γδ T cells protect against lung fibrosis via IL-22

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Inflammation–induced pulmonary fibrosis (PF) leads to irreversible loss of lung function and is a predictor of mortality in numerous lung diseases. Why some subjects with lung inflammation but not others develop PF is unclear. In a mouse model of hypersensitivity pneumonitis that progresses to lung fibrosis upon repeated exposure to the ubiquitous microorganism Bacillus subtilis, γδ T cells expand in the lung and inhibit collagen deposition. We show that a subset of these γδ cells represents the predominant source of the Th17 cytokine IL-22 in this model. Preventing expression of IL-22, either by mutating the aryl hydrocarbon receptor (AhR) or inhibiting AhR signaling, accelerated lung fibrosis. Direct blockade of IL-22 also enhanced collagen deposition in the lung, whereas administration of recombinant IL-22 inhibited lung fibrosis. Moreover, the presence of protective γδ T cells and IL-22 diminished recruitment of CD4+ T cells to lung. These data reveal a protective pathway that involves the inhibition of αβ T cells by regulatory IL-22–secreting γδ T cells.

Hypersensitivity pneumonitis (HP) is an inflammatory lung disease that results from repeated inhalation of aerosolized antigens (Selman, 2003). The etiologic agents are composed of a wide variety of organic particles (e.g., mammalian and avian proteins, fungi, and bacteria) and certain small-molecular-weight volatile and nonvolatile chemical compounds. The severity of the inflammatory response depends on the nature of the antigen, the quantity and duration of exposure, and host–environment interactions (Selman, 2003). In chronically exposed patients, pulmonary fibrosis (PF) occurs in up to 41% of cases, resulting in irreversible pulmonary dysfunction and right heart failure (Mönkäre and Haahela, 1987; Bourke et al., 1989; Lalancette et al., 1993; Yoshizawa et al., 1999). Importantly, lung fibrosis is an independent predictor of mortality in these patients, with a 5-yr mortality of 27% and a median survival of 13 yr (Vourlekis et al., 2004). Although PF is associated with numerous diffuse lung diseases, including autoimmune diseases (e.g., rheumatoid arthritis and systemic sclerosis), idiopathic PF, and sarcoidosis, its pathogenesis is poorly understood with few therapeutic options that arrest progression of fibrotic disease (Luzina et al., 2008).

γδ T cells are a unique subset of lymphocytes whose function is poorly understood. These cells, however, have been implicated in the regulation of the immune response generated against microbial pathogens and allergens (Born et al., 2007). The location of γδ T cells in the subepithelium of alveolar and nonalveolar regions of the lung (Wands et al., 2005) suggests that these cells play an important role in the immune response directed against inhaled particles such as microbial pathogens (Nanno et al., 2007). We recently reported that Vγ6/Vδ1+ γδ T cells isolated from the lung of mice chronically exposed to Bacillus subtilis express IL-17A (Simonian et al., 2009a). In this model of HP, C57BL/6 mice repeatedly treated with B. subtilis by nasal inhalation develop mononuclear infiltrates in the lung and PF that mimics the human disease (Simonian et al., 2006, 2009a; Barrera et al., 2008). Interestingly, large numbers of γδ T cells, nearly all of which are Vγ6/Vδ1+, accumulate in the lung in response to B. subtilis and attenuate lung inflammation and fibrosis upon continued exposure to this ubiquitous environmental microorganism (Simonian et al., 2006). In the absence of γδ T cells, TCR-δ−/− mice treated with B. subtilis have accelerated lung fibrosis but similar levels of IL-17A to...
those of WT mice as the result of a compensatory increase in IL-17A expression by CD4+ T cells. Conversely, IL-22 levels are reduced in TCR-δ−/− mice (Simonian et al., 2009a), suggesting that IL-22 may be an important mechanism by which Vγ6/Vδ1+ γδ T cells attenuate lung inflammation and fibrosis.

In this paper, we show that Vγ6/Vδ1+ γδ T cells are the predominant cell type in the lung producing IL-22 in response to chronic B. subtilis exposure. In addition, these γδ T cells differentially express IL-17A and IL-22, with IL-22 being expressed by a subset of IL-17F–producing Vγ6/Vδ1+ γδ T cells. In the presence of the d allele of the aryl hydrocarbon receptor (AhR), AhRd/d mice had reduced levels of IL-22 with accelerated collagen deposition in the lung. Administration of intratracheal recombinant mouse IL-22 (rIL-22) to B. subtilis–exposed AhRd/d and TCR-δ−/− mice significantly reduced lung inflammation and CD4+ T cell recruitment to lung and collagen deposition, suggesting that Vγ6/Vδ1+ γδ T cells regulate lung inflammation and fibrosis through IL-22.

