Sequential class switching is required for the generation of high affinity IgE antibodies

Huizhong Xiong,1,2 Jayashree Dolpady,1 Matthias Wabl,5 Maria A. Curotto de Lafaille,1 and Juan J. Lafaille1,3,4

1Molecular Pathogenesis Program, Helen L. and Martin S. Kimmel Center for Biology and Medicine at the Skirball Institute of Biomolecular Medicine, 2The Sackler Institute of Graduate Biomedical Sciences, 3Department of Pathology; and 4Department of Medicine; New York University School of Medicine, New York, NY 10016

Abstract

IgE antibodies with high affinity for their antigens can be stably cross-linked at low concentrations by trace amounts of antigen, whereas IgE antibodies with low affinity bind their antigens weakly. In this study, we find that there are two distinct pathways to generate high and low affinity IgE. High affinity IgE is generated through sequential class switching (\(\mu\rightarrow\gamma\rightarrow\epsilon\)) in which an intermediary IgG phase is necessary for the affinity maturation of the IgE response, where the IgE inherits somatic hypermutations and high affinity from the IgG1 phase. In contrast, low affinity IgE is generated through direct class switching (\(\mu\rightarrow\epsilon\)) and is much less mutated. Mice deficient in IgG1 production cannot produce high affinity IgE, even after repeated immunizations. We demonstrate that a small amount of high affinity IgE can cause anaphylaxis and is pathogenic. Low affinity IgE competes with high affinity IgE for binding to Fc\(\varepsilon\) receptors and prevents anaphylaxis and is thus beneficial.

Antibodies belong to five classes (IgM, IgD, IgG, IgE, and IgA) depending on the constant regions of the heavy chain (\(C_H\)) they use. Class switch recombination (CSR) changes the \(C_H\) from \(C_\mu\) to a downstream \(C_H\) (\(C_\gamma, C_\epsilon,\) or \(C_\alpha\)) by DNA recombination and looping-out of the intervening sequences (Selsing, 2006). The irreversible DNA rearrangement in the \(C_H\) region occurs between two switch (S) regions, which lie upstream of each \(C_H\) region (except \(C_\delta\)). CSR is initiated by activation-induced (cytidine) deaminase (AID), which is also essential for the introduction of somatic hypermutations in the variable regions of the BCR (Muramatsu et al., 2000).

IgE is the most stringently regulated secreted Ig, with the lowest serum concentration and the shortest half-life of about 2 d or less (Tada et al., 1975; Stone et al., 2010). In contrast, cell-bound IgE can remain for weeks on the surface of cells bearing Fc\(\varepsilon\) receptors. Mast cells (as well as basophils) are prearmed with IgE through Fc\(\varepsilon\)RI molecules binding to the C region of IgE. Cross-linking of IgE molecules by multivalent antigen results in the activation of mast cells. Upon activation, mast cells degranulate, releasing histamine and other mediators that dilate blood vessels, initiate inflammation, and recruit other effector cells (Kinet, 1999; Kraft and Kinet, 2007; MacGlashan, 2008). The affinity of IgE molecules for their antigens is crucial in determining the fate of IgE-decorated cells. It is possible that mast cells bearing IgE molecules with high affinity for the antigens could be degranulated in the presence of just minute amounts of antigens. In contrast, the high levels of antigen required to cross-link and degranulate mast cells covered by low affinity IgE molecules may not be physiologically achievable. Mast cells are important components of the homeostasis of tissues, and, in such role, they are beneficial. A series of papers have conclusively shown that IgE antibodies are capable of increasing survival of growth factor–deprived mast cells (Asai et al., 2001). There have been no studies comparing the behavior of mast cells covered by IgE antibodies that display low versus high affinity binding.
to a given antigen; instead, affinity correlations were performed by studying strong and weak cross-linking of FcεRI in several alternative ways (Kalesnikoff et al., 2001; Gonzalez-Espinosa et al., 2003; Yamasaki et al., 2004; Kohno et al., 2005; Rivera et al., 2008). These studies showed that a high level of cross-linking of FcεRI receptors results in mast cell degranulation as well as cytokine/chemokine secretion, whereas a low level of cross-linking leads to cytokine and chemokine secretion but no degranulation. However, so far there is no conclusive evidence that the increasing affinity of a given antigen for IgE can turn the function of IgE from beneficial to pathogenic.

At the molecular level, there are two paths to produce IgE antibodies from IgM (Fig. 1 A). One is the direct switching pathway, in which recombination occurs between Sµ and Se; the other is the sequential switching pathway, which is a two-step process observed in both mice and humans. In mice, Sµ first recombines with Sy1, and the hybrid switch region undergoes a second recombination with Se. At the end of sequential switching, Sy1 remnants can be found in some of the rearranged Sµ-Sc junctions (Fig. 1 A). It has been reported that in mice up to 30% of the recombined regions contain Sy1 remnants (Siebenkotten et al., 1992; Mandler et al., 1993). In human cells, the switch to ε in a lymphoid cell line was preceded by an Sµ-Sy recombination (Mills et al., 1992). In B lymphocyte cultures from mice harboring a deletion in the IgM switch region, IgE+ cells could be found at nearly normal levels (70%), and they were derived from CSR events involving Sy1 (Zhang et al., 2010).

