increases in free AA, EPA, and DHA, with concomitant increases in many if not all of their bioactive metabolites, in the LNs of Pla2g2d−/− mice. Remarkably, the levels of AA, EPA, and DHA were significantly lower in Pla2g2d−/− LNs than in Pla2g2d+/+ LNs in the presence or even in the absence of DNFB challenge (Fig. 6 A), suggesting that PLA2G2D plays a role in the constitutive and inducible supply of a large portion of these PUFAs in the LNs. However, the levels of many AA metabolites, including the cyclooxygenase products PGE2, PGF2α, 6-keto-PGF1α (an end-product of PG1α), and thromboxane (TX) B2 (an end-product of TXA2) as well as the 5-lipoxygenase product 5-hydroxyeicosatetraenoic acid (HETE), some of which promote Th1-dominated inflammation (Byrum et al., 1999; Kabashima et al., 2003; Nagamachi et al., 2007; Yao et al., 2009; Nakajima et al., 2010), did not differ significantly between the genotypes in DNFB-treated and -untreated settings (Fig. 6 B). In contrast, the DNFB-induced increases in the AA-derived antiinflammatory prostanoid 15d-PGJ2 and its precursor PGD2 occurred only partially in Pla2g2d−/− mice relative to Pla2g2d+/+ mice (Fig. 6 C). These results suggest that the PLA2G2D-sensitive AA pool is preferentially coupled with the antiinflammatory PGD2/15d-PGJ2 axis, whereas another AA pool remaining in Pla2g2d−/− mice, which may be supplied by cPLA2α (Bonventre et al., 1997; Uozumi et al., 1997), is sufficient for full mobilization of the proinflammatory AA metabolites.

In addition, the EPA and DHA bioactive metabolomes revealed that the levels of ω-3 EPA- and DHA-derived metabolites, which included the potent pro-resolving lipid mediator resolvins D1 (RvD1), were considerably lower in Pla2g2d−/− LNs than in Pla2g2d+/+ LNs regardless of DNFB challenge (Fig. 6 D). Thus, along with the reduction in the inducible production of ω-6 AA-derived 15d-PGJ2 (Fig. 6 C), the absence of PLA2G2D causes a marked reduction in the steady-state levels of ω-3 PUFAs and their enzymatically oxygenated products. The lipidomic profile on day 1, an early peak point of CHS, was similar to that on day 5 (Fig. 6 E). Hence, the constitutive supply of pro-resolving lipid mediators by PLA2G2D may be important for the early entry into resolution, rather than for limiting the maximal inflammatory response. To obtain a possible explanation for these changes in lipid profiles in Pla2g2d−/− LNs, we examined the expression of downstream enzymes responsible for the conversion of PUFAs to bioactive metabolites. Notably, the expression levels of prostanooid-biosynthetic enzymes tended to be higher in Pla2g2d−/− LNs than in Pla2g2d+/+ LNs, whereas that of 12/15-lipoxygenase, an enzyme crucial for the conversion of ω-3 PUFAs to antiinflammatory lipid mediators, was similar in both genotypes (Fig. 6 F). Thus, in Pla2g2d−/− LNs, the diminished supply of AA might be counter-balanced by the elevated expression of downstream enzymes for proinflammatory lipid mediators, while the reduction of ω-3 PUFAs might be linked to the reduction of pro-resolving lipid mediators because of the lack of compensatory up-regulation of 12/15-lipoxygenase.

Lipidomics analysis of the affected skin on day 5 of CHS revealed that none of the free PUFAs and their metabolites were significantly altered in Pla2g2d−/− mice relative to Pla2g2d+/+.
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We found that PLA2G2D selectively decreased several diacyl and alkenyl forms of phosphatidylethanolamine (PE) molecular species containing unsaturated fatty acids, particularly those containing AA and DHA, whereas phosphatidylcholine (PC), phosphatidylinositol (PI), phosphatidylserine (PS), and phosphatidylglycerol (PG) species were barely affected by the enzyme (Fig. 7 A). The decreases of particular PE species by PLA2G2D were accompanied by concomitant increases in lysophosphatidylethanolamine (LysoPE) species with saturated or monounsaturated fatty acid (which are likely to be sn-1 bound), supporting the PLA2 reaction (Fig. 7 B). In agreement with the unaltered PC and PS levels without recombinant mouse PLA2G2D protein. Then, the lipids were subjected to ESI-MS to analyze the reduction of phospholipid species and the increase of lysophospholipids and free fatty acids.

mice (Fig. 6 G). Thus, despite the increased skin inflammation, overall lipid mediator profiles in the skin were unaffected by PLA2G2D deficiency. Altogether, it is likely that PLA2G2D acts mainly in lymphoid tissues, where it supplies a particular pool of PUFAs that is selectively coupled with antiinflammatory/pro-resolving lipid mediator programs and thereby contributes to the resolution of CHS primarily in the LNs and secondarily in the skin.