RESULTS
Lung Vγ6/Vδ1+ γδ T cells express IL-22 in response to B. subtilis
We recently reported that WT C57BL/6 and homozygous Vγ6/Vδ1 transgenic (Vγ6+/+) mice repeatedly exposed to B. subtilis have increased IL-22 levels in the lung compared with mice deficient in γδ T cells (TCR-δ−/−; mice; Simonian et al., 2009a). To determine if γδ T lymphocytes are an important source of IL-22, we performed intracellular cytokine staining on T cells isolated from the lung of WT C57BL/6, Vγ6+/+, and TCR-δ−/− mice. Gating on CD3+γδ T cells, 6 and 9% of ex vivo Vγ6/Vδ1+ γδ T cells isolated from the lung of WT C57BL/6 and Vγ6+/+ mice, respectively, express IL-22 after repeated exposure to B. subtilis for four consecutive weeks (Fig. 1 A). Consistent with a previous study (Martin et al., 2009), IL-22–secreting γδ T cells also express CCR6 (not depicted). CD4+ T cells also produced IL-22 (Fig. 1 B), but the number of IL-22–expressing Vγ6/Vδ1+ γδ T cells was significantly greater than CD4+ T cells in the lung of WT C57BL/6 and Vγ6+ mice (Fig. 1 C). Although TCR-δ−/− mice had increased numbers of IL-22–expressing CD4+ T cells in the lung compared with WT C57BL/6 and Vγ6+ mice, levels of IL-22 were markedly diminished in the absence of γδ T cells (Fig. 1 D). In addition, IL-22 levels were consistently lower in the lung of TCR-δ−/− mice compared with Vγ6+ mice over the 4-wk time course and significantly reduced compared with WT C57BL/6 mice at 3 and 4 wk of repeated exposure to B. subtilis (Fig. 1 B). Collectively, these data suggest that although γδ and CD4+ T cells both express IL-22, Vγ6/Vδ1+ γδ T lymphocytes are the predominant source of IL-22 in the lung in response to repeated B. subtilis exposure.

Differential expression of IL-17A and IL-22 by Vγ6/Vδ1+ γδ T cells
Recent studies show that γδ T cells isolated from the peritoneal cavity after injection of heat-killed mycobacteria can

Figure 1. IL-22 expression by Vγ6/Vδ1+ γδ and CD4+ T cells.
(A) Representative density plots of intracellular IL-22 expression in Vγ6/Vδ1+ γδ T cells isolated from the lung of homozygous transgenic Vγ6/Vδ1 (B6.Vγ6+/-) and WT C57BL/6 mice treated with B. subtilis for 4 wk. Vγ6/Vδ1+ γδ T cells expressing IL-22 were selected from a bivariate dot plot of CD3+/CytC- profiles. The percentage of IL-22–expressing γδ T cells is shown. (B) Representative density plots of intracellular IL-22 expression in CD4+ T cells isolated from the lung of B6.Vγ6+/-, WT C57BL/6, and B6.TCRδ−/− mice treated with B. subtilis for 4 wk. CD4+ T cells were selected from a bivariate dot plot of CD3+/CytC- scatter cells from the lymphocyte gate. The percentage of IL-22–expressing CD4+ T cells is shown. (C) Absolute number of IL-22+ T cells expressing IL-22 is shown. (D) Absolute number of IL-22+ T cells expressing IL-22 is shown. (D) Absolute number of IL-22+ T cells expressing IL-22 is shown. (D) Absolute number of IL-22+ T cells expressing IL-22 is shown.
express IL-22 (Martin et al., 2009). Similar to CD4+ T cells, γδ T cells expressed either IL-17A alone or both IL-17A and IL-22 (Martin et al., 2009). In our model of HP and lung fibrosis induced by chronic exposure to B. subtilis, density plots of CD3+γδ+ T cells revealed two distinct γδ T cell populations (Fig. 2 A). Gating on the entire CD3+γδ+ T cell population (Gate A) demonstrated that γδ T cells isolated from the lung of Vy6/+ and WT C57BL/6 mice expressed IL-22, 11% expressed IL-17A, and very few cells expressed both cytokines. Conversely, γδ T cells isolated from the spleen of WT C57BL/6 and Vy6/+ mice treated with B. subtilis did not express IL-22 (Fig. S1), suggesting a compartmentalized immune response in the lung directed against B. subtilis.

