miR-612 suppresses the invasive-metastatic cascade in hepatocellular carcinoma

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MicroRNAs (miRNAs) play a critical role in tumor metastasis. In this study, we identified a set of 32 miRNAs involved in hepatocellular carcinoma (HCC) metastasis. Among them, miR-612 was shown for the first time to have inhibitory effects on HCC proliferation, migration, invasion, and metastasis. AKT2 was verified to be one of the direct targets of miR-612, through which the epithelial–mesenchymal transition (EMT) and metastasis were inhibited. The level of miR–612 in HCC patients was inversely associated with tumor size, stage, EMT, and metastasis. Of particular importance, miR-612 is involved in both the initial and final steps of the metastatic cascade, by suppressing local invasion and distant colonization. The pleiotropic roles of miR-612 in the HCC metastatic cascade suggest that it could be an effective target for both early and advanced HCC.

As indicated in published statistics, metastases account for 90% of cancer-related deaths (Gupta and Massagué, 2006). Metastasis has always been a bottleneck in tumor prognosis and therapy. The invasive-metastatic cascade is a complex, multistep process that involves the escape of tumor cells from a primary carcinoma, followed by their intravasation into the systemic circulation, survival during transit through the vasculature, extravasation into the parenchyma of distant tissues, establishment of micrometastases, and ultimately outgrowth of macroscopic secondary tumors (Fidler, 2003). Transcriptional expression studies have revealed that sets of genes whose expression profiles in primary tumors are correlated with metastatic relapse and poor survival can be used as prognostic signatures in cancer patients (Ye et al., 2003; Nevins and Potti, 2007). Some of these genes have been validated through functional and clinical studies as candidate mediators of metastasis (Castellano et al., 2008; Sun et al., 2008; Xue et al., 2010). However, few molecules have been implicated in multiple steps of metastatic cascades.

Mature microRNAs (miRNAs) are short, noncoding RNA molecules with 21–24 evolutionarily conserved nucleotides. These small cellular RNAs act as negative regulators of gene expression by binding to the complementary sequences in 3′ untranslated regions (3′UTRs) of specific target mRNAs of protein-coding genes, leading to degradation of the target mRNAs, inhibition of their translation, or both (Ambros, 2004; Bartel, 2004, 2009). Deregulations of miRNAs have been linked to many human diseases, and the central role of miRNAs in the establishment and progression of human tumors has begun to emerge (Voorhoeve et al., 2006; Ma et al., 2007; Tavazoie et al., 2008; Ventura and Jacks, 2009). Recently, high-throughput screening for miRNA expression profiles has revealed that sets of miRNAs (or “signatures”) exist in hepatocellular carcinoma (HCC), which enable clinicians to predict the disease status and prognosis (Ji et al., 2007, 2009; Budhu et al., 2008). Several miRNAs with prometastatic (miR–151, -143, and -30d) or antimetastatic (miR–9, -139, -503, -338, -125, -122, and -101) functions have been identified using HCC cell lines and xenograft animal models (Huang et al., 2009; Li et al., 2009; Tsai et al., 2009;
The highest efficiencies were achieved in these cells with a dose of at least 50, 100, and 100 nM, respectively (unpublished data). The level of miR-612 after transient transfection was then evaluated with a luciferase reporter system, which showed that 100 nM of miR-612 mimic (miR-612-o) decreased luciferase activity by more than 2.4 fold in HCCLM3 cells and 100 nM of miR-612 inhibitor (miR-612-i) increased luciferase activity by more than 2.2 and 4.0 fold in HepG2 and SMMC-7721, respectively (unpublished data). The cell proliferation of HCCLM3 and HepG2 was significantly affected by miR-612-o and miR-612-i, respectively. Treatment with miR-612-o resulted in significant inhibition of HCCLM3 proliferation by 32.23%, whereas miR-612-i enabled HepG2 to accelerate its proliferation by 25.77%, compared with the non-transfected (WT) cells. No statistical differences in HCCLM3 and HepG2 proliferation were found in WT, Lipofectamine, and mock groups (Fig. 1, D and E).

Next, we assessed the possible roles of miR-612 on HCC migration and invasion in three independent experiments. First, we investigated the relationship between the endogenous levels of miR-612 and migration and invasion capability in HCCLM3, MHCC97H, HepG2, and SMMC-7721 cells. The results showed that motility and invasive-ness were inversely associated with endogenous miR-612 levels (Fig. 1 C, right). Second, we further explored the relationship in nine HCCLM3-RFP-derived monoclonal cells and again found a clear negative correlation between the migration and invasion capabilities and endogenous miR-612 levels (unpublished data). Third, loss- and gain-functions of miR-612 were investigated using a transient transfection strategy with miR-612 mimics or inhibitors. The migration and invasion capabilities of HCCLM3 cells were reduced by more than 4.4 and 6.6 fold, respectively, by miR-612-o compared with the capabilities of control cells (P = 0.0017; P = 0.0042; Fig. 1, F [top] and G). In contrast, in HepG2 cells, migration and invasion increased by more than 6.8 and 6.1 fold with miR-612-i treatment, compared with non-transfected counterparts (P < 0.0001, P = 0.0002; Fig. 1, F [bottom] and H). The promigratory and proinvasive effects of miR-612-i were further confirmed in SMMC-7721 cells (unpublished data). Collectively, our findings suggest that miR-612 negatively regulates tumor migration and invasion of HCC.

