A diverse repertoire of high affinity antibodies is required to protect us from foreign antigens. To create such a repertoire, Ig genes in germinal center B cells undergo a high rate of somatic hypermutation (SHM) and class switch recombination (CSR) that lead to the affinity maturation and isotype switching required to generate protective antibodies. These processes are initiated by activation-induced cytidine deaminase (AID), which converts dC to dU in single-stranded DNA (Maul et al., 2011; Stavnezer, 2011). The U:G mismatches created by AID lead to transition mutations through replication or to the recruitment of error-prone base excision repair and mismatch repair (MMR) proteins that introduce additional mutations at the Ig locus (Stavnezer, 2011). Although MMR normally maintains the integrity of the genome, at the Ig genes, it paradoxically mediates extensive mutation and the formation of DNA double-strand breaks (DSBs) at specific switch regions (SRs; Chahwan et al., 2011).

Antibody diversification through somatic hypermutation (SHM) and class switch recombination (CSR) are similarly initiated in B cells with the generation of U:G mismatches by activation-induced cytidine deaminase but differ in their subsequent mutagenic consequences. Although SHM relies on the generation of nondeleterious point mutations, CSR depends on the production of DNA double-strand breaks (DSBs) and their adequate recombination through nonhomologous end joining (NHEJ). MLH1, an ATPase member of the mismatch repair (MMR) machinery, is emerging as a likely regulator of whether a U:G mismatch progresses toward mutation or DSB formation. We conducted experiments on cancer modeled ATPase-deficient MLH1G67R knockin mice to determine the function that the ATPase domain of MLH1 mediates in SHM and CSR. Mlh1GR/GR mice displayed a significant decrease in CSR, mainly attributed to a reduction in the generation of DSBs and diminished accumulation of 53BP1 at the immunoglobulin switch regions. However, SHM was normal in these mice, which distinguishes MLH1 from upstream members of the MMR pathway and suggests a very specific role of its ATPase-dependent functions during CSR. In addition, we show that the residual switching events still taking place in Mlh1GR/GR mice display unique features, suggesting a role for the ATPase activity of MLH1 beyond the activation of the endonuclease functions of its MMR partner PMS2. A preference for switch junctions with longer microhomologies in Mlh1GR/GR mice suggests that through its ATPase activity, MLH1 also has an impact in DNA end processing, favoring canonical NHEJ downstream of the DSB. Collectively, our study shows that the ATPase domain of MLH1 is important to transmit the CSR signaling cascade both upstream and downstream of the generation of DSBs.

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(MLH1 and PMS2), which signals downstream effector functions (Kunkel and Erie, 2005; Iyer et al., 2006; Jiricny, 2006). Both complexes appear equally crucial for the global repair of DNA mismatches, yet their roles in mediating antibody diversity differ. The MutSα complex is required for both SHM and CSR (Chahwan et al., 2011), whereas MutLα seems to function only during CSR (Kim et al., 1999; Phung et al., 1999; Schrader et al., 1999; Ehrenstein et al., 2001; Péron et al., 2008; Stavnezer et al., 2010; van Oers et al., 2010). Although the processes of SHM and CSR are similar in their initial phases because both are triggered by AID-generated U:G mismatches, their outcomes differ in that during CSR, those mutations lead to DSBs that must then be resolved by non-homologous end joining (NHEJ; Kotnis et al., 2009). The activation of the latent endonuclease activity of PMS2 in MutLα complexes contributes to the generation of those DSBs (van Oers et al., 2010). MLH1, as the protein that directly interacts with the mismatch sensor MutSα, anchors the MutL complexes (Kunkel and Erie, 2005; Iyer et al., 2006; Jiricny, 2006) and is required for the recruitment of PMS2 and its endonuclease activity during CSR (Cannavo et al., 2007; van Oers et al., 2010).

MLH1 is a candidate factor to orchestrate the different roles of MutLα in SHM and CSR because it coordinates the MMR process through its physical interactions with the MMR core proteins and through its ATPase activity modulates the timing and spatial relationships of the activities of the MMR complex (Polsinina and Cupples, 2010). The interactome of MLH1 has been shown to contain proteins involved not only in MMR but also in DNA metabolism and repair, among other functions (Cannavo et al., 2007), supporting its role in orchestrating many downstream processes. MLH1 belongs to the GHL (or HATPase) family of ATPases (Bergerat et al., 1997). Inactivating mutations, primarily in the conserved N terminus of hMLH1 or loss of expression of hMLH1, are among the most frequent mutations in hereditary non-polyposis colorectal cancer syndromes (Shah et al., 2010). The recurrent G67R mutation in hMLH1 is located in one of the GxG motifs of the ATPase domain (Fig. 1 A). This particular mutation does not prevent its interaction with PMS2 to form MutL, although it does impede DNA repair functions and promote a strong cancer phenotype (Raevaara et al., 2005; Avdievich et al., 2008).