Gating on the brighter subset of γδ T cells (Gate B), the percentage of IL-17A–expressing γδ T cells increased with a concomitant decrease in the expression of IL-22 in both Vy6/+ and WT C57BL/6 mice (Fig. 2 A, middle), suggesting that this population of γδ T cells more frequently expresses IL-17A. IL-17A expression was as high as 17% in this γδ T cell subset with fewer γδ T cells expressing IL-22 (2%). Gating on the less bright population of γδ T cells (Gate C) resulted in an increased percentage of IL-22–expressing γδ T cells, with as many as 12% of this subset producing IL-22, whereas fewer γδ T cells expressed IL-17A (Fig. 2 A, bottom). Overall, the number of IL-17A–expressing γδ T cells was markedly increased with a concomitant decrease in IL-22 expression in Gate B, whereas IL-22–expressing γδ T lymphocyte numbers were significantly increased in Gate C (Fig. 2 B). Because the vast majority of γδ T cells accumulating in the lung in response to B. subtilis express the Vy6/Vδ1 TCR (Fig. 1 A; Simonian et al., 2006), these data demonstrate that Vy6/Vδ1+ γδ T cells differentially express IL-17A and IL-22 in response to repeated exposure to B. subtilis, suggesting the possibility that subsets of Vy6/Vδ1+ γδ T cells may have distinct functional capabilities.

**Differential expression of IL-17A and IL-17F by Vy6/Vδ1+ γδ T cells**

In addition to IL-17A, IL-17F was detected in the lung of WT C57BL/6, Vy6/+, and TCR−δ− mice after repeated exposure to B. subtilis for four consecutive weeks (Fig. 3 A). As shown in Fig. 3 A, TCR−δ− mice have a twofold increased level of IL-17F in the lung compared with WT C57BL/6 mice and a 1.6-fold increase compared with Vy6/+ mice. Gating on CD3+CD4+ T cells, CD4+ T cells from the lung of TCR−δ− mice treated with B. subtilis differentially express IL-17A and IL-17F (Fig. 3 B). Unlike IL-17A, however, CD4+ T cells expressed high levels of IL-17F even in the presence of γδ T cells in both WT C57BL/6 and Vy6/+ mice (Fig. 3 B). In addition, the absolute number of both IL-17F– and IL-17A–expressing CD4+ T cells was increased in the lung of TCR−δ− mice compared with WT C57BL/6 and Vy6/+ mice (Fig. 3 C). Importantly, microbial elimination was similar in all three strains of mice (Simonian et al., 2009a).

As shown in Fig. 3 D, Vy6/Vδ1+ γδ T cells also differentially express IL-17A and IL-17F, with the majority of IL-17F expression identified in the less bright population of γδ T cells (Fig. 3 D, Gate C, middle), similar to IL-22. As many as 48 and 44% of γδ T cells isolated from the lung of B. subtilis–treated Vy6/+ mice and WT C57BL/6 mice, respectively, expressed IL-17F (Fig. 3 D, Gate A, top). In addition, gating
identified in a subset of IL-17F–expressing Vγ6/Vδ1+ γδ T lymphocytes.

**Decreased expression of IL-22 in AhR<sup>dd/d</sup> mice repeatedly exposed to B. subtilis**

Recent studies suggest that IL-22 expression requires the presence of AhR in CD4<sup>+</sup> and γδ T cells (Veldhoen et al., 2008; Martin et al., 2009). To determine if IL-22 production was affected by expression of the d allele on the C57BL/6 background in this model, we treated AhR<sup>dd/d</sup> mice with...
that were not significantly different than those of cells from *B. subtilis*–treated WT C57BL/6 mice (Fig. 4, A and B). Overall, the number of IL-22–expressing γδ T cells was decreased in the lung of AhR<sup>d/d</sup> mice compared with WT C57BL/6 mice repeatedly exposed to *B. subtilis* in an identical manner, whereas numbers of IL-17A– and IL-17F–expressing γδ T cells were unaffected (Fig. 4 C).

To determine whether decreased expression of IL-22 by Vγ6/Vδ1+ γδ T cells affects *B. subtilis*–induced lung fibrosis, AhR<sup>d/d</sup> mice treated with *B. subtilis* for four consecutive weeks were analyzed with Masson trichrome staining. As shown in Fig. 5 A, AhR<sup>d/d</sup> mice developed mononuclear infiltrates and collagen deposition in a peribronchovascular distribution similar to WT C57BL/6 mice treated in an identical fashion. We have previously shown that TCR–δ<sup>−/-</sup> mice develop significantly increased lung fibrosis after *B. subtilis* exposure and are included for comparison (Simonian et al., 2009a). By quantitative analysis using Sirius red, AhR<sup>d/d</sup> mice develop accelerated lung fibrosis with a 1.4-fold increased collagen content in the lung compared with *B. subtilis*–treated WT C57BL/6 mice (P < 0.05) and a nearly fourfold increase in collagen deposition compared with PBS–treated control animals (P < 0.001; Fig. 5 B). In addition, collagen deposition in the lungs of AhR<sup>d/d</sup> mice at 4 wk of repeated exposure to *B. subtilis* was similar to the exaggerated fibrotic response seen in TCR–δ<sup>−/-</sup> mice.