miR-612 suppresses the initial step of tumor metastasis in vivo

Local invasion is the initial step in tumor metastasis. The inhibitory effects of miR-612 on in vitro cell migration and invasion of HCC indicated that miR-612 may also inhibit in vivo metastatic initiation. To address this possibility, we stably overexpressed miR-612 in HCCLM3 (unpublished data), the cell line with the highest metastatic potential, and then established an orthotopic xenograft model in nude mice. After 42 d, WT and mock tumors displayed conspicuous intraliver and microvascular invasion (Fig. 2 A). In stark contrast, miR-612 overexpressed xenograft tumors exhibited almost no local invasion (Fig. 2 A). These findings demonstrate...
Figure 1. Systematic identification and functional evaluation of HCC metastasis-related miR-612. (A) Hierarchical clustering of 32 differentially expressed miRNAs. The scale bar at the bottom depicts the standard deviation from the mean. PT and MT represent primary tumor and metastatic tumor, respectively. (B) Pathway perturbation assays. The heat map highlights the highest coefficient of miRNA interference on the signal pathway (red bracket). (C) Association between endogenous levels of miR-612 and motility and invasiveness in HCC cells. (C, left) Real-time PCR analysis to quantify the endogenous levels of miR-612 in HCCLM3, MHCC97H, HepG2, and SMMC-7721 cells. U6 was included as a control and data were normalized to the level of HCCLM3 cells. n = 3. (C, right) Transwell analysis to determine motility and invasion abilities of HCC cells. (D and E) Cell proliferation of HCCLM3 and HepG2 after transient transfections with indicated oligonucleotides. WT, Lipo2000, mock, miR-612-o, or miR-612-i were defined as nontransfected, Lipo2000-treated, negative control oligonucleotide transfected, miR-612 mimic transfected, or miR-612 inhibitor transfected, respectively. Data are mean ± SEM and are representative of three independent experiments.
Figure 2. The suppressive effects of miR-612 on HCC invasion, colonization, and distant metastasis. (A) Pathology analysis of tissue sections from recipient mice at week 6 after transplantation. (top) Hematoxylin and eosin (H&E) staining; (bottom) anti-mouse CD34 staining. Boxed area highlights intraliver dissemination and vascular invasion of tumor cells. (B and C) Lung colonization and outgrowth in recipient mice on day 42 after tail vein injection with $1.0 \times 10^5$ tumor cells. (B) Quantitation of tumor foci (IOD of fluorescent area) in lung of mice injected HCCLM3 and HepG2 cells. (C) Images showing the colonization and outgrowth foci of HCCLM3-RFP and HepG2-GFP cells in the lung of recipient mice. (D) Images showing the lung colonization in the recipient mice on day 3 after tail vein injection with $1.0 \times 10^5$ HCCLM3-RFP and HepG2-GFP cells. (E-G) Intraliver and lung metastasis in the recipient mice on day 42 after HCCLM3-RFP orthotopic transplantation. (E) Images showing metastatic foci in liver (top) and lung (bottom). (F) Quantitation of tumor foci (IOD of fluorescent area) in the liver and lung of recipient mice. (G) Incidence of intraliver dissemination and lung metastasis in WT, mock, and miR-612-o HCCLM3 xenografts. WT, mock, miR-612-o, or miR-612-i was defined as nontransfected, negative control oligonucleotide transfected, miR-612 mimic transfected, or miR-612 inhibitor transfected, respectively. Bars: (A) $100 \mu$m; (C) fluorescent image $2.0 \text{ mm}$; (C, H&E image) $100 \mu$m; (D) $100 \mu$m; (E, liver) 6.0 mm; (E, lung) 2.0 mm. Data are mean ± SEM ($n=5$) and are representative of three independent experiments.
that ectopic forced expression of miR-612 was able to suppress tumor local invasion, the initial step of the metastatic cascade.

miR-612 suppresses the final step of tumor metastasis in vivo

Colonization and outgrowth at a distant site are the last key step in the metastatic cascade. To test whether miR-612 is involved in this step, a tail vein injection model was used to imitate the pathophysiological process of colonization and outgrowth of disseminated tumor cells. We first injected HCCLM3-RFP cells pretreated with none, mock, or miR-612-o into athymic nude mice via the tail vein. On day 42 after injection, the fluorescent area of tumor foci in the lung of recipient mice was reduced by fourfold in miR-612-o group compared with WT and mock groups (Fig. 2, B and C [left]). We then injected HepG2-GFP cells pretreated with none, mock, or miR-612-i using the same protocol. As expected, inhibition of miR-612 activities resulted in a massive colonization and macrotumor foci of HepG2-GFP in the lung of nude mice (Fig. 2, B and C [right]). Similar results were also found in SMMC-7721, a HCC cell line with a high level of miR-612 and little metastatic potential (unpublished data). To pinpoint the effect of miR-612 on HCC colonization, HCCLM3-RFP and HepG2-GFP cells pretreated as above were injected into the tail vein of nude mice. After 3 d, the number of colonized HepG2-GFP cell blocks in lung was significantly increased in the miR-612-i group compared with controls, whereas the number of HCCLM3-RFP cell blocks was markedly decreased in the miR-612-o treatment group (Fig. 2 D). Thus, miR-612 appears to play a suppressive role in HCC colonization and outgrowth in the last stage of the invasive-metastatic cascade.