A recently generated mouse model carrying the MLH1G67R mutation and expressing almost normal protein levels showed a unique opportunity to study the impact of its ATPase activity on the multiple functions of MLH1 during SHM and CSR. We show here that although SHM is not affected by the loss of the ATPase activity of MLH1, CSR is dramatically impaired. The ATPase-dependent functions of MLH1 promote the generation of DSBs that are necessary during CSR and ultimately influence the NHEJ mechanisms that repair such DSBs at the SRs.

RESULTS AND DISCUSSION
Normal SHM in Mlh1GR/GR mice
A highly conserved GxG sequence motif (Fig. 1 A) is located in the loops defining the boundary of the ATP binding pocket of bacterial MutL (Dutta and Inouye, 2000). The mammalian MLH1 ATP binding pocket behaves similarly in a crystal structure resolved at 2.5 Å (Protein Data Bank accession no. 3NA3; Fig. 1 A). Accordingly, the GxG motif is in a polypeptide stretch devoid of secondary structures, suggesting that a G67R mutation should not significantly affect the overall integrity of the ATPase or the adjacent MutS interaction domains. However, G67 appears to be in close proximity to the adenine base of the ATP molecule (Fig. 1 A), suggesting that a G67R mutation could greatly alter the bulkiness and charge of the ATP pocket, thereby hindering the accommodation of the ATP molecule.

ATP binding and/or hydrolysis by the ATPase of MLH1 is believed to induce large conformational changes (Sacho et al., 2008; Johnson et al., 2010) capable of mediating an MMR signaling cascade by recruiting and activating downstream effector proteins after MutS’ binding (Ban et al., 1999; Dutta and Inouye, 2000; Spampinato and Modrich, 2000; Tomer et al., 2002; Chahwan et al., 2011). Although MLH1 and its ATPase activity are required for efficient DNA MMR in vitro and in vivo (Käschele et al., 2002; Tomer et al., 2002; Jiricny, 2006; Avdievich et al., 2008), there has been some uncertainty about the role of MLH1 in the resolution of the AID-generated mismatches during SHM. In fact, the frequency and characteristics of mutations in MLH1-deficient mice were previously shown to be altered in an artificial exogenous transgene but normal when an endogenous Ig variable gene was studied in knockout mice (Kim et al., 1999; Phung et al., 1999). To determine whether the continued presence of the protein but the loss of the ATPase activity of MLH1 had any effect on SHM, three WT and three Mlh1GR/GR mice were immunized with NP36-CGG to promote SHM in the rearranged V<sub>H</sub>186.2 gene. We found no significant differences in the overall mutation frequencies between WT and Mlh1GR/GR mice (P = 0.4252; Fig. 1 B), and the distribution of mutations was similar (P > 0.0576; Fig. 1 C). Although there was a slight tendency for more G:C mutations in mutant mice, the number of transversions or transitions (P < 0.2652), the different types of substitutions (P > 0.0576), and the cumulative spectrum of mutations from G, C, A, or T sites (P > 0.2183) were also comparable between WT and mutant mice (Fig. 1, B and D). Furthermore, the overall mutation frequencies of the hotspots for AID
Figure 1. The MLH1G67R mutation does not have a significant impact on SHM. (A) Alignment of the GxG motif of MLH1 ATPase domain and a 2.5-Å resolution of the backbone of hMLH1 ATP-binding pocket with ATP (Protein Data Bank accession no. 3NA3). G67 of human MLH1 is highlighted with a gray oval, and the ATP molecule is shown inside the MLH1 ATPase pocket. Asterisks signify that the amino acid conservation is identical, and colons signify similarity at the respective amino acid residues. On the right, a cartoon backbone and a surface representation of the first 347 aa residues of mammalian MLH1 along with an ATP molecule trapped in the ATPase domain are shown. The G67 residue does not seem to be in a region having secondary structures but is in close proximity to the adenine base of the ATP molecule and not to the triphosphate group. Although MLH1 was crystallized as a homodimer, for simplicity, only the monomer is shown here. (B) Global analysis of unique mutation frequencies corrected for base composition according to the SHMTool algorithm. (C) Distribution of mutations in the V<sub>186.2</sub> sequence (273 bp). (D) The spectrum of base substitutions is expressed as frequencies of mutation (x10<sup>-2</sup> mutations/base) and corrected for base composition. The cumulative spectrum of mutations from G, C, A, or T sites is also shown (Sum). (E) Proportion of sequences with different load of mutations (indicated in each fraction) for antigen and nonantigen selected regions (V<sub>186.2</sub> and Jh2-Jh4, respectively). The center of the pies represents the total number of sequences analyzed in each group. (F) Relative frequency of mutations at G:C versus A:T sites observed in VH186.2 and Jh2-Jh4 regions.