**Figure 4.** γδ T cells from B6.AhR<sup>d/d</sup> mice have reduced IL-22 expression. (A) Cytokines from supernatants of lung homogenates were analyzed by ELISA from WT C57BL/6 (wt) and B6.AhR<sup>d/d</sup> mice treated with either *B. subtilis* or PBS for 4 wk. The mean is shown as a solid line. Data are compiled from five individual mice from at least two separate experiments. (B) Representative density plots of intracellular IL-22, IL-17A, and IL-17F expression in γδ and CD4+ T cells isolated from the lung of B6. AhR<sup>d/d</sup> mice treated with *B. subtilis* for 4 wk. γδ T cells expressing IL-22, IL-17A, or IL-17F were selected from a bivariate dot plot of CD3+/C<sup>δδ</sup> cells from the lymphocyte gate. CD4+ T cells that express IL-22, IL-17A, and IL-17F were selected from a bivariate dot plot of CD3+/side scatter–expressing cells in the lymphocyte gate. The percentage of T cells expressing IL-22, IL-17A, or IL-17F is shown in the top right quadrant of the density plot. Data are representative of at least five individual mice per experiment from at least three separate experiments. (C) Absolute number of IL-22–, IL-17A–, and IL-17F–expressing γδ T cells in the lung of B6.AhR<sup>d/d</sup> and WT C57BL/6 mice treated with *B. subtilis* for 4 wk. Data represent individual mice per experiment from at least three separate experiments (mean ± SD). NS, not significant. P > 0.05.
(Simonian et al., 2009a). Collectively, these findings suggest that $\gamma_6/\delta^{-}$ $\gamma_6 T$ cells protect against lung fibrosis through expression of IL-22.

**Recombinant IL-22 decreases lung inflammation and fibrosis in AhR$^{\delta^{-}}$ and TCR-$\delta^{-/-}$ mice**

To confirm that IL-22 protects against $B. subtilis$-induced lung inflammation and fibrosis, we treated mice with decreased IL-22 production (i.e., AhR$^{\delta^{-}}$ and TCR-$\delta^{-/-}$ mice) with intratracheal rIL-22 (100 pg and 100 ng) in combination with $B. subtilis$ for 3 d each week for four consecutive weeks. As shown in Fig. 6 A, total cells and lymphocytes were significantly decreased in the lung of AhR$^{\delta^{-}}$ and TCR-$\delta^{-/-}$ mice treated with intratracheal rIL-22 and $B. subtilis$ compared with control mice exposed to vehicle and $B. subtilis$. In addition, no significant changes in the number of total lung cells or lymphocytes in WT C57BL/6 mice were seen (Fig. S2). There was no significant change in the number of $\gamma_6 T$ cells in the lung of both WT C57BL/6 and AhR$^{\delta^{-}}$ mice treated with rIL-22 and $B. subtilis$ (unpublished data). Conversely, CD4$^+$ T cells were significantly decreased in WT C57BL/6, AhR$^{\delta^{-}}$, and TCR-$\delta^{-/-}$ mice that received rIL-22 (Fig. 6 B), whereas treatment with rIL-22 alone had no effect on lung inflammation (not depicted).

Masson trichrome staining of lungs of AhR$^{\delta^{-}}$ and TCR-$\delta^{-/-}$ mice treated with $B. subtilis$ and rIL-22 showed reduced mononuclear cell infiltrates and collagen deposition in a peribronchovascular distribution (Fig. 6 C). Using Sirius red colorimetric assay for collagen quantification, AhR$^{\delta^{-}}$ and TCR-$\delta^{-/-}$ mice that received intratracheal rIL-22 had decreased levels of collagen deposition in the lung compared with mice treated with $B. subtilis$ alone. For example, there was a 2- and 1.5-fold decrease in collagen deposition in AhR$^{\delta^{-}}$ mice treated with 100 ng and 100 pg of rIL-22, respectively, compared with mice treated with $B. subtilis$ alone (Fig. 6 D). TCR-$\delta^{-/-}$ mice also had a significant decrease in collagen deposition in the lung with a 1.6- and 1.3-fold decrease in collagen content in the lung of mice treated with 100 ng and 100 pg of rIL-22, respectively, compared with TCR-$\delta^{-/-}$ mice exposed to $B. subtilis$ alone (Fig. 6 D). WT C57BL/6 treated with the higher concentration of rIL-22 (100 ng) had decreased collagen deposition compared with $B. subtilis$ treatment alone, whereas $\gamma_6/\delta^{-}$ mice, which have higher levels of IL-22 in the lung throughout the course of $B. subtilis$ exposure (Fig. 1 D), did not have any additional decrease in lung fibrosis at either dose of rIL-22 (Fig. 6 D).

In combination, these data demonstrate that exogenous rIL-22 is capable of modulating lung inflammation and fibrosis induced by chronic exposure to $B. subtilis$.