miR-612 suppresses metastasis of HCC in vivo

The preceding results indicated that miR-612 could suppress both the initial invasion and final colonization of HCC tumor cells. Thus, we speculated that miR-612 would suppress HCC metastasis in an orthotopic xenograft model. Again, a miR-612 stably overexpressed HCCLM3 xenograft model was used. On day 42 after transplantation, miR-612-overexpressed xenograft tumors displayed significantly less intraliver dissemination and lung metastasis in nude mice compared with WT and mock xenograft tumors (Fig. 2, E–G). The number and size (integrated optical density [IOD] of fluorescence) of metastatic foci normalized to primary tumor volume both in liver and lung of recipient mice were significantly decreased by ectopic overexpression of miR-612 (Fig. 2, E and F). In addition, the ratio of intraliver and lung metastasis in the miR-612 overexpressed group was dramatically lower than that of controls (Fig. 2 G). The average size of miR-612 overexpressed tumors was smaller than that of control tumors (P = 0.0443; Fig. 3, A and B). To account for this difference, we reevaluated the number of lung metastases in HCCLM3-RFP xenograft model with a tumor volume–matched endpoint. The results showed that miR-612 overexpression resulted in a dramatic decrease in lung metastasis (Fig. 3, C and D).

miR-612 suppresses the epithelial–mesenchymal transition (EMT)

A typical morphological change of HCCLM3, HepG2, and SMMC-7721 was noted after treatment with miR-612 mimic or inhibitor. Compared with WT cells, HepG2 and SMMC-7721 cells treated with miR-612-i had more changes resembling the EMT, in which the cells scattered and assumed a spindly or star-like morphology, whereas HCCLM3 cells treated with miR-612-o condensed and assumed a cobblestone-like morphology (Fig. 4 A). These phenomena suggested that miR-612 is involved in the EMT of HCC. Therefore, several putative EMT-related markers, such as E/N-cadherin, β-catenin, and vimentin were evaluated by real-time PCR. The mRNA levels of E-cadherin and vimentin were significantly increased and decreased, respectively, in miR-612-o-treated HCCLM3 cells (Fig. 4, B and C). In contrast, the mRNA levels of E-cadherin and vimentin were significantly decreased and increased in both miR-612-i-treated HepG2 and SMMC-7721 cells (Fig. 4, B and C). No significant differences in the expression of other genes were found (unpublished data). The protein changes of these genes were further confirmed by Western blots (Fig. 4, D–F). These observations indicate that miR-612 can suppress the EMT of HCC.

AKT2 is a direct target of miR-612

To elucidate the underlying mechanisms of the suppressive effects of miR-612 on EMT and metastasis of HCC, we used several bioinformatics methods to help identify the target human genes of miR-612. Among the 167 targets predicted by the search programs of miRanda, TargetScan, and miRTarget2, AKT2 was determined to be the gene that localized at one of two main centers of the net comprised of 104 predicted genes and pathways (Fig. 5 A). This finding suggested that AKT2 is a key target of miR-612. Interestingly, the basal levels of AKT2 in several HCC cell lines were negatively correlated with the endogenous levels of miR-612 (Fig. 1 C [left] and Fig. 5 B). Furthermore, the level of AKT2 in HCCLM3 cells was dramatically reduced by miR-612-o, whereas that in HepG2 (Fig. 5 C) and SMMC-7721 (unpublished data) cells was significantly promoted by miR-612-i. The promoting effects of miR-612-i on HepG2 and SMMC-7721 migration and invasion was mostly compromised by AKT2 knockdown mediated by shRNA (AKT2-i) (Fig. 5 D), whereas restoration of AKT2 (AKT2-o) with the expression vector lacking the miR-612 target site in the 3’UTR increased the motility and invasiveness of miR-612-o–treated HCCLM3 cells (Fig. 5 D). In addition, we found one AKT2 inhibitor (AKT inhibitor VIII, VIII) that could abolish the pro-metastatic role of miR-612-i on HepG2 cells in vivo in a dose-dependent manner, and that was notably associated with the attenuated level of phosphorylated AKT2 (Ser 474) in a dose-dependent manner without affecting the total level of AKT2 (Fig. 5, E and F). In other words, AKT2 is a functional target gene of miR-612.

To test whether AKT2 is a direct target gene of miR-612, we cloned a 3’UTR element of AKT2, which has a
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In miR-612-i-treated HepG2 cells, mesenchyma-like cells returned to the epithelial phenotype, whereas co-transfection of miR-612-o with an AKT2 expression vector lacking the miR-612 target site in the 3’UTR abolished the cobblestone-like change in the phenotype of HCCLM3 cells (Fig. 6, A and C). The morphological changes were followed by changes in the mRNA and protein levels of E-cadherin and vimentin (Fig. 6, B and D). These findings suggest that miR-612 regulates the EMT of HCC cells through suppression of AKT2.