(WRC/GYW) and Polη (WA/TW), which are major hallmarks of the SHM process, were similar in both groups (P ≥ 0.5287; Fig. 1 B). Finally, the strand bias signatures previously reported in antigen-selected V(D)Js (Steele, 2009; Roa et al., 2010; van Oers et al., 2010), which show WA>TW and GYW>WRC biased frequencies, were similarly evident in both WT and Mlh1<sup>-/-</sup> mice (Fig. 1, B and D). To determine whether selection pressures on the V<sub>186.2</sub> coding region concealed hypermutation differences, we compiled unique mutations within the intrinsic Jh2-Jh4 region of splenic B cells from unimmunized mice. As expected, Jh2-Jh4 sequences yielded fewer mutations than V<sub>186.2</sub> sequences (Fig. 1 E), but the overall mutation frequencies and the relative spectrum of mutations in both scenarios were comparable between the mutant and WT mice (Fig. 1, E and F). Fig. 1 F also shows a difference in G:C/A:T ratios between V<sub>186.2</sub> and Jh2-Jh4 regions, which is not completely understood yet but has been connected before to intrinsic differences between antigen and nonantigen selected regions (Steele, 2009).

Other MMR factors, including MutSo, the exonuclease EXO1, the ubiquitylated form of PCNA, and error-prone polymerases such as Polη, have been clearly shown to behave as error-prone factors at the Ig locus, modifying the spectrum of substitutions and introducing mutations at A:T sites within the V region (Chahwan et al., 2011). However, our data here on the catalytic mutant MLH1 expand previous findings that show little or no effect of MLH1 (Kim et al., 1999; Phung et al., 1999) or PMS2 deficiency (Péron et al., 2008; van Oers et al., 2010) in SHM of B cells. The reason why the heterodimer MLH1–PMS2 collaborates with MSH2/MSH6/PCNA/EXO1/Polη outside the V region and is essential in global MMR but seems to be irrelevant for SHM remains unknown. It is likely that there is some mechanism to prevent MutLa from being recruited by the MutSo complex or
becoming active at the V region so as to prevent it signaling for additional downstream elements and reduce the likelihood of DSBs.

**Mlh1GR/GR B cells show reduced efficiencies of CSR**

In contrast to the lack of a significant effect of the loss of MLH1 and of its ATPase activity on SHM, Mlh1GR/GR mice have a substantial decrease in CSR (Schrader et al., 1999; Stavnezer et al., 2010; Eccleston et al., 2011). Because this decrease in CSR could be caused by the absence of the MLH1–PMS2 complex, including its scaffolding functions, or by the loss of the ATPase activity of MLH1, we examined whether the selective inactivation of the ATPase activity of MLH1 would affect CSR. We purified splenic B cells from mutant and WT mice and stimulated them with LPS or with LPS + IL4 to induce ex vivo switching from IgM to IgG3 and IgG1, respectively. After normalization of the efficiencies of switching to WT levels, Mlh1GR/GR mice showed an ~70% reduction in CSR, to all studied isotypes (P < 0.001; Fig. 2, A and B). No appreciable differences in proliferation that might delay CSR were detected between WT and Mlh1GR/GR B cells, as assessed by CFSE dilution histograms (Fig. 2 C).