**Inhibition of IL-22 expression increases lung inflammation and fibrosis**

We next investigated whether inhibition of AhR signaling in $\gamma_6/\delta^{-}$ $\gamma_6 T$ cells would reduce IL-22 expression and accelerate the $B. subtilis$-induced inflammatory and fibrotic process. Because $\gamma_6/\delta^{-}$ mice have increased levels of IL-22 in the lung and develop significantly less inflammation and fibrosis, we treated these mice with a chemical inhibitor of AhR, CH-223191. CH-223191 is a novel chemical compound that potently inhibits AhR-dependent transcription induced by 2,3,7,8-Tetrachlorodibenzo-p-dioxin (TCDD; Kim et al., 2006). In addition, CH-223191 has been shown to block binding of TCDD to AhR and inhibit TCDD-mediated nuclear translocation and DNA binding of AhR, thus preventing expression of target genes of AhR without detectable AhR agonist activity (Kim et al., 2006).

$\gamma_6/\delta^{-}$ mice were treated with $B. subtilis$ and either 10 mM or 10 $\mu$M of CH-223191 for three consecutive days each week for four consecutive weeks. As shown in Fig. 7 A,
of increased numbers of CD4+ T cells, lung fibrosis was accelerated, as assessed by both Masson's trichrome staining and quantitatively by Sirius red colorimetric assay, with a twofold increase in collagen deposition in the lung of Vγ6+/- mice treated with 10 mM CH-223191 compared with control mice treated in an identical manner with B. subtilis and vehicle (Fig. 7, E and F).

To directly determine if blocking IL-22 results in increased lung inflammation and fibrosis, we treated WT C57BL/6 and TCR-δ-/- mice with anti–IL-22 mAb in combination with B. subtilis for 4 wk. Lung inflammation, as assessed by total cell numbers, was not significantly different between mice treated with the anti–IL-22 antibody compared with isotype controls (Fig. 8 A). In contrast to γδ T cells in the lung of WT C57BL/6 mice, which were unaltered by the addition of the anti–IL-22 antibody, CD4+ T cells were treatment of Vγ6+/- mice with B. subtilis in combination with the AhR inhibitor resulted in decreased frequency of γδ T cells that express IL-22 compared with mice treated with B. subtilis and vehicle. In addition, the number of IL-22–expressing γδ T cells and the level of IL-22 in the lung were markedly reduced in Vγ6+/- mice treated with the AhR inhibitor at both concentrations relative to mice treated with B. subtilis and vehicle (Fig. 7, B and C). With reduced IL-22 expression, lung inflammation was accelerated with increased total cells, whereas the number of γδ T cells was relatively unaffected in the lung of Vγ6+/- mice treated with the AhR inhibitor compared with mice exposed to B. subtilis and vehicle (Fig. 7 D). CD4+ T cells, however, were markedly increased (more than threefold) in the lung of Vγ6+/- mice treated with the AhR inhibitor compared with mice exposed to B. subtilis treated with vehicle alone (Fig. 7 D). In the presence of increased numbers of CD4+ T cells, lung fibrosis was accelerated, as assessed by both Masson's trichrome staining and quantitatively by Sirius red colorimetric assay, with a twofold increase in collagen deposition in the lung of Vγ6+/- mice treated with 10 mM CH-223191 compared with control mice treated with vehicle alone (Fig. 7 D). In the presence...
expressing IL-22 were selected from a bivariate dot plot of CD3+/side B. subtilis
scatter cells from the lymphocyte gate. The percentage of CD3+ T cells in
(B) Number of IL-22–expressing least five individual mice per experiment treated with CH-223191 at each
each quadrant of the density plot is shown. Data are representative of at
density plots of intracellular IL-22 expression in γδ T cells isolated from
the lung of B6.Vγδ6+ mice treated with a combination of AhR inhibitor, CH-223191 and
B. subtilis or vehicle for four consecutive weeks. Data were compiled from at least five mice per
experiment from two separate experiments. (D) Number of cells in the lung of B6.Vγδ6+ mice treated with a
combination of AhR inhibitor, CH-223191, and B. subtilis or vehicle for four consecutive weeks. Data were
compiled from at least five mice per experiment from two separate experiments. (E) Rep-
acceleration of lung inflammation and fibrosis in trans-
genic Vγδ6/Vδ1 mice treated with an AhR inhibitor. (A) Representative
density plots of intracellular IL-22 expression in γδ T cells isolated from
the lung of B6.Vγδ6+ mice treated with the AhR inhibitor CH-223191 and
B. subtilis or B. subtilis with vehicle for four consecutive weeks. γδ T cells
expressing IL-22 were selected from a bivariate dot plot of CD3+/side scatter cells from the lymphocyte gate. The percentage of CD3+ T cells in
each quadrant of the density plot is shown. Data are representative of at
least five individual mice per experiment treated with CH-223191 at each
concentration with B. subtilis from at least two separate experiments.
(B) Number of IL-22–expressing γδ T cells in the lung of B6.Vγδ6+ mice (+/+)
treated with a combination of AhR inhibitor, CH-223191, and B. subtilis or
significantly increased in the lung of WT C57BL/6 mice treated with anti–IL-22 antibody compared with isotype
treatment control (Fig. 8 A). In the presence of an already exaggerated lung
defensive myeloid response (Simonian et al., 2006), anti–IL-22 antibody had no effect on the recruitment of CD4+ T cells to the lung of TCR-δ−/− mice (Fig. 8 A). As shown in Fig. 8 B, WT C57BL/6 mice treated with anti–IL-22 antibody had accelerated collagen deposition in the lung compared with mice treated in an identical fashion with an isotype control antibody. Exposure of TCR-δ−/− mice to B. subtilis and anti–IL-22 antibody did not result in a further elevation of
collagen deposition in the lung compared with mice treated with the isotype control (Fig. 8 B), likely because of the pre-
ence of significantly decreased IL-22 in the lung of TCR-δ−/− mice relative to WT C57BL/6 mice (Fig. 1 D). Collectively,
these data provide direct evidence that IL-22 regulates the accumulation of CD4+ T cells in the lung and B. subtilis–
induced lung fibrosis.