PLA2G2D releases PUFAs from phosphatidylethanolamine in LN membranes

Having established that the deficiency of PLA2G2D reduces the LN levels of PUFAs such as AA and DHA, we next sought to gain insights into phospholipid subclasses from which PLA2G2D could release these PUFAs. To this end, we extracted total lipids from LNs and incubated them with or without recombinant mouse PLA2G2D protein. Then, the lipids were subjected to ESI-MS to analyze the reduction of phospholipid species and the increase of lysophospholipids and free fatty acids.

We found that PLA2G2D selectively decreased several diacyl and alkenyl forms of phosphatidylethanolamine (PE) molecular species containing unsaturated fatty acids, particularly those containing AA and DHA, whereas phosphatidylcholine (PC), phosphatidylinositol (PI), phosphatidyserine (PS), and phosphatidylglycerol (PG) species were barely affected by the enzyme (Fig. 7 A). The decreases of particular PE species by PLA2G2D were accompanied by concomitant increases in lysophosphatidylethanolamine (LysoPE) species with saturated or monounsaturated fatty acid (which are likely to be sn-1 bound), supporting the PLA2 reaction (Fig. 7 B). In agreement with the unaltered PC and PS levels
Figure 6. LC-ESI/MS profiling of PUFAs and their bioactive mediators. (A–D) LC-ESI/MS of free PUFAs (A), proinflammatory AA metabolites (B), antiinflammatory AA metabolites (C), and EPA or DHA metabolites (D) in the LNs of Pla2g2d+/+(+/+) and Pla2g2d−/− (−/−) mice on day 5 of CHS, with or without DNFB treatment (mean ± SEM from five mice per group). Values are AUC (area under the curve) per mg tissue, as evaluated by comparison with an internal standard (1 AUC = ~50 pg). (E) LC-ESI/MS of free PUFAs and their metabolites in the LNs of Pla2g2d+/+ and Pla2g2d−/− mice on day 1 after DNFB challenge (mean ± SEM from three mice per group). (F) Quantitative RT-PCR of lipid mediator-biosynthetic enzymes relative to Gapdh in the LNs on day 5 after DNFB challenge (mean ± SEM from three mice per group). (G) LC-ESI/MS of free PUFAs and their metabolites in the ear skin of DNFB-challenged Pla2g2d+/+ and Pla2g2d−/− mice on day 5 (mean ± SEM from 10 mice per group). In E and G, values are expressed as AUC ratio, with the level of each product in Pla2g2d+/+ mice being regarded as 1. HEPE, hydroxyeicosapentaenoic acid; HDHA, hydroxydocosahexaenoic acid; PD1, protectin D1. Representative results of two (E and F) or three (A–D) experiments and pooled results of two experiments (G) are shown. *, P < 0.05; **, P < 0.01.
(Fig. 7 A), there were no increases in lysophosphatidylcholine (LysoPC) and lysophosphatidylserine (LysoPS) species (Fig. 7 B). Among the PUFAs, DHA and AA were increased in marked preference to linoleic acid (LA; C18:2) after the treatment with PLA2G2D (Fig. 7 C), an observation that agreed with the selective reduction of PE species bearing DHA or AA (Fig. 7 A). Moreover, this PUFA preference in vitro was recapitulated by marked abundance of free DHA and AA relative to LA in the LNs in vivo (Fig. 7 D). These results collectively suggest that PLA2G2D selectively hydrolyzes PUFA-containing PE in LN membranes to provide DHA and AA.