miR-612, AKT2, E-cadherin, and vimentin expression in HCC patients

Based on the similar results of experiments using HCC cell lines and xenograft models, it appears that the decreased expression of miR-612 would induce the EMT, and thus promote HCC invasion and metastasis through the AKT2 pathway. To investigate whether this is true in HCC patients, the relationship between miR-612 level and metastatic status was first tested.
Figure 4. The inhibitory roles of miR-612 on EMT of HCC cells. (A) Image showing the morphological changes of HCCLM3, HepG2, and SMMC-7721 after treatment with indicated reagent at hour 72. Bars: (HCCLM3 and SMMC-7721 cell) 100 µm; (HepG2 cell) 200 µm. (B and C) Real-time PCR analysis to quantify the level of E-cadherin and vimentin in HCCLM3, HepG2, and SMMC-7721 cells. β-Actin was included as a loading control and all data were normalized by WT cells. (D–F) Western blot analysis to evaluate the protein levels of E-cadherin (E-cad) and vimentin (VIM) in HCCLM3, HepG2, and SMMC-7721. β-Actin was used as a loading control. WT, mock, miR-612-o, or miR-612-i were defined as nontransfected, negative control oligonucleotide transfected, miR-612 mimic transfected, or miR-612 inhibitor transfected, respectively. Data are mean ± SEM (n = 3) and are representative of three independent experiments.
Figure 5. Direct regulation of miR-612 on AKT2 expression. (A) Bioinformatics analysis to predict target gene of miR-612 and assay pathway perturbation. (B) Western blot analysis to quantify endogenous levels of AKT2 in HCCLM3, MHCC97H, HepG2, and SMMC-7721 cells. (C) Protein levels of AKT2 in HCCLM3 and HepG2 cells after indicated treatments were detected by Western blots. (D) Motility and invasion assays for HCCLM3 (left) and HepG2 (right) cells after treatment with miR-612 inhibitors or control. (E) Luciferase activity assay for HCCLM3 and HepG2 cells with 3’UTR of AKT2 or vehicle.
These findings revealed a significant association between the low expression of miR-612 and HCC local dissemination, as well as distant metastasis. Next, we investigated whether miR-612 level was associated with the protein levels of AKT2 and EMT biomarkers in these patients. Immunohistochemical assays revealed distinct expression patterns of AKT2, E-cadherin, and vimentin in the 14 metastatic and 23 nonmetastatic patients. Higher protein levels of AKT2 and vimentin were found in the metastatic patients, whereas a higher protein level of E-cadherin was found in the nonmetastatic patients (Fig. 7, C–E). Furthermore, tumors with a high level of miR-612 tended to express low levels of AKT2 and vimentin and a high level of E-cadherin (as illustrated in patient 9 with no metastasis), whereas tumors with a low level of miR-612 (as seen in patient 37 archived primary liver tumors using real-time PCR. The clinicopathological features of these patients are listed in Table S1. Tumors from 14 patients who had portal vein thrombosis, microvascular invasion, and intrahepatic dissemination tended to express lower levels of miR-612 compared with those from nonmetastatic patients (0.2771 ± 0.0886 vs. 1.475 ± 0.3217; P = 0.0015; Fig. 7 A). The low expression levels of miR-612 were correlated with poor metastasis-free survival (P = 0.0177; Fig. 7 B). Four of the 18 patients with low levels of miR-612 developed lung (3 patients) and bone (1 patient) metastasis within 1 yr, whereas no distant metastases were found in the 19 patients with high levels of miR-612 (4/18 vs. 0/19; P = 0.046). In addition, the level of miR-612 had a significant inverse correlation with tumor size and stage, intrahepatic metastasis, and microvascular invasion (Table S2). These findings revealed a significant association between the low expression of miR-612 and HCC local dissemination, as well as distant metastasis.

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Clinical validation of miR-612, AKT2, E-cadherin, and vimentin. (A) Real-time PCR analysis to quantify the endogenous levels of miR-612 in HCC patients with or without local dissemination/metastasis. U6 was used as a loading control. The amount of miR-612 was normalized to the mean value of all patients. (B) Kaplan-Meier curves of 37 HCC patients after stratification by the level of miR-612 were used for depicting metastasis-free survival. (C–E) Relative levels of AKT2, E-cadherin, and vimentin in HCC patients with or without local dissemination/metastasis. (F–H) Relative levels of AKT2, E-cadherin, and vimentin in patients with low and high miR-612 expression levels. (I and J) Images to visualize positive staining of AKT2, E-cadherin, and vimentin in patients with low and high miR-612 expression levels. The slides are constructed by serial section method. Bars: (left) magnification x100, (right) magnification x400. Data are mean ± SEM and are representative of three independent experiments.
10 with local metastasis) tended to express high levels of AKT2 and vimentin and a low level of E-cadherin (Fig. 7, F–J). Hence, it appears that miR-612 negatively regulates the EMT and tumor metastasis in HCC patients by targeting AKT2.

**DISCUSSION**

Genome-wide miRNA analysis with the miRAGE method revealed that miR-612 was down-regulated in tumors versus nontumorous colorectal tissues (Cummins et al., 2006). However, the biological function of miR–612, especially in the progression of cancer, was unknown. In this study, we revealed for the first time that miR–612 has pleiotropic inhibitory effects on cell proliferation, migration, invasion, and metastasis of HCC. We also verified that the levels of miR-612 were inversely associated with tumor size and stage, local invasion, and distant metastasis in HCC patients, but were positively correlated with metastasis-free survival. These data indicate that miR-612 suppresses metastasis in HCC, but it is unclear whether miR-612 plays an antitumorigenic role in other types of tumors. Therefore, we modulated miR-612 level in the colon cancer cell line HCT116 and found that overexpression of miR-612 reduced liver metastasis of colon cancer in a HCT116 spleen xenograft mouse model and vice versa (Unpublished data). In the present study, we further verified that AKT2 is one of direct targets of miR-612, and its suppressive role on HCC metastasis was facilitated by EMT inhibition through the AKT2 pathway.

The complex invasive-metastatic cascade can be conceptually simplified into two major phases: one is the physical translocation of cancer cells from the primary tumor to a distant tissue, and the other is the colonization and outgrowth of translocated cells within a secondary organ (Chaffer and Weinberg, 2011). We scrutinized the roles of miR-612 in the HCC metastatic cascade step by step. For successful translocation, HCC cells must acquire the ability of migration and invasion in the initial stage. Indeed, up-regulation of miR-612 was able to suppress HCC cell invasion into the surrounding normal liver and microvascular tissue in nude mice model. Clinical data also confirmed that miR-612 levels were inversely associated with intrahepatic metastasis and microvascular invasion of patients. These findings suggest that miR-612 plays a suppressive role in the initial step of the HCC metastatic cascade.