CD4+ T cells promote lung fibrosis in response to B. subtilis
In the absence of γδ T cells, we have previously shown that an increased number of CD4+ T cells are recruited to the
lung (Simonian et al., 2006). To determine whether the activation and differentiation state of CD4+ T cells is affected by the presence or absence of IL-22–secreting Vγδ6/Vδ1+ γδ T cells after B. subtilis exposure, we analyzed lung CD4+ T cells from WT C57BL/6, TCR-δ−/−, and Vγδ4−/− mice. As compared with CD4+ T cells from WT C57BL/6 mice, a significantly increased percentage of CD4+ T lymphocytes from the lung of TCR-δ−/− and Vγδ4−/− mice expressed high levels of CD44, a memory cell marker, and CD69, a marker of recent activation (Fig. 9 A). In addition, a reduced
percentage of T cells expressed CD45RB, CD27, and CD62L, indicating that a larger fraction of CD4+ T cells in B. subtilis–treated TCR-δ−/− and Vγδ4−/− mice had a

B. subtilis and vehicle for four consecutive weeks. Data were compiled from at least five mice per experiment from two separate experiments (mean ± SD). (C) IL-22 concentration in the lung of B6.Vγδ6+ mice treated with a
combination of AhR inhibitor, CH-223191, and B. subtilis or B. subtilis and vehicle for four consecutive weeks. The mean is shown as a solid line. Data were compiled from five individual mice from at least two separate experi-
ments. (D) Number of cells in the lung of B6.Vγδ6+ mice (+/+) treated with a combination of AhR inhibitor, CH-223191, and B. subtilis or vehicle for four consecutive weeks. Data were compiled from at least five mice per experiment from two separate experiments (mean ± SD). (E) Rep-
sentative Masson's trichrome staining of lungs from B6.Vγδ6+ mice repeatedly treated with the AhR inhibitor, CH-223191, and B. subtilis compared with mice treated with B. subtilis and vehicle for four consecutive weeks is shown. Arrow denotes collagen deposition in a peribronchovascular
distribution. Data shown represent at least five mice from at least two separate experiments (40×). Bars, 100 µm. (F) Quantification of collagen
collesting content using Sirius red colorimetric assay in the lungs of individual B6.
Vγδ6+ mice treated with the AhR inhibitor CH-223191 compared with mice treated with vehicle and B. subtilis or sterile PBS alone for four con-
secutive weeks is shown. The mean is shown as a solid line. Data were compiled from five individual mice from at least two separate experiments.

Figure 7. Acceleration of lung inflammation and fibrosis in trans-
genic Vγδ6/Vδ1 mice treated with an AhR inhibitor. (A) Representative
density plots of intracellular IL-22 expression in γδ T cells isolated from
the lung of B6.Vγδ6+ mice treated with the AhR inhibitor CH-223191 and
B. subtilis or B. subtilis with vehicle for four consecutive weeks. γδ T cells
expressing IL-22 were selected from a bivariate dot plot of CD3+/side scatter cells from the lymphocyte gate. The percentage of CD3+ T cells in
each quadrant of the density plot is shown. Data are representative of at
least five individual mice per experiment treated with CH-223191 at each
concentration with B. subtilis from at least two separate experiments.
(B) Number of IL-22–expressing γδ T cells in the lung of B6.Vγδ6+ mice (+/+)
treated with a combination of AhR inhibitor, CH-223191, and B. subtilis or

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highly activated effector phenotype, as compared with those from the lung of WT C57BL/6 mice (Fig. 9 A).