Figure 7. Hydrolysis of LN-derived phospholipids by recombinant PLA2G2D. (A–C) ESI-MS of phospholipids (A), lysophospholipids (B), and fatty acids (C) in total lipids extracted from the LNs and incubated with (+) or without (−) 200 nM recombinant mouse PLA2G2D (mean ± SEM, n = 3 per group). (D) ESI-MS of LA, AA, and DHA in the LNs of WT mice (mean ± SEM from five mice). Results in A–D are representative of three independent experiments. *, P < 0.05; **, P < 0.01.
The lipid products altered in Pla2g2d<sup>−/−</sup> LNs dampen the DC-mediated immune responses

Finally, to assess whether the lipid products altered in Pla2g2d<sup>−/−</sup> LNs could indeed modulate the immune response, LN cells from DNFB-sensitized mice were stimulated with DNBS in the presence of the lipids. EPA, DHA, 15d-PGJ<sub>2</sub> and RvD1 dampened IFN-γ secretion, whereas LA and AA were ineffective (Fig. 8A), confirming the immune regulation by the ω-3 PUFA products and the ω-6 cyclopentenone prostanooid. These antiinflammatory lipids also efficiently blocked IFN-γ production by Pla2g2d<sup>−/−</sup> LN cells (unpublished data). The levels of these lipids in WT LNs were estimated to be within a range of nanomolar to micromolar orders (Fig. 6A), which were sufficient to block IFN-γ production by LN cells (Fig. 8A). Furthermore, DHA, 15d-PGJ<sub>2</sub> and RvD1 reduced IL-12 secretion (Fig. 8B) and surface expression of MHC class II (Fig. 8C, D) in LPS-stimulated BMDCs. Together with previous studies reporting the antiinflammatory effects of ω-3 PUFA-rich fish oils in various diseases (Yokoyama et al., 2003; Serbina et al., 2003; Allenspach et al., 2008), our present results indicate that the lipid mediator balance is shifted toward a proinflammatory status in favor of an antiinflammatory status in Pla2g2d<sup>−/−</sup> LNs during the resolution phase of CHS.

**DISCUSSION**

Although the last decade has seen progress in deciphering the pathways that activates innate and acquired immunity, the mechanisms that limit these responses to secure homeostasis have not yet been fully defined. Antiinflammatory or pro-resolving lipid mediators can explain some of the beneficial actions of ω-3 PUFA-rich fish oils in various diseases (Yokoyama et al., 2007). Although the understanding of PUFA-oxygenated products with pro-resolving activities has recently been expanding (Serhan, 2007; Serhan et al., 2008), an interest in exploring the role of PLA<sub>2</sub> that can supply PUFA precursors upstream of their biosynthetic conversion to pro-resolving lipid mediators has emerged. Here, we have shown that PLA2G2D is an immunosuppressive sPLA<sub>2</sub> that is abundantly expressed in CD11c<sup>+</sup> cells in lymphoid organs and that its absence delays the resolution of CHS. Importantly, pro-resolving lipid mediators and their precursors were reduced in Pla2g2d<sup>−/−</sup> LNs, implying that PLA2G2D resolves inflammation by driving antiinflammatory lipid mediators in this tissue. Because of its pro-resolving role in CHS, along with a recent study showing an immunosuppressive effect of PLA2G2D-Fc fusion protein (von Allmen et al., 2009), we propose that PLA2G2D represents a “resolving sPLA<sub>2</sub>,” that mobilizes antiinflammatory or pro-resolving lipid mediators in lymphoid tissues.

PLA2G2D is mainly located in a cell population expressing CD11c, a molecular signature of DCs. DCs are antigen-presenting cells specialized to capture and process antigens for presentation on MHC products and then to control differentiation of T cells (Villadangos and Schnorrer, 2007), and they are highly diverged in terms of their locations, surface marker expressions, and functions (Heath and Carbone, 2009). Lymphoid-resident DCs present antigen that drains to LNs and stimulate the initial steps of T cell activation; migratory DCs transport antigen from peripheral tissues to regional LNs and stimulate the next steps of T cell activation; as immune stimulation continues, blood monocytes recruited to peripheral sites develop into inflammatory DCs, which migrate into LNs to provide a continuing stimulus for T cell activation; and TipDCs, another unique DC subset, mediate T cell–independent innate immune defense (Itano et al., 2003; Serbina et al., 2003; Allenspach et al., 2008;
target phospholipids of PLA2G2D are particular PE molecular pro- versus antiinflammatory lipid mediators. The preferred explanation for the different coupling of PLA2G2D with calized in distinct cell populations, could provide partly an downstream biosynthetic enzymes, each of which may be lo-

ular basis for the distinct compartmentalization of the pro-

plored compartmentalization of distinct PUFA pools toward

AA-derived mediators appear to be mostly derived from the

PLA2G2D participates in the steady-state supply of a particu-

mice even under noninflamed conditions, suggesting that

LN levels of free AA, EPA, and DHA were lower in

activation of CD4+ DCs.