We previously reported that ε sterile transcripts and ε switched transcripts, which are generated before and after the switch to Ce, respectively, were abundantly found in IgG1+ cells. Furthermore, purified IgG1+ cells could switch to IgE in vitro and in vivo. In addition, affinity-enhancing somatic mutations found in the IgG1+ cells were also found in IgE+ B cells, although with some delay (Erazo et al., 2007). We proposed that sequential class switching through an IgG1 intermediate was a way to generate high affinity IgE antibodies. The IgE cells were absent from germinal centers (GCs), and we proposed that they inherited somatic mutations and affinity maturation from the preceding IgG1 phase.

Despite prior studies by multiple laboratories (Yoshida et al. [1990], Erazo et al. [2007], and references therein), it is unclear whether sequential switching to IgE through an IgG1 intermediate is an obligatory step to generate high affinity IgE antibodies. In this study, we demonstrate that, indeed, high affinity IgE antibodies in the mouse are generated through an IgG1 intermediate.

RESULTS

A simple assay to quantify sequential DNA switching in IgE+ cells

Because our hypothesis is that high affinity IgE antibodies are generated through an IgG intermediate, we developed a PCR assay to quantify Sy1 remnants in the Sµ-Sc junctions of IgE+ cells. As schematized in Fig. 1 A, the finding of Sy1 remnants is always indicative of sequential switching. In our assay, after we ran a PCR over the Sµ-Sc junctions, the products were subjected to quantitative PCR (qPCR) on the Sy1 repeats.

We first validated the method using four IgE hybridomas and five subclones from one of them. PCR was performed using primers located at 5’ of Sµ and 3’ of Sc. The size of the PCR-amplified Sµ-Sc junctions was between 3 and 7 kb (Fig. 1 B). There was a dominant band for each hybridoma. The four hybridomas had different sized bands, and the five subclones had the same sized band as the original hybridoma (Fig. 1 B). 24 minipreps ran from cloned Sµ-Sc junctions to a given antigen; instead, affinity correlations were performed by studying strong and weak cross-linking of FcεRI in several alternative ways (Kalesnikoff et al., 2001; Gonzalez-Espinosa et al., 2003; Yamasaki et al., 2004; Kohno et al., 2005; Rivera et al., 2008). These studies showed that a high level of cross-linking of FcεRI receptors results in mast cell degranulation as well as cytokine/chemokine secretion, whereas a low level of cross-linking leads to cytokine and chemokine secretion but no degranulation. However, so far there is no conclusive evidence that the increasing affinity of a given antigen for IgE can turn the function of IgE from beneficial to pathogenic.

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of each hybridoma indicated that all inserts were of the same size, reassuring us that no major recombination events took place during the PCR and cloning steps.

The second PCR step was a quantitative real-time PCR on Sy1 to target the characteristic 49-mer Sy1 consensus sequences (Mowatt and Dunnick, 1986). Sy1 contains a long (∼11.5 kb) region characterized by >120 imperfectly repeated 49-mer sequences. Actual consensus Sy1 sequence repeats perfectly only nine times, whereas the other repeats have small but complicated changes like overlaps and truncations. qPCR detected the Sy1 remnant in one of the four hybridomas and all of its subclones. The other three hybridomas were negative (Fig. 1 C). Both positive and negative qPCR results were confirmed by DNA sequencing. Because it was apparent that the IgE-producing hybridomas only retained one Spl-Se allele, the Sy1 qPCR yielded a quantitative signal of either >10,000 or 0, i.e., Sy1 was either present or absent.

To evaluate the frequency of sequentially switched IgE cells within a population of cells during an immune response, we defined an Sy1 ratio, which is the number of Sy1+ junctions among the total number of Spl-Se junctions. Even though not all sequentially switched cells had Sy1 remnants (Fig. 1 A), as a population, the frequency of Sy1+ cells still provided a comparative measure of the extent of sequential switching. The Sy1 ratio in hybridomas was either 0 or 1, and in an vivo IgE response will have a ratio anywhere between 0 and 1.

Increased sequential switching to IgE correlates with increased IgE affinity for antigen

For the present experiments on affinity maturation of the IgE response, we used two experimental systems. The TBmc system (Curotto de Lafaille et al., 2001) consists of heavy and light chain influenza hemagglutinin (HA)-specific Ig knockin mice crossed with OVA-specific TCR transgenic mice (D011.10) and RAG1 KO mice, all on a BALB/c genetic background. The HA-specific knockin Ig genes were derived from the high affinity BCR 17-9 hybridoma (Schulze-Gahmen et al., 1988), and as a consequence, the antibodies are intrinsically of high affinity for HA. Because of both the absence of natural regulatory T cells (Treg cells) and the BALB/c background, these mice display a high HA-specific IgE response after one immunization with the cross-linked antigen OVA-HA. There is no increase in affinity-enhancing mutations of the anti-HA response after OVA-HA immunization (Erazo et al., 2007). To study affinity maturation, we used a single mutant HA peptide, PEP1, in which a phenylalanine residue replaced one of the tyrosine residues in HA. Binding of the endogenous anti-HA antibodies to PEP is 1,000 times weaker than to HA (Pimilla et al., 1993). In TB monoclonal (TBmc) mice, immunization with OVA-PEP1 allows the tracking of affinity maturation both by the appearance of PEP1-binding B cells (undetectable if mice are not immunized with PEP1) and PEP1 affinity-enhancing mutations in the Ig genes.