Tumor colonization is an utterly inefficient and rate-limiting step in the metastatic cascade, in which millions of cancerous cells may be released by a primary tumor into systemic circulation every day, but only a small minority of these cells will colonize a distant organ (Gupta and Massagué, 2006). This study also underscored the influence of miR–612 during this stage using a tail vein injection model, which is theoretically analogous to the colonization and outgrowth phase of the tumor metastatic cascade. Our results showed that overexpression of miR–612 strikingly diminished the number of lung tumor foci in HCCLM3-RFP xenograft model, and the loss of function of miR–612 markedly enhanced lung colonization in HepG2. These results suggest that miR–612 plays a pivotal role in multiple rate-limiting steps in the metastatic cascade.

Obtaining an understanding of the process of metastatic dissemination of cancer cells is in sight. The EMT appears to be essential for cancerous cells to acquire the capability of migration and invasion and is a key driver of tumor cell translocation (Polyak and Weinberg, 2009). This idea is supported by our finding that miR–612 negatively regulated EMT, local invasion, and dissemination of HCC. As tumor colonization and outgrowth in a distant organ seems to represent a far more complex set of phenomena with a relatively small number of unifying, generalizable principles (Chaffer and Weinberg, 2011), we cannot yet define the underlying mechanism of miR–612 in this process. However, a cancer stem cell (CSC)–like state of HCC cells induced by EMT, accompanied by gaining the traits of self-renewal and apoptosis resistance (Charafe-Jauffret et al., 2009; Thierry et al., 2009), may provide a reasonable explanation of the role of miR–612 in this stage. This hypothesis is based on the following observations: (a) overexpression of miR–612 greatly inhibited in vitro cell proliferation and suppressed in vivo tumor growth of HCC in nude mice; (b) restoration of miR–612 first reduced HCCLM3 cells blocks in lung and then inhibited the outgrowth to a second tumor, whereas the inhibition of miR–612 in HepG2 cells resulted in the opposite results; and (c) tumors of patients with high levels of miR–612 were two times smaller than those of patients with low levels of miR–612. Therefore, we speculate that miR–612 may interfere with the initiation of a CSC-like state of HCC through the inhibition of the EMT, which thus suppresses tumor growth and metastasis. Although the detail mechanisms need to be further investigated, miR–612 did suppress both the initial and final steps of the invasive-metastatic cascade of HCC.

The full spectrum of signaling agents that contribute to EMTs of carcinoma cells remains unclear. Evidence indicates that activation of the EMT process during tumorigenesis and progression most often requires signals from reactive stroma (Yang and Weinberg, 2008). Recently, the involvement of miRNAs in the EMT process was demonstrated (Bracken et al., 2008; Gregory et al., 2008; Park et al., 2008). For example, miR–205 and miRNAs from the miR–200 family (miR–200a, -200b, -200c, -141, and -429) are closely related with regulation of the EMT. ZEB1 and ZEB2/SIP1 are subsequently associated with the loss of function of miR–612. Hence, EMT inhibition or a low endogenous level of miR–612 expression was inversely correlated with EMT phenotype. Forced down-regulation of translocated cells within a secondary organ (Chaffer and Weinberg, 2011), this idea is supported by our finding that miR–612 negatively regulated EMT, local invasion, and dissemination of HCC. As tumor colonization and outgrowth in a distant organ seems to represent a far more complex set of phenomena with a relatively small number of unifying, generalizable principles (Chaffer and Weinberg, 2011), we cannot yet define the underlying mechanism of miR–612 in this process. However, a cancer stem cell (CSC)–like state of HCC cells induced by EMT, accompanied by gaining the traits of self-renewal and apoptosis resistance (Charaf...
of miR-612 on the early and final steps of the HCC invasive/metastatic cascade suggest that it could be a useful target for interfering with HCC metastasis in clinical practice.

MATERIALS AND METHODS

Cell lines. Four human hepatoma cell lines and nine monoclonal cell strains from HCCLM3-RFP cells were used in this study. HepG2 and SMMC-7721 cells were purchased from the Shanghai cell bank, Chinese Academy of Sciences. MHCC97H, HCCLM3, and its red fluorescence protein (RFP)–expressing derivatives were described previously (Sun et al., 1999; Li et al., 2004; Yang et al., 2008). HepG2-GFP cells were established previously by our group (Yang et al., 2008). Human colon cancer cell HCT116 was provided by C.H. Tang (Minghong Biotechnology Inc., Shanghai, China). All these cells were cultured under standard conditions.