mice over WT mice in the

Haan et al., 2000; Pooley et al., 2001). PLA2G2D is prefer-

with the antiinflammatory property of this enzyme. In addi-

$a$ sets, CD8$^+ $ DCs are responsible for cell-mediated immune

responses, whereas CD4$^+$ DCs regulate humoral responses

(Maldonado–López et al., 1999; Pulendran et al., 1999; den

Haan et al., 2000; Pooley et al., 2001). PLA2G2D is prefer-

entially expressed in lymphoid tissue-resident CD4$^+ $CD11b$^+$

DCs as well as in dermal DCs, monocyte-derived inflam-

matory DCs/macrophages, and ex vivo-derived, GM-CSF-

driven BMDCs.

Although PLA2G2D is largely dispensable for normal de-

velopment of DCs as well as lymphocytes and other myeloid

cells, its deficiency impairs resolution, rather than progression,

of CHS with over-production of proinflammatory markers,

while holding antiinflammatory markers unchanged, in both

skin and regional LNs. Robust elevation of DC/macrophage

surface markers and proinflammatory cytokines/chemokines

in hapten-challenged Pla2g2d$^{-/-}$ mice over WT mice in the late phase of CHS indicates that PLA2G2D deficiency per-

turs the termination program for the recruitment and activa-

tion of monocyte-derived inflammatory DCs/macrophages.

In support of this, GM-CSF-driven BMDCs from Pla2g2d$^{-/-}$
mice have elevated expression of cell activation markers and can elicit skin inflammation after transferred into mice. The decrease of PLA2G2D expression in activated DCs also accords with the antiinflammatory property of this enzyme. In addition, a modestly increased splenic CD4$^+$ T cells in Pla2g2d$^{-/-}$ mice under normal condition might result from the increased activation of CD4$^+$ DCs.

During the process of inflammation, lipid mediator pro-

files switch from proinflammatory to antiinflammatory products, which in turn contribute to sequestration of the recruitment of inflammatory cells to the site (Levy et al., 2001; Serhan et al., 2008). Of importance, lipidomics profiling has delineated a criti-
cal role of PLA2G2D in the supply of a pool of free PUFAsthat can be selectively linked to an array of pro-resolving lipid products in the skin-draining LNs. Thus, several DHA- and EPA-oxygenated products and AA-derived 15d-PGJ$_2$ were significantly reduced in the LNs by PLA2G2D deletion. The LN levels of free AA, EPA, and DHA were lower in Pla2g2d$^{-/-}$ mice even under noninflamed conditions, suggesting that PLA2G2D participates in the steady-state supply of a particu-

lar PUFA pool within the LNs. In contrast, Th1-promoting AA-derived mediators appear to be mostly derived from the PLA2G2D-independent AA pool, thus revealing an unex-

plored compartmentalization of distinct PUFA pools toward pro-

versus antiinflammatory pathways. Although the molecu-

lar basis for the distinct compartmentalization of the pro-

and antiinflammatory lipid pathways in the LNs is unclear at present, it may rely on when and where PLA2G2D secreted from DCs could encounter its target phospholipids during the process of immune response. The differential expression of downstream biosynthetic enzymes, each of which may be lo-

lized in distinct cell populations, could provide partly an explanation for the different coupling of PLA2G2D with pro-

versus antiinflammatory lipid mediators. The preferred target phospholipids of PLA2G2D are particular PE molecular species with PUFAs such as DHA and AA. Considering that PE usually resides in the inner leaflet of the plasma membrane in resting cells, PLA2G2D might act on these PE species exposed on activated cells by a process of flip-flop or those released extracellularly from the cells (e.g., exosomes or microvesicles). Supporting the latter possibility, sPLA$_2$ acts on microvesicles shed from cells at inflamed sites (Fourcade et al., 1995) and leukocyte-derived microvesicles, which are temporally generated in inflammatory exudates during resolution, contain esterified biosynthetic precursors of pro-resolving mediators (Norling et al., 2011).