To test whether CD4+ T cells represent a pathogenic T cell subset and promote lung fibrosis in response to B. subtilis, we adoptively transferred CD4+ and CD8+ T cells from the spleen of naive WT C57BL/6 mice into TCR-δ−/− mice and subsequently treated them with B. subtilis by nasal inhalation. As shown in Fig. 9 B, mice receiving CD4+ T cells developed peribronchovascular mononuclear infiltrates similar to WT C57BL/6 mice treated in an identical fashion with B. subtilis (Simonian et al., 2006). By quantification of collagen content using Sirius red colorimetric assay in the lungs of individual WT C57BL/6 (wt) and B6.TCRδ−/− (δ−/−) mice treated with B. subtilis in combination with anti–IL-22 (il22) or isotype control (IC) for 3 d a week for four consecutive weeks is shown. Data were compiled from at least five individual mice from two separate experiments (mean ± SD). (B) Quantification of collagen content in the lungs of naive WT C57BL/6 mice (wt) and B6.TCRδ−/− mice treated with B. subtilis in combination with anti–IL-22 (il22) or isotype control (IC) for 3 d a week for four consecutive weeks is shown. The mean is shown as a solid line. Data represent five individual mice per experiment from two separate experiments. NS, not significant. P > 0.05.

Figure 8. Administration of anti–IL-22 enhances the B. subtilis–induced CD4+ T cell alveolitis and accelerates lung fibrosis. (A) Total and differential cell counts performed on lung homogenates from WT C57BL/6 (wt) and B6.TCRδ−/− mice treated with B. subtilis in combination with anti–IL-22 (il22) or isotype control (IC) for 3 d a week for four consecutive weeks is shown. Data were compiled from at least five individual mice from two separate experiments (mean ± SD). (B) Quantification of collagen content using Sirius red colorimetric assay in the lungs of individual WT C57BL/6 (wt) and B6.TCRδ−/− mice treated with B. subtilis in combination with anti–IL-22 (il22) or isotype control (IC) for 3 d a week for four consecutive weeks is shown. The mean is shown as a solid line. Data represent five individual mice per experiment from two separate experiments. NS, not significant. P > 0.05.

by nasal inhalation (P < 0.01; Fig. 9 C). TCR-β−/−δ−/− mice reconstituted with CD8+ T cells also had a slight increase in collagen content in the lung when chronically exposed to B. subtilis, compared with PBS-treated CD8+ T cell–reconstituted control mice. However, similar levels of collagen deposition were seen in B. subtilis–treated TCR-β−/−δ−/− mice that were sham reconstituted by injecting sterile PBS into the tail vein (Fig. 9 C). Therefore, CD8+ T cells do not appear to have a significant role in the development of PF. Collectively, these data suggest that activated CD4+ T cells that accumulate in the lung in response to repeated B. subtilis exposure promote lung fibrosis.

rIL-22 reduces accumulation of CXCR3-expressing CD4+ T cells in the lung

Although Th1 cells typically express CXCR3, Th17 cells have also been shown to express this chemokine receptor (Acosta-Rodriguez et al., 2007). In addition, CXCR3 expression has been shown to be important in other models of lung injury/fibrosis such as bleomycin-induced lung fibrosis (Jiang et al., 2004). To determine if IL-22 affects CXCR3 expression on CD4+ T cells and recruitment of activated CD4+CXCR3+ T lymphocytes in the lung, we treated WT C57BL6, Ahr/d/d, and TCR-δ−/− mice with rIL-22 and B. subtilis for 4 wk. As shown in Fig. 10 (A and B), the percentage and number of CXCR3-expressing CD4+ T cells isolated from the lung of WT C57BL6, Ahr/d/d, and TCR-δ−/− mice treated with rIL-22 was significantly decreased compared with mice treated with B. subtilis and vehicle.

Next, we evaluated total lung homogenates from Vγ6+/+, WT C57BL/6, and TCR-δ−/− mice for the CXCR3 ligands CXCL9, 10, and 11. As shown in Fig. 10 C, levels of CXCL9 and CXCL10 were significantly increased in the lung of TCR-δ−/− mice compared with Vγ6+/+ and WT C57BL/6 mice repeatedly exposed to B. subtilis over 4 wk, whereas CXCL11 was no different between the various strains of mice. To determine if exogenous rIL-22 could alter the levels of CXCR3 ligands in the lung, we treated WT C57BL6, TCR-δ−/−, and Ahr/d/d mice with B. subtilis and rIL-22 for 4 wk. As shown in Fig. 10 D, levels of CXCL9 were decreased in the lung of WT C57BL/6 mice treated with 100 ng rIL-22 but not at the lower dose. CXCL9 levels, however, were decreased in the lung of TCR-δ−/− and AhR/d/d mice at both concentrations of rIL-22 (Fig. 10 D). Levels of CXCL10 were not affected by treatment with rIL-22, suggesting that γδ T cells may affect CXCL10 expression independent of IL-22 (Fig. S3). Conversely, levels of CXCL9 were increased in the lung of Vγ6+/+ mice treated with B. subtilis for 4 wk in the presence of the AhR inhibitor CH-223191 (Fig. 10 E). Collectively, these data suggest that IL-22 alters the expression of the CXCR3 ligand CXCL9, resulting in a reduced recruitment of CD4+CXCR3+ T cells to the lung in response to repeated exposure to B. subtilis.