Constructs, oligonucleotides, and transfection. Three GV248 lentiviral particles containing short hairpin sequences targeting the human AKT2 gene and the GV142 vector with human AKT2 complementary DNA without miR-612 targeting sites in 3′UTR, as well as the negative plasmids, were purchased from GeneChem. The three pooled shRNA sequences that yielded the greatest knockdown were: 5′-GACAGGAGGCTCCTGCTGATGCGGTAAGCTGAGAA-3′, 5′-ACAAGGTAATCCCGATGGAACCTGTG-3′, and 5′-CCCTTAAACAACTTCTCCGTACTCGAGTACGGAGAAGTT-3′. Primers for amplification of human AKT2 were as follows: 5′-CCACCTTTTGTCGAAAAAGTTGGCC-3′ (sense) and 5′-AAATGCCA-ACCTCCTGGTAAAGG-3′ (antisense). The oligonucleotides including miRIDIAN miR-612 hairpin inhibitor, mimic, and negative control (Thermo Fisher Scientific) for inhibition and restoration of miR-612 were used in this study. All these constructs and oligonucleotides were transfected into HCC cells using Lipofectamine 2000 according to the product manual (Invitrogen). A luciferase reporter system that contains putative recognition elements of the binding sites for miR-612 in the 3′UTR, as well as the negative plasmids, were provided by C.H. Tang (Minghong Biotechnology Inc., Shanghai, China). All these cells were cultured under standard conditions.

Luciferase reporter assay. The binding sites for miR-612 in the 3′UTR sequence of AKT2 were cloned into the pmirGLO Dual-Luciferase miRNA Target Expression Vector (Promega). The miR-612 recognition elements were as follows: 5′-AAACTAGGCAGGTGGCCCTACCTTTTGTGCGGACACAGGTTTTT-3′ (sense) and 5′-CTAGGAAATCCTGGTCTGGGGCACAAAGGGTGTTAGG-3′ (antisense). AKT2 complementary DNA was purchased from Fulengen Co. Ltd. These constructs (1.0 µg) were co-transfected into target cells in 24-well plates together with 100 nM of miR-612 hairpin inhibitors or mimic using Lipofectamine 2000. Luciferase activity was measured 24 h after transfection using the Dual-Luciferase Reporter Assay System (Promega). The levels of firefly luciferase activities were obtained by normalizing to Renilla luciferase activities and relative to a control, as previously reported (Tavaoe et al., 2008).

Stable expression of miR-612 with lentiviral vector. Human miR-612 gene was PCR amplified from normal genomic DNA and cloned into pEZK-Am03-mC FP (mCherry Fluorescent Protein) for ectopic expression of miR-612. Primers used for amplification were 5′-CCACGAGAAGATCCAGTGAGTGGAGGACAC-3′ (sense) and 5′-AATATACCC-TTCTCGAGACACACACTGCTGTGCTGCT-3′ (antisense; Xhol site marked as bold). The following primers were used for PCR to confirm the insertion: 5′-ATGAGGCTCTAGATTACTTACAG-3′ (MR30-F) and 5′-CATAGGCGGGTGATGGACACGAC-3′ (forward) and 5′-ACACTTGTAACGTGCGGACGAC-3′ (reverse) for E-cadherin; 5′-CCGCTGGATTCACCTGCTCTGGAGG-3′ (forward) and 5′-TGGCGGGGATCCGCGAGGACC-3′ (reverse) for N-cadherin; 5′-GATCCACTGATGMCGGAGACGAC-3′ (forward) and 5′-TGGTAGGTCAGCCTTTCAATGTGCACT-3′ (reverse) for vimentin; 5′-ATGCGAAGATCCCTGGTGATGGACACGAC-3′ (forward) and 5′-CCAGATGCGGAGAGGAGGACGAC-3′ (reverse) for N-cadherin; 5′-CTTCTCTGACCTGATGACGAGGGAC-3′ (forward) and 5′-TGGTAGGTCAGCCTTTCAATGTGCACT-3′ (reverse) for vimentin; 5′-ATGAGGCTCTAGATTACTTACAG-3′ (forward) and 5′-CATAGGCGGGTGATGGACACGAC-3′ (forward) and 5′-ACACTTGTAACGTGCGGACGAC-3′ (reverse) for E-cadherin; 5′-CCGCTGGATTCACCTGCTCTGGAGG-3′ (forward) and 5′-TGGCGGGGATCCGCGAGGACC-3′ (reverse) for N-cadherin; 5′-GATCCACTGATGMCGGAGACGAC-3′ (forward) and 5′-TGGTAGGTCAGCCTTTCAATGTGCACT-3′ (reverse) for vimentin.

Analysis of miRNA expression. For analysis of miRNA expression, we used the CapitalBio Mammalian miRNA microarray and the service to process the samples (CapitalBio Inc.). The primary tumors and their paired lung metastatic specimens were isolated from orthotopic xenograft models of HCCLM3-RFP cells (Sun et al., 1996; Yang et al., 2008). Total RNAs were prepared with TRIzol Reagent (Invitrogen). miRNAs were purified by precipitation methods with PEG solutions (Thomson et al., 2004; Watanabe et al., 2005). A total of 4 µg RNA from each sample was labeled with Cy3 (Thermo Fisher Scientific), as described previously (Thomson et al., 2004), and hybridized to mammalian miRNA chips containing capture probes against all known human miRNAs (Sanger miRNA version 12.0). After hybridization, signals were scanned, quantified, and normalized with a global mean algorithm. Differentially expressed miRNAs were selected according to the following criteria: the change fold was no less than two times and Q value was set at 5% (SAM, version 2.1; He et al., 2005). Hierarchical clustering of variance-normalized expression values was performed with Cluster 3.0 software for Windows using the metric of Euclidean distance and average linkage. The raw and normalized microarray data have been deposited in the National Institutes of Health NCBI Gene Expression Omnibus under accession no. GSE26323.