The second experimental system is a well-studied system of immunization of WT mice with the hapten Nitrophenyl (NP) conjugated with carrier proteins. The results using both experimental systems were in agreement, as shown in the following paragraphs.

Immunizing TBmc multiple times at 3-wk intervals with OVA-PEP1 enabled us to observe the gradual increase of IgE affinity to PEP1 (Fig. 2 A). After each immunization, we determined the serum titer against PEP1 and sorted IgE+IgG1−Fashigh cells from the spleen (not depicted). As the number of immunizations of TBmc mice with OVA-PEP1 increased from one time to four times, the PEP1-specific IgE serum titer raised from 50 to 4,000 (Fig. 2 A). IgE+IgG1−Faslow cells were also sorted but did not contain amphilifiable switch regions (not depicted); they were positive for FceRI (not depicted), indicating that they were IgE-coated cells. As we reported previously, a mild acid treatment is sufficient to dislodge IgE molecules from CD23, the low affinity IgE receptor, but not from the high affinity FceRI (Erazo et al., 2007).

DNA was extracted to perform Sp1-Se PCR. We observed a smear of different sizes (not depicted). Interestingly, as the number of immunizations increased, we observed dominant Sp1-Se bands, which was not seen in other experimental models such as the NP-immunized human metallogenprotein H ε (hMT) mice and the anti-IgD–treated WT mice (see below). We applied the aforementioned single colony qPCR assay to determine the Sy1 ratio. As we increased the number of immunizations, the Sy1 ratio went up from 0.03...
to 0.18 and then to 0.61 (Fig. 2, A and B), indicating that sequentially switched IgE was gradually enriched as the IgE affinity increased. To validate the sensitivity and specificity of Sy1 qPCR, we randomly chose 40 Sp–Se junctions for sequencing and compared the sequencing data with qPCR results (Table S1). The Sy1 qPCR assay was 100% sensitive and 100% specific. The minimal Sy1 length that was detected was 108 bp. Many Sy1 remnants had incomplete consensus sequences, indicating that the qPCR assay could deal with small Sy1 fragments with imperfect 49-mer repeats.

More switch region mutations accumulate in junctions undergoing sequential switching than direct switching

It has been reported that somatic hypermutations in both VDJ regions and switch regions are driven by the same AID machinery (Petersen et al., 2001; Goodman et al., 2007; Maul and Gearhart, 2010). The mutation levels in switch regions can therefore be used to evaluate AID activity. To compare the level of AID activity during sequential and direct switching to IgE, we analyzed the mutations in Sp–Se junctions of PEP1-immunized TBmc mice. We found that >95% of the mutations in switch regions accumulated in a 1,000-bp region. Conversely, two thirds of Sy1+ junctions had >10 mutations, whereas only 19% of the Sy1− junctions had >10 mutations. 67% of the Sy1+ clones (sequentially switched) had >10 mutations, whereas only 19% of the Sy1− clones had >10 mutations. Conversely, two thirds of Sy1− junctions had zero to four mutations, whereas only 17% of the Sy1+ junctions had such low mutation levels (Fig. 3 B). Over 70% of the mutations had the AID target motif WGCW (Goodman et al., 2007; Han et al., 2011). The difference in the number of mutations between Sy1+ and Sy1− junctions increased as the number of immunizations augmented (5.7 in the Sp–Se direct switching junctions vs. 9.8 in the Sp–Sy1 sequential switching junctions, after four immunizations; P = 0.04; Fig. 3 C). Furthermore, the distribution of the number of mutations in Sy1− junctions was discontinuous, with the appearance of a bimodal distribution (Fig. 3 B, left). It is possible that the highly mutated Sy1− sequences represent sequential switching events in which the Sy1 remnants were absent (Fig. 1 A). However, these mutations were particularly enriched in intra-switch recombination events, as four out of six Sy1− sequences with >20 point mutations had intra-switch recombination in Sp; therefore, we cannot be sure that the highly mutated Sy1− sequences represent sequential switching events.

The lower mutation frequency in Se compared with Sp could be explained, at least partially, by the lower number and density of AID target motifs in the Se region; the preferred AID motif WGCW is present 415 times in Sp (density of 134.7/kb), 120 times in Se (density of 41.9/kb), and 401 times in Sy1 (density of 34.9/kb). Collectively, these results suggest that sequential switching correlates with a higher AID activity or two separate AID events from the two-step recombination process.

IgG1-deficient hMT mice produce normal levels of IgE but have impaired affinity maturation of the IgE response

If, as the correlations show, high affinity IgE antibodies are generated through sequential switching through an IgG1 intermediate, then IgG1 deficiency should have an impact on the affinity maturation of the IgE response. hMT mice have a replacement at the IgY1 region (5′ of Sy1) by an hMT promoter, which doesn’t respond to a variety of inductive stimuli or get involved in any recombination process. As a result, these hMT-deficient mice cannot generate sterile γ1 transcripts and therefore lack IgG1+ cells and IgG1 antibodies (Lorenz et al., 1995). It has been shown that these mice can still have

Figure 3. Sequentially switched Sp–Se junctions harbor a large number of mutations. (A) TBmc mice were immunized two or four times with OVA-PEP1. The DNA was extracted, and the Sp–Se junctions were amplified. 33 clones from each group were sequenced. Diamonds show the approximate position of the mutations at the junction. (B) Sy1− and Sy1+ junctions from mice immunized four times were sequenced and analyzed using pies (left) to show the distribution of point mutation frequency and columns (right) to show the overall mutation frequency. Error bars indicate SEM. (C) Mean number of point mutations at the junctions after two and four immunizations. Sy1+ junctions were split into two junctions, Sp–Sy1 and Sy1–Se, and counted separately.
normal levels of total IgE (Jung et al., 1994), indicating that IgE does not necessarily derive from sequential switching. However, we propose that the IgE affinity maturation in hMT mice is impaired.