Collectively, we conclude that PLA2G2D is preferen-
tially coupled with the biosynthesis of pro-resolving lipid mediators that could resolve LN inflammation. Our results are compatible with a recent report showing that mouse lymphoid tissues produce RvD1, which can regulate antibody production (Ramon et al., 2012). Also, free PUFAsthe-

m selves might be protective against inflammation through act-

ing on PUFA-sensing receptors (Diep et al., 2000; Oh et al.,

2010) or through reducing the stress response (Ariyama et al.,

2010). As such, the lack of PLA2G2D, and accordingly the decrease of the pro-resolving lipid mediators driven by this enzyme, may tip the balance toward a more aggressive in-

flammatory response in the LNs and thereby in the skin. The lipid-mobilizing role of PLA2G2D in local skin microenvi-

roments remains to be elucidated.

PLA2G5 is expressed in macrophages and DCs (Giannattasio

et al., 2010; Satake et al., 2004), and we have shown here that it is preferentially localized in CD11b$^+ $CD11c$^-$ splenic macro-

phages. The expression of PLA2G2D in lymphoid organs and in CD11c$^-$ cells is much higher than that of PLA2G5 and other sPLA$_2$ isoforms. Beyond their distinct expression pat-

terns, these two sPLA$_2$s afford opposite effects on DC func-

tions and immunity; PLA2G5 facilitates antigen capture and presen-

tation by DCs toward airway inflammation (Giannattasio et al.,

2010), while PLA2G2D has pro-resolving roles in CHS (as shown in this study by gene targeting) and in experimental multiple sclerosis and colitis (as shown by injection of PLA2G2D–Fc fusion protein; von Allmen et al., 2009). Although the mech-

anism underlying the functional difference between these two sPLA$_2$s is unclear, it might rely at least partly on their distinct expression levels and on their different substrate selectivity in that PLA2G2D prefers PE bearing PUFAsthis study), whereas PLA2G5 prefers PE and PC bearing fatty acids with low degree of unsaturation (Chen and Dennis, 1998; Mitsuishi et al., 2007). Non-redundant roles of multiple sPLA$_2$s in a given process have also been documented for arthritis (offensive PLA2G2A versus defensive PLA2G5; Boilard et al., 2010) and for male reproduction (PLA2G3 for sperm maturation versus PLA2G10 for sperm activation; Escoffier et al., 2010; Sato et al., 2010). The contrasting roles of PLA2G2D versus PLA2G5 in the DC-related immunity represent another example of these occasions. Thus, individual sPLA$_2$s, which exhibit distinct substrate preferences or cellular locations in a given tissue or cell type, act spatiotemporally on different membrane targets, thereby linking to distinct biological outcomes. Although pan-sPLA$_2$
inhibitors, which block classical sPLA₂s broadly, attenuate several disease models (Rosenson et al., 2009), dampening offensive and defensive sPLA₂s altogether might limit their therapeutic use (Bowton et al., 2005; Bradley et al., 2005). Our results point to an alternative strategy to use PLA₂G2D, a resolving sPLA₂, for treatment of human disorders.

MATERIALS AND METHODS

Targeted disruption of the Pla2g2d gene. The Pla2g2d-targeting vector was constructed with a neomycin resistance (Neo) gene that was inserted between exons 1 and 2 of the Pla2g2d gene. ES cell transfection and embryo injections were performed by the Transgenic Resources Program (Department of Comparative Medicine, University of Washington, Seattle, WA). ES clones with homologous recombination were screened by PCR and Southern blotting. After germline transmission, Pla2g2d<sup>−/−</sup> mice were bred with C57BL/6NCrSlc (Japan SLIC) mice (n > 12). Genotyping was performed on genomic DNA from tail biopsies by PCR using the primer pairs 5′-CCATCTGAACTTGAGTGGGAAGTGCCA-3′ and 5′-GGCTATGGTGCCCTCCTATGAG-3′ (Gene SQnosys), which amplified a 490-bp fragment specific for the WT allele, and 5′-TGTCGTCAAGAAGCATGTGTTG-3′ and 5′-GGCTATGGTGCCCTCCTATGAG-3′, which amplified a 530-bp fragment specific for the mutant allele. The reaction was 95°C for 10 s and then 35 cycles of 95°C for 0 s and 65°C for 1 min on Applied Biosystems 9800 Fast Thermal Cycler (Applied Biosystems). The PCR products were analyzed by 1.5% (wt/vol) agarose gel electrophoresis with ethidium bromide. All mice were housed in climate-controlled (23°C) specific pathogen–free facilities with a 12-h light-dark cycle, with free access to standard laboratory food (CE2; Laboratory Diet; CLEA Japan) and water. All procedures involving animals were performed in accordance with protocols approved by the Institutional Animal Care and Use Committees of the Tokyo Metropolitan Institute of Medical Science, Showa University, and the University of Washington.