RNA extraction and real-time PCR assays for miRNA and mRNA detection. Total RNA was extracted from cultured cells or freshly iced HCC tissues with TRIzol Reagent (Invitrogen). The quality of RNA was examined by A260 absorption, and then 2 µg of total RNA were used for first-strand DNA synthesis. Real-time PCR was performed in triplicate by the SYBR. Green PCR method using an All-in-One miRNA qPCR Detection kit (GeneCopoeia Inc.). The following forward primers were synthesized by GenoCopoeia; 5′-TTCAGAATTCCAGAAGCTGCA-3′ for hsa-miR-26a, 5′-GCCCCCCTCCTACCTGTA-3′ for hsa-miR-26b, and 5′-CCAGGCTCCTGAGCTCTTATTAA-3′ for hsa-miR-30d, 5′-GGCCCGGTTGGGCGGCCCTAA-3′ for hsa-miR-638, 5′-GCAGGCTCTTGCAGCTCTTATTAA-3′ for hsa-miR-612, and 5′-CAAAATCGTGAAGCGTTCTCATAT-3′ for U6 small nuclear RNA (U6). The common reverse primer was purchased from the same company. For miRNA detection, 500 ng of total RNA were used for complementary DNA synthesis with a PrimeScript RT reagent kit (Takara Bio, Inc.). Real-time PCR was performed in triplicate using SYBR Premix Ex Taq (Takara Bio, Inc.). The primers for the genes of interest (E/N-cadherin, β-catenin, vimentin) were synthesized by Sangon Biotech Co., Ltd., as follows: 5′-CTGTCAGGCTCTCATCTAGGAA-3′ (forward) and 5′-ACCTGTTAGACCTGGCAGACT-3′ (reverse) for E-cadherin; 5′-CCCTGGTAAATGGGCA-3′ (forward) and 5′-GATCCACTGATGMCGGAGACGAC-3′ (reverse) for N-cadherin; 5′-GATCCACTGATGMCGGAGACGAC-3′ (forward) and 5′-TGGTAGGTCAGCCTTTCAATGTGCACT-3′ (reverse) for vimentin; 5′-ATGAGGCTCTAGATTACTTACAG-3′ (forward) and 5′-CATAGGCGGGTGATGGACACGAC-3′ (forward) and 5′-ACACTTGTAACGTGCGGACGAC-3′ (reverse) for E-cadherin; 5′-CCGCTGGATTCACCTGCTCTGGAGG-3′ (forward) and 5′-TGGCGGGGATCCGCGAGGACC-3′ (reverse) for N-cadherin; and 5′-GATCCACTGATGMCGGAGACGAC-3′ (forward) and 5′-TGGTAGGTCAGCCTTTCAATGTGCACT-3′ (reverse) for vimentin.

Inhibition of AKT2 activation. AKT inhibitor VIII (Isozyme-selective, AKT1-1/2) (EMD Millipore) was used to inhibit AKT2 activation and its corresponding downstream signal pathway. Final doses of AKT inhibitor VIII ranging from 0.5 to 5 µM were used in the experiments.

In vitro cell proliferation assays. Cell proliferation was determined by the Cell Counting kit-8 (Dojin Laboratories) assay. In brief, HCCLM3 and HepG2 cells were seeded into 96-well plates at an initial number of 2,000 cells per well. At 24, 48, or 72 h after oligonucleotide transfection, 10 µl of the kit reagent was added to each well, and 2 h later all plates were scanned by a microplate reader (Thermo Fisher Scientific) at 450 nm. Cell proliferation was calculated on the basis of absorbency.
In vitro migration and invasion assays. Cell migration and invasion were analyzed by a Transwell Permeable Support system with 8-µm pores (Corning). For motility assays, 5 × 10^4 cells were seeded into upper uncoated inserts; for invasion assays, 1.0 × 10^5 cells were seeded into upper inserts with a Matrigel-coated membrane (BD). Cells were seeded in 1% serum medium and translocated to 1% serum media for 24 or 48 h. After removal of the nonmigrating or noninvading cells, the remaining cells were fixed, stained, and analyzed by inverted microscopy. A total of 100 nM miR-612 inhibitors, mimics, or mock were transfected 60 h before plating using Lipofectamine 2000, as well as AKT2 shRNA (60 nM) or expression vector (80 nM). AKT inhibitor VIII at a concentration ranging from 0.5 to 5.0 µM was used to treat cells for 60 h before seeding.

miRNA-target prediction, pathway perturbation, and miRNA ranking. In silico target genes of all human miRNAs were predicted by miKanda, TargetScan, and miRInvese2 algorithms. The predicted gene accordance in at least two algorithms were retained and used for KEGG pathway annotation, which was obtained from the ”has_pathway.hst” file downloaded from KEGG ftp (ftp://ftp.genome.jp/pub/kegg/). Next, the miRNA pathway relationships were achieved according to the relationships between miRNAs and genes and between genes and the pathway (namely the earlier annotated KEGG pathway of the genes). A miRNA-pathway-weighted matrix MP (m, n) was composed to denote the relationships between all m miRNAs and n pathways. Furthermore, pathway perturbation by differentially expressed miRNAs (*) was evaluated, and the pseudo pathway expression profile was constructed. The GlobalAncova test (Hummel et al., 2008) was performed on the pseudo pathway expression profile to estimate the perturbation degree in samples from primary tumor to lung metastasis. The perturbation degree of each pathway was judged by the resultant F value. Finally, miRNAs were ranked according to their contributions to pathway perturbation. All statistical analyses were performed using R/Bioconductor software (http://www.bioconductor.org).