We immunized hMT and their WT littermates with NP19KLH four times at 3-wk intervals and collected serum to assess the NP-specific antibody responses. Plates were coated with NP4BSA and NP23BSA. NP4 is a highly conjugated molecule that binds both low and high affinity antibodies, whereas NP23 has a low degree of cross-linking and binds only high affinity antibodies. A higher NP4/NP23 ratio indicates a higher affinity to NP (Herzenberg et al., 1980; Allen et al., 1988; Smith et al., 1994; Chen et al., 2000; Shimizu et al., 2003; O’Connor et al., 2006). IgE antibodies from hMT mice had a much lower NP4/NP23 ratio than IgE antibodies from WT mice (Fig. 4 A, left). Absolute absorbance of NP23-bound IgE was similar between hMT and WT mice, but the binding of NP4 was much weaker in hMT IgE than in WT IgE (Fig. 4 A, right). As expected, total IgE levels were similar in hMT and WT (Fig. 4 B), indicating that the low NP4/NP23 ratio in hMT mice was not caused by the deficiency of IgE production but rather by the affinity maturation of the IgE response. We applied the same ELISA technique to study the IgG2a and IgG1 responses in the same mice. IgG2a constitutes a good isotype to identify whether any nonspecific effect on IgE could be taking place. hMT mice did not have any IgG1, whereas IgG1 of high affinity was readily detected in WT mice (Fig. 4 C, left). The affinity maturation of the IgG2a response was the same in hMT and in WT mice (Fig. 4 C, right), indicating that IgG2a affinity maturation occurs independent of γ1 sequential switching.

We sought to demonstrate that the high affinity NP-specific IgE response was accompanied by molecular remnants of sequential switching through IgG1. We sorted IgE+ cells and ran the Sγ1-Sc PCR in the same way we did on TBmc mice. In contrast to the TBmc situation, dominant Sc-Sc bands were not observed after four immunizations in hMT mice or the WT littermates. We then calculated the Sγ1 ratio based on 100 cloned Sc-Sc junctions derived from four WT mice and a similar number of hMT mice. The Sγ1 qPCR assay showed that, depending on the mice, 10–30% of WT Sc-Sc junctions had Sc1 remnants (Fig. 4 D). Interestingly, Sc-Sc junctions from hMT mice not only lacked Sc1 remnants, which was expected, but contained no other intervening S region (not depicted). Thus, in hMT mice, IgE antibodies are exclusively generated through direct switching.

**Somatic mutations that enhance affinity to NP are not present in IgE genes from IgG1-deficient mice**

In the C57BL/6 (B6) mouse strain, the repertoire of B cells responding to NP has been extensively studied (Cumano and Rajewsky, 1986; Furukawa et al., 1999; Weiser et al., 2011). It was determined that V18186.2 was preferentially used in the anti-NP response, paired with IgL1.

The maturation of the NP-specific response includes two different mutation paths: in one of them, the first junctional residue in the CDR3 is Y99. In those B cells, affinity to NP is greatly increased by a W33L mutation. Additional mutations do not further enhance the affinity for NP in the Y99-type cells. The second path involves heavy chains with a junctional CDR3 residue G99, encoded by N nucleotides. In this case, high affinity is achieved through the accumulation of several mutations, mainly K59R, G57D, S66N, and other contributors, and allows for much higher affinities (Furukawa et al., 1999; Weiser et al., 2011). However, there is a timing trade-off: it has been determined that the Y99/W33L type of cells appears relatively early during the anti-NP response, whereas the G99/K59R type of cells appears late, but the latter takes over upon secondary immunization as it accumulates additional affinity-enhancing mutations (Jacob et al., 1993; Takahashi et al., 2001; Murakami et al., 2010; Nishimura et al., 2011).

In contrast to the B6 strain, knowledge on the molecular basis of affinity maturation of the NP response in BALB/c mice is scanty. Because of the fact that B6 mice make poor IgE responses, our work was performed in hMT mice (and littermates) backcrossed.