Hapten-induced CHS. On day 5, mice (7–12 wk-old, male) were sensitized with 50 µl of 0.8% (vol/vol) DNFB in acetone/olive oil (4/1, vol/vol) on shaved abdominal skin. On day 0, ears were challenged by 20 µl of 0.3% DNFB to monitor ear swelling. Ear thickness was measured for each mouse before and after elicitation for various time periods at a predetermined site with a micrometer, and the difference is expressed as ear swelling.

Quantitative RT-PCR. Total RNA was extracted from tissues and cells using TRIzol reagent (Invitrogen). First-strand cDNA synthesis was performed using a High Capacity cDNA Reverse transcription kit (Applied Biosystems). PCR reactions were performed using Power SYBR Green PCR system (Applied Biosystems) or TaqMan Gene Expression System (Applied Biosystems) on the ABI7700 Quantitative PCR system (Applied Biosystems). The probe/primer sets used were listed in Table S3.

Immunohistochemistry. Paraffin-embedded tissue sections (~4–7 µm thick) were incubated with Target Retrieval Solution (Dako), and then with rabbit anti-mouse or human PLA2G2D antibody, which did not cross-react with other sPLA₂s (Degounee et al., 2002), or control antibody at 1:2,000 dilution in 10 mM Tris-HCl (pH 7.4) containing 0.15 M NaCl and 0.1% BSA (TBS-BSA) overnight at 4°C. The sections were then treated with an EnVision+ System Staining kit (Dako) with diaminobenzidine substrate, followed by counterstaining with hematoxylin (Murakami et al., 2008; Sato et al., 2010). Human LN sections were obtained by surgery at Toho University Ohmon Hospital (Tokyo, Japan) after approval by the ethical committee of the faculty and informed consents from patients.

Confocal laser microscopy. Mouse frozen tissues mounted in OCT compound (Sakura Finetek) were cut into 20-µm thick sections using a cryomicrotome (CM3050S; Leica). The sections were incubated with 1.5 x Block Ace (DS Pharma Biomedical) in TBS for 30 min, and then with rabbit anti-mouse PLA2G2D antibody, phycoerythrin-conjugated hamster anti-mouse CD11c antibody (N418, ebioscience), or FITC-labeled rat anti-mouse F4/80 antibody (BM8; BioLegend) in TBS-BSA overnight at 4°C. For human tissues, rabbit anti-human PLA2G2D antibody and Alexa Fluor 488-labeled mouse anti-human CD11c antibody (3.9; BioLegend) were used. The sections were then treated with 1 µg/ml Alexa Fluor 647-labeled goat anti-rabbit IgG (Invitrogen) in TBS-BSA for 1 h, mounted in a VectaShield mounting medium (Vector laboratories), and analyzed with a confocal laser scanning microscope (LSM510 META; Carl Zeiss).

Flow cytometry. Mouse spleens or inguinal LNs were excised, minced in Hanks’ solution (Nissui Pharmaceutical) with 2% (vol/vol) heat-inactivated FBS (Invitrogen) and 0.05% (wt/vol) sodium azide (Nacalai Tesque), and incubated with 400 U/ml collagenase type II (Worthington) with shaking for 30 min at 37°C. After adding 10 mM EDTA, the suspensions were passed through Cell Strainer 70-µm Nylon (Falcon; BD), and then centrifuged at 300 g for 5 min at 4°C. Splenocytes were treated for 2 min on ice with 10 mM Tris-HCl (pH 8.0) containing 0.84% (wt/vol) ammonium chloride to lyse red blood cells, centrifuged, and suspended in a 1:4 volume of appropriate MicroBeads (Miltenyi Biotec). After incubation for 15 min at 4°C, cells (<10<sup>6</sup> cells/500 µl) were resuspended in AutoMACs buffer (PBS, 0.5% [wt/vol] BSA, and 2 mM EDTA), blocked with 0.5 µg/ml rat anti-mouse CD16/CD32 antibody (mouse BD Tc Block; BD) for 5 min at 4°C, and were added with 1/4 volume of appropriate MicroBeads (Miltenyi Biotec). For flow cytometry, the cells were blocked with mouse BD Tc Block, incubated with marker antibodies (listed in Table S4), and analyzed with a FACSaria III (BD) and FlowJo (Tree Star) software.