A similar phenotype has been shown in the absence of PMS2, an essential partner of MLH1 during CSR and postreplicative MMR (Schrader et al., 1999; Ehrenstein et al., 2001; Péron et al., 2008; van Oers et al., 2010). These reduced efficiencies in CSR have been associated with impaired generation and resolution through NHEJ of DSBs, both of which are critical for CSR (Stavnezer et al., 2010). However, little is known about the functional domains of MLH1 and the role, if any, of its distinct catalytic activities during CSR. Based on the deficiency in CSR exhibited here by Mlh1GR/GR B cells, it became plausible to explore the hypothesis that the ATPase activity of MLH1 might play a role in the generation and/or resolution of the DSBs produced during CSR.

The generation of DSBs is decreased in Mlh1GR/GR activated B cells

Because MLH1–PMS2 ATPase activities are thought to be required for the activation of the MutL endonuclease function (Pillon et al., 2010; Polosina and Cupples, 2010) leading to DSBs in SRs (van Oers et al., 2010), we determined whether DSB generation during B cell activation was affected in Mlh1GR/GR mice. To examine whether there was a specific decrease of DSBs at SRs, chromatin immunoprecipitation (ChIP) experiments using anti-53BP1 antibodies were conducted on activated B cells from WT, Mlh1GR/GR, and Mlh1GR/GR mice. 53BP1 has been shown to be essential for CSR and the mediation of DNA end joining (Ramachandran et al., 2010; Bothmer et al., 2011), and therefore the recruitment of 53BP1 to the targeted SRs serves as a reporter for the presence of DSBs. As expected, 53BP1 protein was enriched approximately two- to fourfold on the Sα and Sγ1 region in LPS + IL4–stimulated WT B cells compared with the respective Mlh1GR/GR (Fig. 3 A). We did not find a significant increase on the Sy3 region, which switches infrequently after LPS + IL4 treatment (Stavnezer et al., 2010), suggesting that the observed 53BP1 enrichment was specific. Furthermore, we used Western blots to examine the levels of H2AX that undergo rapid phosphorylation (γ-H2AX) in response to the generation of DSBs in the stimulated B cells (Rogakou et al., 1999). As expected, all cells stimulated with LPS or with LPS + IL4 had considerably higher γ-H2AX signal compared with nonstimulated cells (Fig. 3 B). However, the increase in the overall amount of γ-H2AX was much lower in activated Mlh1GR/GR splenic B cells than in WT cells (Fig. 3 B), also suggesting a smaller accumulation of DSBs in the mutant cells.

Collectively, these data suggest that there is a decrease in the formation of DSBs in the relevant SRs in the MLH1-ATPase mutant B cells that is comparable with that found in mice lacking MLH1. This is consistent with reports that MMR factors are upstream of IgH DSB formation (Schrader et al., 2007; Péron et al., 2008). Our observations further suggest that at least some of the DSBs generated during CSR are orchestrated by MLH1 through its ATPase activity. One possibility is that the ATPase-dependent conformational changes of MLH1 that undergo rapid phosphorylation (γ-H2AX) in response to the generation of DSBs in the stimulated B cells (Rogakou et al., 1999). As expected, all cells stimulated with LPS or with LPS + IL4 had considerably higher γ-H2AX signal compared with nonstimulated cells (Fig. 3 B). However, the increase in the overall amount of γ-H2AX was much lower in activated Mlh1GR/GR splenic B cells than in WT cells (Fig. 3 B), also suggesting a smaller accumulation of DSBs in the mutant cells.

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Microhomology-mediated end joining mechanisms are enhanced in Mlh1<sup>GR/GR</sup> switching B cells

The lack of MMR function and the reduction of DSBs have been attributed to delayed recombination efficiencies and differences in the length of microhomologies detected at the switch junctions (Lieber, 2010; Stavnezer et al., 2010). The use of long microhomologies during CSR has been interpreted to suggest that the alternate pathway of NHEJ (alt-NHEJ) is being used, and so the analysis of S-S junctions has been proven to be very informative of the role of end-joining factors (Yan et al., 2007; Kotnis et al., 2009). To determine the nature of the DSB repair pathway or pathways during CSR in activated Mlh1<sup>GR/GR</sup> B cells, we analyzed recombined switch junctions. We activated control and Mlh1<sup>GR/GR</sup> splenic B cells with LPS for 96 h and then amplified and sequenced Sµ–Sy3 junctions. The distribution of breakpoints within the SRs was similar when distances up to 1 kb from the edges of the Sµ–Sy3 amplicon were explored (Fig. 4 A).