![Figure 4](https://example.com/figure4.png)

**Figure 4.** IgE affinity is impaired in IgG1-deficient mice.

(A) WT and hMT mice were immunized with NP19KLH four times. Serum was analyzed by ELISA with NP4- and NP23-coated plates. (left) NP4/NP23 ratio. (right) Absolute absorbance value. Error bars indicate SEM. Each dot represents one mouse. (B) Same sera used in A were subjected to total IgE ELISA. (C) IgG1 and IgG2a ELISA with NP4/NP23 plates. (D) Sγ1 ratio in WT and hMT mice using the Sγ1 qPCR assay. Each dot represents one animal (four animals per group). The number of Sγ1+ junctions in each mouse is shown in the right panel, divided by the total number of Sc-Sc sequences. (A–D) Horizontal bars indicate the mean.
onto the BALB/c background. Our characterization of the NP-specific response in mice of BALB/c background indicated that the preferred Vιι gene was homologous to Vι1.186.2 and that the ιgL chain also dominated the NP response in BALB/c mice (not depicted). We immunized BALB/c background mice with NP0KLH, sorted IgG1+ cells that did or did not bind NP-PE (not depicted), and sequenced the VDJ segments. Interestingly, the Y99/W33L path was not found on >300 DNA sequences from NP+ B cells, at any time point ranging from day 7 to day 80 after immunization (Fig. 5 A). Instead, acquisition of NP binding was accompanied by an increase in the frequency of the K59R mutation, which in approximately half of the cases had G57D and/or S66T/N mutations. G99 was found only in some of these antibodies, but all the junctions contained N nucleotides (Table S2). Thus, the pathway that confers the highest NP binding affinity in B6 mice is the one that operates in BALB/c mice.

With this knowledge in hand, we sought to analyze the affinity-enhancing mutations in IgE, IgG2a, and IgG1 in WT and hMT mice immunized with NP0KLH. We amplified Vι1.1 sequences (homologous to the B6 Vι1.186.2) in IgE+, IgG2a+, and IgG1+ cells. We then studied the mutations in 96 DNA sequences for IgE, 82 sequences for IgG2a, and 62 sequences for IgG1 (the latter only from WT mice). The IgE VDJ repertoire in hMT mice had significantly fewer mutations than WT mice (Fig. 5, B and C). Only 8% of the IgE cDNAs from hMT mice had 20–24 nucleotide mutations, and no cDNA had 25–29 mutations. In contrast, in WT mice, 13% and 17% of the IgE VDJ regions contained 20–24 and 25–29 mutations, respectively. The percentage of sequences with high frequency of mutations in IgE genes from WT mice was slightly lower than that of IgG1 genes from the same mice (Fig. 5 B), consistent with the observation in TBmc anti-PEP1 response (Erazo et al., 2007). In contrast with IgE, the frequency of mutations in IgG2a VDJ genes was similar between hMT and WT mice (Fig. 5 B). Note that the reduction in the frequency of mutations is less pronounced than the reduction in affinity, indicating that the selection of high affinity mutants is more affected than the overall level of mutations.

Next we analyzed the typical mutations that confer higher binding affinity to NP in mice of BALB/c background. In IgE genes, the K59R mutation was found only in WT IgE but not hMT IgE (Fig. 5 A). In contrast, the K59R mutation had a similar frequency in IgG2a from both WT and hMT mice. About half of the K59R IgE clones in WT mice had the additional G57D and/or S66T/N mutations (Fig. 5 A).

Finally, we also studied the J and D segment usage in these clones. In C57BL/6 mice, the early NP-specific B cells (Y99 type) have highly enhanced usage of the Dfl16.1-Jι1 segments, and even the late-arising NP-specific B cells (G99 type) predominantly use the Dfl16.1 segment (Furukawa et al., 1999). In the BALB/c background mice, there was no strict preference for the usage of certain JH (not depicted) or DH (Table S2) segments; however, there was a preference of JH3 in NP +IgG1+ cells, whereas NP −IgG1+ cells used more the JH4 segment (P = 0.0001; not depicted). Thus, JH3 usage correlates with higher affinity for NP in the BALB/c background. In accordance with this, the JH usage in total IgE+ cells from hMT mice showed preference for JH4 over JH3, whereas the higher affinity IgE+ cells from WT mice showed preference for JH3 over JH4 (P = 0.0001; not depicted). In summary, these data strongly support the model in which high affinity IgE is generated through sequential switching involving Sy1.

Figure 5. Mutations in the VDJ region of IgG1-deficient mice and their WT littermates. (A) Total splenocytes from the same hMT and WT littermate mice used in Fig. 4 A were prepared for cDNA. PCR was performed using a common Vι1.1 forward primer and Cι and Cy2a reverse primers. NPhigh and NPlow IgG1+ cells were sorted by staining with NP-PE. cDNA was prepared, and Vι1.1- Cy1 PCR was performed. After sequencing, the NP affinity-enhancing mutations were tabulated. (B and C) Total nucleotide mutation analysis of the same sequences described in A (B) and the analysis of total mutation frequency (C). Error bars indicate SEM.

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Antigen-driven high affinity but not low affinity IgE causes anaphylaxis