Culture of LN cells. LN cells (10<sup>6</sup> cells/ml) were isolated from mice that had been sensitized with DNFB for 5 d and cultured with or without 100 µg/ml DNBS (Alfa Aesar) in culture medium (RPMI 1640 containing 10% heat-inactivated FBS, 50 µM 2-ME, 2 mM L-glutamine, 25 mM Heps, 1 mM nonessential amino acids, 1 mM sodium pyruvate, 100 U/ml penicillin, and 100 µg/ml streptomycin). After appropriate periods, RNA was extracted from the cells and subjected to quantitative RT-PCR, and the supernatants were subjected to ELISA for cytokines (ebioscience). As required for experiments, PUFAs, 15d-PGJ<sub>2</sub>, and RvD1 (Cayman Chemicals) were added to the culture.

Culture of BMDCs and BMMDs. BM cells were cultured with mouse GM-CSF (10 ng/ml; PeproTech) for 9 d and with mouse M-CSF (100 ng/ml; Kyowa Kirin) for 3 d to obtain BMDCs and BMMDs, respectively (Lutz et al., 1999). The cells (2 × 10<sup>6</sup> cells/ml) were stimulated with LPS (Sigma-Aldrich) for 24 h to assess Pla2g2d expression by quantitative RT-PCR, cytokine release by ELISA, or surface expression of DC activation markers by FACS. For transfer of BMDCs into mice, the cells (2 × 10<sup>6</sup> cells/ml) were treated with 100 µg/ml DNBS for 24 h, resuspended in PBS at 10<sup>6</sup> cells/200 µl, and injected intravenously into recipient mice. After 5 d, the ears were challenged with 40 µl of 0.3% DNFB to monitor ear swelling.

Immunoblotting. Spleen (10 mg) was homogenized in 100 µl of 20 mM Tris-HCl (pH 7.5) with a Polytron homogenizer. Protein concentrations were determined with a BCA protein assay kit (Thermo Fisher Scientific). The homogenates (10 µg protein) were subjected to SDS-PAGE on 15% (wt/vol) gels under nonreducing conditions and were electroblotted onto PVDF membranes (Bio-Rad Laboratories) with a semi-dry blotter (Transblot SD; Bio-Rad Laboratories). The membranes were blocked with 5% (wt/vol) skim milk in PBS containing 0.05% (vol/vol) Tween 20 (PBS-T), probed with goat anti-PLA2G2D antibody (sc-104297; Santa Cruz Biotechnology, Inc.) at 1:5,000 dilution in PBS-T for 2 h, incubated with horseradish peroxidase–conjugated anti-goat IgG (Zymed) at 1:5,000 dilution in PBS-T for 2 h, and then visualized by ECL Prime Western blotting detection reagent (GE Healthcare) with LAS-4000 (Fuji Film).
Tris-HCl (pH 7.4) containing 4 mM CaCl₂ and then incubated for 1 h at 37°C. The tissue-extracted lipids (250 µg) was sonicated for 5 min in 100 mM acetonitrile: methanol = 4:1 (vol/vol) at a flow rate of 50 µl/min at 45°C. The samples injected by an autosampler (10 µl) were separated by a step gradient with mobile phase A (water containing 0.1% acetic acid) and mobile phase B (acetonitrile: methanol = 4:1; vol/vol) at a flow rate of 50 µl/min at 45°C. The detection of each lipid was performed by multiple reaction monitoring (Yamamoto et al., 2011).

PLA₂ reaction. Lipids were extracted from mouse LNs by the method of Bligh and Dyer (Bligh and Dyer, 1959). The LN membrane mimic composed of the tissue-extracted lipids (250 µg) was sonicated for 5 mm in 100 mM Tris-HCl (pH 7.4) containing 4 mM CaCl₂ and then incubated for 1 h at 37°C with 200 nM recombinant mouse PLA₂G2D (Rouault et al., 2007). After incubation, the lipids were mixed with internal standards (250 pmol of PE or PC with 14:0-14:0), extracted, and subjected to ESI-MS for detection of fatty acids, phospholipids, and lysophospholipids (Sato et al., 2010; Yamamoto et al., 2011).

Statistical analysis. Data are presented as the mean ± SEM. Statistical significance was evaluated by two-tailed Student’s t test and Dunnett test at a significance level of P < 0.05.

Online supplemental material. Fig. S1 shows flow cytometry of splenocytes used in Fig. 1 D. Tables S1 and S2 show serum parameters and circulating blood cells in Pla2g2d⁻/⁻ and Pla2g2d⁺/⁺ mice under normal conditions. Table S3 is a list of primers used in quantitative R-T-PCR. Table S4 is a list of antibodies used in flow cytometry. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20121887/DC1.