In contrast, comparison of the actual recombination sites between the Sµ and Sy3 regions from mutant B cells to those of controls revealed profound differences. WT sequences contained ~20% direct (blunt) junctions and only a few insertions, with most of the remainder being short microhomology junctions (1.64 bp mean; Fig. 4, B and C). In contrast, only 5% of direct junctions and also few insertions were found in Mlh1<sup>GR/GR</sup> B cells, which showed more junctions with longer microhomologies (5.45 bp mean) than the WT cells (P < 0.0001; Fig. 4, B and C).

Our finding that sequence microhomology at Sµ–Sy3 junctions is greater in Mlh1<sup>GR/GR</sup> switching B cells than in WT suggests a deficiency in canonical NHEJ in MLH1 mutant B cells and provides additional information regarding the role of MLH1 during the DNA end processing of SRs. The use of long microhomologies is unusual in canonical NHEJ at CSR junctions and suggests the participation of alt-NHEJ (Yan et al., 2007; Kotnis et al., 2009). This has been suggested to be the result of additional end processing of DSBs or the impaired recruitment of NHEJ proteins in the absence of MLH1–PMS2 proteins (Stavnezer et al., 2010). In fact, DNA helicases and other DNA repair factors have been shown to be part of the MutLα interactome (Cannavo et al., 2007) and may require the ATPase activity of MutLα for recruitment to the complex. More specifically, these interactome studies have identified EXO1 and DNA-PKcs as partners of MLH1, both of which have been shown to function during CSR in B cells (Stavnezer et al., 2010). EXO1 is not only an exonuclease member of the MMR complex but has also been shown to be involved in DNA end resection downstream of a DSB, whereas DNA-PKcs is a kinase protein integral for the mediation of DSB repair using the canonical NHEJ pathway (Kotnis et al., 2009; Lieber, 2010; Chahwan et al., 2011). During MMR, MutLα plays a role in the termination of EXO1-mediated strand excision by RPA (Genschel and Modrich, 2009). It is possible that the defect in the ATPase activity of MutLα interferes with this activity also during DNA end processing of SRs, which might lead to longer excision tracts and increased use of microhomologies during NHEJ.

Increased mutation frequencies at Sµ–Sy3 junctions in Mlh1<sup>GR/GR</sup> switching B cells

Recombinant junctions usually accumulate mutations at the donor Sµ and acceptor Sy regions (Stavnezer et al., 2010). To test whether the switching deficiencies observed in Mlh1<sup>GR/GR</sup> mice were accompanied by any alteration in SR mutations, we sequenced Sµ–Sy3 junctions from LPS-activated mutant B cells. The comparison with WT sequences showed a significant increase in the overall frequency of mutations accumulated at the Sµ–Sy3 junctions of Mlh1<sup>GR/GR</sup> B cells (P < 0.01; Fig. 4 D). This approximately twofold increase affected all types of mutations and hotspots in Mlh1<sup>GR/GR</sup> B cells.

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that the loss of the endonuclease activity of PMS2 did not affect the preferential use of canonical NHEJ (van Oers et al., 2010), whereas alt-NHEJ with its longer microhomologies appears to be favored in the \textit{Mlh1GR/GR} B cells studied here. Therefore, the MLH1GR mutation does not only result in the loss of activation of the latent endonuclease of PMS2 but also must result in the loss of additional functions of MLH1.

It is becoming clearer that MMR factors can have separate scaffolding and catalytic functions, which could explain the distinct roles of these proteins during MMR processes. In this sense, mouse models representing separation of function mutations have proven to be useful in exploring the multiple roles of MMR protein complexes during error-free DNA repair, error-prone SHM, or DSB-mediated CSR (Chahwan et al., 2011). The role of the endonuclease activity of PMS2 during CSR was separated from the scaffolding functions of MutL\(\alpha\) in the \textit{Pms2EK/EK} mice (van Oers et al., 2010). Here, we suggest that MLH1 has ATPase-dependent functions that influence both the generation and the NHEJ repair of DSBs at the SRs. In contrast, other ATPase-independent functions of MLH1, such as those involved in the apoptotic response to DNA damage, are retained in the mutant \textit{MLH1GR/GR} but not in the null mice (Avdievich et al., 2008). Therefore, some MLH1 functions are independent of its ATPase activity and seem to be irrelevant to CSR because they are not sufficient to enable moderate or normal efficiencies of switching in \textit{Mlh1GR/GR} mice.