The differential pathological function of IgE antibodies with high versus low affinity for their antigens has been proposed but not fully explored. Anaphylaxis is a life-threatening reaction caused by antigen-induced cross-linking of IgE molecules bound to mast cells. In this study, we used a local passive cutaneous anaphylaxis (PCA) assay in the ears to quantify the effect of high and low affinity IgE antibodies in vivo. The amount of IgE, from IgG- and IgA-depleted serum, was normalized in all cases. First, mice were injected in the ears with anti-HA IgE, followed by cross-linked HA (OVA-HA) together with Evans blue by systemic route. This combination induced strong extravasation of Evans blue into the ears (Fig. 7 A, left). However, there was virtually no Evans blue leakage when injection of the same anti-HA IgE antibodies was followed by low affinity antigen OVA-PEP1 (Fig. 7 A, middle left). As indicated above (Erazo et al., 2007), HA-specific antibodies are \( \times 1,000 \) fold less reactive to OVA-PEP1 than to OVA-HA. When IgE affinity to PEP1 increased through affinity maturation caused by repeated immunizations, these IgE antibodies displayed the capability to cause Evans blue extravasation upon OVA-PEP1 challenge (Fig. 7 A, middle right). As expected, unconjugated monovalent soluble PEP1 could not cross-link anti-PEP1 IgE and did not cause the PCA (Fig. 7 A, right). The results were confirmed by spectrophotometric determination of the leaked dye upon cutting and dissolving the affected and control ears (Fig. 7 B). Next, we titrated the amount of injected IgE in the ears. In serial dilution experiments, very diluted anti-HA IgE (1:100 dilution) caused a strong PCA reaction by OVA-HA, and the same happened in the case of anti-PEP1 IgE with OVA-PEP1. However, in the presence of OVA-PEP1, anti-HA IgE did not cause anaphylaxis, even when injected undiluted (Fig. 7 C). To illustrate the importance of affinity maturation of IgE in anaphylaxis, we studied IgE obtained after
Sequential switching produces high affinity IgE | Xiong et al.

IgG1 phase. Our data showed that affinity-enhancing mutations in VDJ segments from IgE cDNAs but not from the IgG2a cDNA, whose C region lies next to Cε, were greatly reduced in IgG1-deficient mice. Thus, affinity maturation of IgE can occur through a GC IgG1 phase. Whether these IgG1+ cells that switch to IgE+ cells are immediate post-GC cells, memory cells, or cells committed to become plasmas blasts remains the subject of future studies. We previously showed that GC IgG1 cells can express sterile as well as switched ε transcripts, but double expressing IgG1+/IgE+ cells produced in homozygous knockin TBmc mice do not have a GC phenotype (Erazo et al., 2007).

It has been known for a long time that the dose and frequency of immunizations have an impact on the amount and ratio of IgE and IgG1 antibodies (Vaz et al., 1971). It is likely that at higher doses, Treg cells can be recruited to the response and suppress the IgE response (Curotto de Lafaille et al., 2001).

We previously reported that IgE cells were absent from GCs during Nippostrongylus brasiliensis infection as well as during immunization of TBmc with cross-linked OVA-HA (Erazo et al., 2007). In fact, the evidence that GCs are not favorable structures for IgE+ cells is mounting. Bcl6-deficient zero, one, two, and four immunizations with OVA-PEP1, using normalized amount of total IgE. Only IgE from mice immunized four times was capable of causing the full anaphylactic reaction (Fig. 7 D), which was confirmed by spectrophotometric measurement (Fig. 7 E).

DISCUSSION
Sequential switching $\mu \rightarrow \gamma \rightarrow \epsilon$ was discovered >20 yr ago (Yoshida et al., 1990). However, previous studies did not investigate whether $\gamma$ remnants represented transient markers of an instant sequential switching event or there was a cellular

![Figure 7. Anaphylaxis caused by IgE of high affinity to its antigen can be competed by IgE of low affinity for the same antigen. TBmc mice were immunized with OVA-HA or OVA-PEP1 two times, and sera were collected and depleted of IgG and IgA. IgE concentration was measured by ELISA. 25 ng of total IgE was injected intradermally into the ear of recipient BALB/c mice. 24 h later, 50 µg OVA-HA, OVA-PEP1, or unconjugated PEP1 was injected intravenously together with 1% Evans blue. (A and B) 30 min later, mice were sacrificed, and the extent of Evans blue leakage was determined either visually (A) or spectrophotometrically (B; three mice per group). (C) Before injecting into the ears, sera were diluted as indicated. (D and E) Similar PCA assay was performed using sera from TBmc mice immunized with OVA-PEP1 one, two, or four times. The PEP1-specific IgE titer is shown above the photos. A total of 25 ng IgE was injected per ear. 24 h later, recipient BALB/c mice were injected intravenously with OVA-PEP1 and Evans blue. The ears are shown in D, and Evans blue was measured spectrophotometrically in E. (F) 6 ng of high affinity anti-PEP1 IgE was mixed with 100 times more PBS, anti-HA IgE, IgG1, or IgM to perform the PCA assay mediated by OVA-PEP1. The extent of Evans blue leakage was determined visually. (G) Quantification of Evans blue leakage by spectrophotometry in the ears injected with 1, 20, 100, and 500 times more IgM, IgG, or low affinity IgE (three mice each group). (B, C, E, and G) Error bars indicate SEM.](https://jem.rupress.org/content/jem/99/2/360.full.pdf)
mice have severely impaired GC formation and no affinity maturation, but they harbor an increased number of IgE+ cells (Ye et al., 1997). Similarly, deficiency of Dock8 (dedicator of cytokinesis 8) causes unstable GCs in mice (Randall et al., 2009) and hyper IgE syndrome in humans (Engelhardt et al., 2009; Zhang et al., 2009). IL-21 is a critical cytokine produced by Tfh cells; similarly to Bcl6 and Dock8, IL21R-deficient mice have higher IgE and lower IgG1 level (Ozaki et al., 2002). Thus, the incompatibility between IgE and GCs can be explained by our sequential switching model, in which IgG1+ intermediate cells undergo the GC phase.