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REFERENCES


B subunits (p65, p50) in stimulated rheumatoid synovial...


\[
\Delta^12,14-\text{prostaglandin J2 inhibits multiple steps in the NF-κB signal-
\]


**SUPPLEMENTAL MATERIAL**

Mike et al., http://www.jem.org/cgi/content/full/jem.20121887/DC1

Figure S1. Flow cytometry of mouse splenocytes. Mouse splenocytes were separated into CD11c+ and CD11c− populations by AutoMACS. CD11c− cells were further sorted into CD3+ T cells, B220+ B cells, and CD11b+ macrophages. Purities of individual cell populations, which were used in Fig. 1 D, are indicated by boxes.

Table S1. Serum biochemical parameters in Pla2g2d+/+ and Pla2g2d−/− mice under normal housing conditions

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Genotypes</th>
<th>Units</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkaline phosphatase</td>
<td>135.3 ± 70.5</td>
<td>U/liter</td>
</tr>
<tr>
<td>Alanine aminotransferase</td>
<td>30.9 ± 11.1</td>
<td>U/liter</td>
</tr>
<tr>
<td>Bilirubin</td>
<td>0.3 ± 0.1</td>
<td>mg/dl</td>
</tr>
<tr>
<td>Albumin</td>
<td>3.7 ± 0.5</td>
<td>mg/dl</td>
</tr>
<tr>
<td>Nitrogen urea</td>
<td>28.5 ± 4.9</td>
<td>mg/dl</td>
</tr>
<tr>
<td>Total cholesterol</td>
<td>77.4 ± 28.1</td>
<td>mg/dl</td>
</tr>
</tbody>
</table>

Male 15-wk-old Pla2g2d+/+ and Pla2g2d−/− mice were anesthetized and blood samples were collected by cardiac puncture. Sera were applied to a clinical chemistry analyzer VetScan with V-DPP rotors (n = 8). Data are pooled results from two experiments. None of these parameters differed significantly between the genotypes.
Table S2. Circulating blood cells in Pla2g2d+/+ and Pla2g2d−/− mice under normal housing conditions

<table>
<thead>
<tr>
<th>Cell populations</th>
<th>Genotypes</th>
<th>Units</th>
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</thead>
<tbody>
<tr>
<td></td>
<td>Pla2g2d+/+</td>
<td>Pla2g2d−/−</td>
</tr>
<tr>
<td>Leukocytes</td>
<td>8.18 ± 1.11</td>
<td>7.89 ± 1.40</td>
</tr>
<tr>
<td>Lymphocytes</td>
<td>7.02 ± 1.04</td>
<td>6.47 ± 1.07</td>
</tr>
<tr>
<td>Granulocytes</td>
<td>0.76 ± 0.11</td>
<td>0.96 ± 0.29</td>
</tr>
<tr>
<td>Monocytes</td>
<td>0.39 ± 0.15</td>
<td>0.46 ± 0.18</td>
</tr>
<tr>
<td>Erythrocytes</td>
<td>9.97 ± 0.36</td>
<td>9.98 ± 0.13</td>
</tr>
<tr>
<td>Platelets</td>
<td>0.59 ± 0.04</td>
<td>0.55 ± 0.04</td>
</tr>
</tbody>
</table>

Blood samples of 15-wk-old male Pla2g2d+/+ and Pla2g2d−/− mice were collected from tail biopsy with 10 mM EDTA as an anti-coagulant. The samples were applied to the clinical blood cell analyzer VetScan HM to determine the numbers of circulating blood cells (n = 9). Data are pooled results from two experiments. None of these parameters differed significantly between the genotypes.

Table S3. A list of primers used in quantitative RT-PCR

<table>
<thead>
<tr>
<th>Gene</th>
<th>Accession number</th>
<th>Forward primer</th>
<th>Reverse primer</th>
<th>Probe number (Roche)</th>
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<tr>
<td>Ccl2</td>
<td>NM_011333.3</td>
<td>5′-CATCCACGTGTTGGCTCA-3′</td>
<td>5′-GATCATCTTGCTGGAATGCT-3′</td>
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<tr>
<td>Ccr2</td>
<td>NM_009915.2</td>
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<td>Cd11b</td>
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*Used in combination with biotin-labeled antibodies.