Protein levels detected by Western blot analysis. Lysates were obtained from cultured cells with a mixture of ProteoJET Mammalian Cell Lysis Reagent (Fermentas) and phenylmethylsulfonyl fluoride (Roche) and PhosSTOP (Roche). Proteins were separated by sodium dodecyl sulfate-PAGE and transferred onto polyvinylidene fluoride membranes. After being blocked in 5% nonfat milk or bovine serum albumin; probed with antibodies against human AKT2 (1:1,000; Cell Signaling Technology), phospho-AKT2 (S474) (1:1,000; Abcam), E-cadherin (1:1,000; Cell Signaling Technology), α-smooth muscle actin (1:1,000; Sigma-Aldrich); and tested with goat anti-rabbit or anti-mouse IgG (1:10,000 for both; Jackson ImmunoResearch Laboratories); and detected with enhanced chemiluminescent reagents (Pierce). Bands were visualized with 3,3′-diaminobenzidine. Density of target proteins, except CD34 used for vascular labeling, was determined as previously reported (Zhu et al., 2008).

Immunohistochemistry assays. Immunohistochemistry for the target molecules was performed on single serial sections made from surgical samples. The slides were probed with a primary antibody against AKT2 (1:200; Cell Signaling Technology), E-cadherin (1:100; Cell Signaling Technology), vimentin (1:100; Bioworld Technology Inc.), and CD34 (1:400; Santa Cruz Biotechnology, Inc.) and then incubated with horseradish peroxidase-conjugated IgG (1:500; Invitrogen), and the proteins in situ were visualized with 3,3′-diaminobenzidine. Density of target proteins, except CD34 used for vascular labeling, was determined as previously reported (Zhu et al., 2008).

Statistical analysis. Data were analyzed using GraphPad Prism 5 software (Tavazoie et al., 2008). Quantitative variables were expressed as means ± SEM and analyzed by one-way ANOVA. Student’s t test, Kruskal–Wallis test, or Mann–Whitney test. Qualitative variables were compared using Pearson χ² test or Fisher exact test. The log-rank test was used to determine the statistical significance of the differences between metastasis-free survival curves. R/Bioconductor software was used for all bioinformatics analysis. Results were considered statistically significant at P < 0.05.

Online supplemental material. Table S1 shows clinicopathological features of HCC patients. Table S2 shows the relationships between intratumoral levels of miR-612 and clinicopathological features of HCC patients. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20120153/DC1.

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Table S1. Clinicopathological features of HCC patients

<table>
<thead>
<tr>
<th>Features</th>
<th>Values/Counts (n = 37)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years, median (range))</td>
<td>55 (30-74)</td>
</tr>
<tr>
<td>Gender (male/female)</td>
<td>29/8</td>
</tr>
<tr>
<td>Liver cirrhosis (yes/no)</td>
<td>28/9</td>
</tr>
<tr>
<td>Tumor size (cm, median (range))</td>
<td>4.0 (2.0–15)</td>
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<tr>
<td>Encapsulation (complete/none)</td>
<td>21/16</td>
</tr>
<tr>
<td>Tumor differentiation (I–II/III–IV)</td>
<td>30/7</td>
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<tr>
<td>Intrahepatic metastasis (yes/no)</td>
<td>5/32</td>
</tr>
<tr>
<td>Microvascular invasion (yes/no)</td>
<td>13/24</td>
</tr>
<tr>
<td>Portal venous emboli (yes/no)</td>
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</tr>
<tr>
<td>TNM stage (I/II/IIIa/IVA)</td>
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</table>
Table S2. Relationships between intratumoral levels of miR-612 and clinicopathological features

<table>
<thead>
<tr>
<th>Variables</th>
<th>miR-612 level</th>
<th>P-value</th>
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<tbody>
<tr>
<td></td>
<td>High (n = 19)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Low (n = 18)</td>
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</tr>
<tr>
<td>Age, years&lt;sup&gt;a&lt;/sup&gt;</td>
<td>51.89 ± 2.13</td>
<td>53.67 ± 2.49</td>
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<tr>
<td>Gender&lt;sup&gt;b&lt;/sup&gt;</td>
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<td></td>
</tr>
<tr>
<td>Male</td>
<td>15</td>
<td>14</td>
</tr>
<tr>
<td>Female</td>
<td>4</td>
<td>4</td>
</tr>
<tr>
<td>Liver cirrhosis&lt;sup&gt;a&lt;/sup&gt;</td>
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<tr>
<td>Yes</td>
<td>14</td>
<td>14</td>
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<tr>
<td>No</td>
<td>5</td>
<td>4</td>
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<tr>
<td>Tumor size (cm)</td>
<td>3.8 ± 0.40</td>
<td>7.21 ± 0.87</td>
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<tr>
<td>Tumor encapsulation&lt;sup&gt;c&lt;/sup&gt;</td>
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<td>Complete</td>
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<td>8</td>
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<tr>
<td>None</td>
<td>6</td>
<td>10</td>
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<tr>
<td>Tumor differentiation&lt;sup&gt;b&lt;/sup&gt;</td>
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<tr>
<td>I–II</td>
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<td>7&lt;sup&gt;e&lt;/sup&gt;</td>
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</tr>
<tr>
<td>I</td>
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<td>II</td>
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</tr>
<tr>
<td>IVa</td>
<td>0</td>
<td>2</td>
</tr>
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</table>

<sup>a</sup> Student’s t test, data described as mean ± SEM.
<sup>b</sup> Fisher’s exact test.
<sup>c</sup> Pearson chi-square test; one-sided.
<sup>d</sup> Chi-square test for trend.
<sup>e</sup> One patient with uncertain microvascular invasion is classified into this subgroup.