As the number of immunizations increased, the frequency of Spu–Se junctions that contain Sy1 remnants increased, in parallel with an increase in IgE affinity. It is remarkable that during the course of immune responses we did not see S remnants other than Sy1 in Spu–Se junctions. The reason for the strong Sy1 dominance is that sterile transcripts from Sy1 and Ig promoters share some of the triggers, such as IL-4/STAT6, which could turn on Sy1 and Se in the same cell, whereas other sterile transcripts such as Ig2a or Ig2b would remain silent. If this is correct, in other IgE responses in which IgG1 is not activated, we may possibly find remnants other than Sy1 in Spu–Se junctions.

We did not find evidence of the contribution of the memory IgM+ B cells to the high affinity IgE response (Dogan et al., 2009; Tomayko et al., 2010; Pape et al., 2011), as we found almost no affinity maturation in the absence of IgG1. However, it is possible that memory IgM+ B cells are prone to undergo switching to IgE via IgG1 rather than through the direct switching route.

In B6 mice, the high affinity to NP first comes from a combination of Vh186.2 and IgL1. Of the two paths that further increase Vh186.2 affinity for NP, the first path (Y99/W33L) does not have N nucleotides in the VD junctions, whereas the second path (G99/K59R) uses N nucleotides. In BALB/c mice, we found NP-binding cells were also highly enriched in IgLa, and the K59R, with other affinity-enhancing mutations were found. Interestingly, there were many N nucleotides in these junctions, even though they did not necessarily encode G99. These observations show that the mechanism used by BALB/c mice to produce high affinity anti-NP IgE is very similar to the second path in B6 mice, which can be time consuming, but end up with a higher affinity than the first path.

IgE antibodies are spontaneously increased in mice with T cell immunodeficiencies such as nu/nu, Cd4−/−, MHC class II−/−; these IgEs have been referred to as natural IgE antibodies (McCoy et al., 2006). Like the conventional IgE antibodies, natural IgE antibodies are dependent on T cells, CD40–CD40L interactions, and IL-4; however, organized lymphoid structures such as GC are not required, and natural IgE antibodies do not accumulate affinity-enhancing somatic hypermutations (McCoy et al., 2006). Importantly, lymphopenia-induced proliferation in T cell–deficient mice induces T cells that develop predominantly along the Th2 pathway (Milver et al., 2007). The high IgE found in Lgα136f knockin mice could have a similar genesis (Aguado et al., 2002). It thus appears that during some T cell lymphopenic disorders, IgE switching resembles much of what we showed here for directly μ→ɛ switched antibodies. A nonexclusive alternative is that natural IgE antibodies may be the result of switching to IgE at the immature B cell developmental stage. It has recently been shown that immature B cells in bone marrow and spleen switch to IgE in a direct μ→ɛ CSR, whereas the mature B cells have a propensity to switch via an IgG1 intermediate (Wesemann et al., 2011). In contrast to natural IgE generation, conventional IgE responses, which involve affinity maturation, are generated through a different mechanism, involving a key IgG phase.

Natural IgE antibodies may be involved in conditions such as hyper IgE recurrent infection syndrome (HIES), Wiskott–Aldrich syndrome, Omenn syndrome, and DiGeorge syndrome. The combination of immunodeficiency and Th2 bias sometimes results in the production of extremely high levels of IgE (50,000 U/ml) without, however, allergic symptoms such as rhinitis, asthma, urticaria, and anaphylaxis (Grimbacher et al., 2005; Ozcan et al., 2008; Minegishi, 2009; Freeman and Holland, 2010). If all these IgEs had undergone affinity maturation to bind environmental antigens with high affinity, these patients would have severe allergic reactions upon minimal exposure to antigen. Thus, it is likely that hyper IgE responses in HIES patients are of low affinity to environmental antigens and are directly switched μ→ɛ, like natural IgE responses. Compared with HIES patients, children with allergic asthma have lower levels of IgE; however, these IgEs showed hypermutations and affinity maturation (Kerzel et al., 2010).

IgE responses have traditionally been considered important in the clearance of some helminthes, which induce a massive IgE production caused by very efficient Th2 differentiation induced by parasite components. Interestingly, although some of the IgE antibodies are specific for the helminthes (Jarrett and Haig, 1976; Pochanke et al., 2007), the vast majority of the IgE antibodies do not react to the parasites. Evolutionarily, for the parasite, it could result in a delayed rejection, as low affinity IgE competes with parasite-specific IgE for FcεRI receptors and cross-linking of host effector cells would be perturbed (Pritchard, 1993). For the host, helminth nonspecific IgE binding to mast cells would reduce immunopathology and prevent serious systemic allergic reactions.

It has been long known that recombinant Fc fragments and synthetic peptides from Ce can inhibit antigen-specific IgE reactions when present at 200- or 1,000-fold excess, respectively (Geha et al., 1985; Burt and Stanworth, 1987; Nio et al., 1992). We observed inhibition of high affinity IgE-mediated anaphylaxis by the addition of intact IgE of low affinity for the antigen, although complete inhibition of anaphylaxis required large molar excess. It clearly demonstrates the beneficial and pathogenic functions of low and high affinity IgE, respectively.