**MATERIALS AND METHODS**

**Mice.** The \textit{Mlh1\(^{-/-}\)} mice were generated previously by deleting exon 2 of the \textit{Mlh1} gene (Edelmann et al., 1996). The \textit{Mlh1\(^{GR/GR}\)} mice were generated more recently and carry a missense mutation within exon 2 of the \textit{Mlh1} gene (Avdievich et al., 2008). The G67R mutation slightly reduced...
the levels of the mutant MLH1 protein, but this effect was very modest and did not affect the interaction of MLH1 with its MutLo partner PMS2 (Avdievich et al., 2008). All mouse experiments were approved by the Albert Einstein College of Medicine Animal Use Committee.

Immunizations and SHM assay. To analyze the V_{H}186.2 region, three 8-wk-old Mhl1^{G67R/G67R} mice along with three WT littermates were immunized intraperitoneally with 200 μg NP_{60}-CGG (Biosearch Technologies) in alum (Thermo Fisher Scientific). To analyze the Jh2-Jh4 region, two aged mice (>8 mo) per genotype were used. In both cases, splenic B cells were purified to analyze SHM as previously described (Roa et al., 2010). Alignment and analysis of unique mutations were performed using SeqMan (DNASTAR) and SHMTool (MacCarthy et al., 2009). As consensus sequences, GenBank/EMBL/DDJB no. J00530.1 (nucleotides 224–496) for V_{H}186.2, GenBank no. NT_039551.8 (nucleotides 25620771–25619943) for Jh2-Jh4, and GenBank no. NT_039551.8 (nucleotides 25521839–25521801) for the rearranged IgG1 constant region were used. The presence of significant A>T and WA>TW bias in the WT sequences was used as a quality assessment of mutation datasets (Steele, 2009).

Ex vivo CSR assay. After depletion of T cells by complement-mediated lysis, splenic B cells were obtained from six WT and seven Mhl1^{G67R/G67R} 8-wk-old mice. Purified B cells were plated at a concentration of 0.5 × 10^6 cells/ml and stimulated with either 50 μg/ml LPS (Sigma-Aldrich) or 50 μg/ml LPS and 50 ng/ml rIL-4 (R&D Systems) for 4 d. Simultaneously, purified B cells were preincubated for 2 h at 4°C with Protein A–agarose beads (Roche) to remove free IgG and then incubated overnight at 4°C with 2 µg 53BP1 antibody. Washes and isolation of DNA, as well as quantitative PCR conditions and primers are described previously (Kuang et al., 2009).

Analysis of S_p–S_y3 switch junctions. Genomic DNA was prepared from splenic B cells cultured for 4 d with 50 μg/ml LPS to induce specific switching from IgM to IgG3. Functional S_p–S_y3 DNA was amplified using high-fidelity PfuTurbo DNA polymerase (Agilent Technologies) as previously described (van Oers et al., 2010). Alignment and analysis of sequenced switch junctions were performed using BLAST2seq (NCBI) with the 3 consensus sequences. As consensus sequences, GenBank/EMBL/DDBJ no. J00530.1 (nucleotides 224–496) for V_{H}186.2, GenBank no. NT_039551.8 (nucleotides 25620771–25619943) for Jh2-Jh4, and GenBank no. NT_039551.8 (nucleotides 25521839–25521801) for the rearranged IgG1 constant region were used. The presence of significant A>T and WA>TW bias in the WT sequences was used as a quality assessment of mutation datasets (Steele, 2009).

Western blotting and quantitative ChIP. For Western blot analysis, 1.0 × 10^6 purified splenic B cells was sonicated chromatin from 2.0 × 10^6 purified splenic B cells was prepared according to published protocols (Kuang et al., 2009). In brief, sonicated chromatin from 2.0 × 10^6 purified splenic B cells was preincubated for 2 h at 4°C with Protein A–agarose beads (Roche) to avoid nonspecific binding. After centrifugation at maximum speed at 4°C for 5 min, the supernatant was incubated overnight at 4°C with 2 μg 53BP1 antisera (Novus Biologicals). The mixture was then centrifuged in the preceding step, and chromatin–antibody complexes were collected by incubation with Protein A–agarose beads for 90 min at 4°C. Washes and isolation of DNA, as well as quantitative PCR conditions and primers are described previously (Kuang et al., 2009).

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