Besides the effect on basophil and mast cell degranulation, IgE antibodies have also been shown to facilitate antigen
presentation, an effect which is antigen-specific. Thus, high affinity recognition of antigens by IgE could also increase T cell proliferation, as was described both in mice (Oshiba et al., 1997) and humans (Turcanu et al., 2010).

There are large health implications of the affinity maturation of IgE antibodies. By describing a distinct mechanism for the generation of the high affinity IgE response, our study makes possible the detection of high affinity responses even when triggering antigens are unknown, using Syremnants in switch regions as indicators of affinity maturation. The existence of a distinct mechanism also allows the design of therapeutic interventions specifically targeted to affect sequential generation of IgE cells, an intervention which would preserve the positive effects of low affinity IgE antibodies on tissue homeostasis.

**MATERIALS AND METHODS**

Mice, immunization, and injection. TBmc mice were previously described (Curotto de Lafaille et al., 2001). Heterozygous heavy and light chain knockin mice were used. Mice were immunized by intraperitoneal injection of 100 µg OVA-HA (YPYDVPDYASLRS) or with 100 µg OVA-PEP1 (YPYDVPDYASLRS) in alum. hMT mice (Lorenz et al., 1995) were provided by C. Jacob (University of Southern California, Los Angeles, CA). Mice were backcrossed to BALB/c genetic background and immunized by intraperitoneal injection of 50 µg NP, KLH (purchased from Biosearch) in alum. BALB/c mice were purchased from the Jackson Laboratory. Mice were injected intraperitoneally with 200 µl of goat anti-mouse IgD serum (eBioscience). All procedures involving mice were approved by New York University’s Institutional and Animal Care Use Committee.

**Hybridomas.** IgE-producing hybridomas (Böttcher et al., 1980; Rudolph et al., 1981) were grown in RPMI medium supplemented with FBS, Hepes, t-glutamine, β-mercaptoethanol, penicillin, and streptomycin.

**ELISA.** Total and antigen-specific IgE and IgG1 antibodies were quantified by ELISA as described previously (Curotto de Lafaille et al., 2001). To measure antigen-specific IgE in serum, IgG and IgA were depleted by incubating with protein G-Sepharose (GE Healthcare) overnight at 4°C. The plate was coated with 2 µg/ml PEP1 for anti-PEP1 ELISA or with 10 µg/ml NP1BSA and 10 µg/ml NP2BSA separately for anti-NP ELISA. Absorbance was detected by spectrometer at 450 nm. Serum titer was defined as the lowest dilution that rendered an optical density value (OD450) >0.1 after background subtraction.

**Flow cytometry.** Before staining for FACS analysis, single-cell suspensions from spleen were treated for 1 min with ice-cold acid buffer (0.085 M NaCl, 0.005 M KCl, 0.01 M EDTA, and 0.05 M NaAcetate, pH 4) to remove extrinsic IgE antibodies noncovalently bound to CD23. The samples were then neutralized with a large volume of cell culture medium and washed twice before staining. The following reagents were used: FITC-anti-IgG1 (A85-1), biotin-anti-IgE (R35-72), PerCP–anti-B220 (RA3-6B2), PE–anti-Fas (Jo2), PE-Cy7–anti-Fas (Jo2), NP-PE (Biosearch), and APC–streptavidin (BD).

**Sy1 qPCR.** Cells were sorted by a MoFlo cytometer. DNA was extracted from the cells with lysin buffer where proteinase K at 50 µg/ml was freshly added. The following primers were used to amplify Sµ-Sε junctions: SµF, 5′-TTTGGGGAAGGAAATATTTAA-3′; and SεR, 5′-TAGGCT-GTGGTCTAGATG-3′. Amplified junctions were cloned into TA vector (Invitrogen) and miniprepped. qPCR was performed with the following Sy1 primers:Sy1F, 5′-AGCCAGGACAGGTGGAAT-3′; and Sy1R, 5′-CTGCTCTGCTTGGTCA-3′.

**DNA sequence analysis.** For VDJ sequencing, total RNA extraction and cDNA synthesis were performed by standard procedures. PCR reactions were performed with high fidelity Ex Taq polymerase (TaKaRa Bio Inc.) and the common forward primer VH1.1 in combination with primers specific for IgE, IgG1, or IgG2a constant regions (Erazo et al., 2007). For switch region sequencing, PCR–amplified junctions were cloned into TA vector and miniprepped. All the sequences were performed by Macrogen Corp. Point mutations in Syl regions with length of >1,000 bp were counted in the first half and second half of Syl separately and were included in the overall mutation frequencies of the Sµ–Syl junction and the Syl–Se junction, respectively.

**PCA assay.** 10–20 µl IgE-containing serum was injected intradermally into the ears of BALB/c mice. 24 h later, mice were injected intraperitoneally with antigen and 1% Evans blue. 30 min later, mice were sacrificed. The ears were cut and digested in protease K–containing lys buffer. Supernatant was subjected to spectrometry at 650 nm to quantify Evans blue.

**Statistical analysis.** The two-tailed χ2 test was performed to JH preference.

**Online supplemental material.** Table S1 shows that the sequencing results validate the sensitivity and specificity of the Syl qPCR assay, which is related to Fig. 3. Table S2 shows the usage of D segments and sequences of K59–containing sequences, which is related to Fig. 5. Online supplemental material is available at http://www.jem.org/cgi/content/full/jem.20111941/DC1.

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