DISCUSSION

We developed a mouse model of HP to investigate the role of different T cell subsets in lung inflammation and fibrosis induced by chronic exposure to a common environmental
of WT C57BL/6 compared with TCR-

each time point. (B) Representative H
tom) staining of lungs from TCR-

CD62L on CD4+ T cells isolated from the lungs of WT C57BL/6, TCR-

induce lung fibrosis.

Data represent at least four individual mice from at least two separate

Arrow denotes collagen deposition in a peribronchovascular distribution.

Figure 9. CD4+ T cells represent the pathogenic T cell subset and induce lung fibrosis. (A) Expression of CD44, CD69, CD45RB, CD27, and CD62L on CD4+ T cells isolated from the lungs of WT C57BL/6, TCR-δ−/−, and Vγ4−/−δ−/− mice treated with B. subtilis for 4, 8, and 12 consecutive weeks is shown. Data represent the mean ± SD of at least five mice at each time point treated with B. subtilis from at least two separate experiments. The asterisks denote a statistically significant (P < 0.05) difference in the percentage of CD4+ T cells that express each surface marker from the lung of WT C57BL/6 compared with TCR-δ−/− mice treated with B. subtilis at each time point. (B) Representative H&E (top) and Masson trichrome (bottom) staining of lungs from TCR-β−/−δ−/− mice reconstituted with either CD4+ or CD8+ T cells or sterile PBS followed by intranasal treatment with either B. subtilis or sterile PBS for four consecutive weeks is shown. Arrow denotes mononuclear infiltrates in the peribronchovascular space. Arrow denotes collagen deposition in a peribronchovascular distribution. Data represent at least four individual mice from at least two separate experiments.

B. subtilis

microorganism, B. subtilis (Simonian et al., 2006). In this model, Vγ6/Vδ1+ γδ T cells expand in the lung in response to the microorganism and attenuate B. subtilis–induced lung inflammation and fibrosis (Simonian et al., 2006). In addition to IL-17A, the current study shows that Vγ6/Vδ1+ γδ T cells isolated from the lung of mice chronically exposed to B. subtilis express IL-22 and that expression of IL-22 is an important mechanism by which these T cells attenuate lung inflammation and fibrosis.

Vγ6/Vδ1+ γδ T cells are a rare population of γδ T cells that normally reside in the female reproductive tract (Itohara et al., 1990), tongue (Itohara et al., 1990), and lung (Sim et al., 1994) but are rarely found in other tissues (Roark et al., 2004). Expansions of this TCR-invariant subset have been reported in the liver of Listeria monocytogenes–infected mice (Roark et al., 1996), tests during Listeria–induced orchitis (Mukasa et al., 1997), peritoneum of Escherichia coli–infected mice (Matsuzaki, Takada, and Nomoto, 1999), kidneys of mice infected intrarenally with L. monocytogenes (Ikebe et al., 2001), kidneys of rats treated with Adriamycin (Ando et al., 2001), and brains of mice with experimental autoimmune encephalitis (Olive, 1995). In response to intratracheal B. subtilis, the lung γδ T cell expansion is almost completely limited to the Vγ6/Vδ1+ γδ T cell subset and substantially exceeds that of both CD4+ and CD8+ αβ-expressing T cells (Simonian et al., 2006, 2009a). As a result of its invariant TCR, it is likely that the TCR ligand for Vγ6/Vδ1+ γδ T cells is a host molecule whose expression is induced during inflammation. However, we and others have previously shown that live bacteria are required to induce a γδ T cell response (Griffin et al., 1991; Boom et al., 1992; Skeen and Ziegler, 1993; Simonian et al., 2006).

Consistent with previous studies (Weaver et al., 2007; Veldhoen et al., 2008, 2009; Simonian et al., 2009a), we identified distinct populations of Vγ6/Vδ1+ γδ T cells that differentially express IL-17A and IL-22. In addition, IL-17A and IL-17F were also differentially expressed, with IL-22 expression identified as a subset of IL-17F–expressing Vγ6/Vδ1+ γδ T cells. These findings suggest that distinct Vγ6/Vδ1+ γδ T cell subsets may possess a range of functional capabilities.

Recent data suggest that IL-17A and IL-17F may have complementary roles in host defense against Staphylococcus aureus (Ishigame et al., 2009). IL17a−/−IL17f−/− mice were more susceptible to mucocutaneous abscesses that contained S. aureus than either IL17a−/− or IL17f−/− mice, suggesting that S. aureus infection can be independently controlled by IL-17A or IL-17F (Ishigame et al., 2009). Although IL-17A and IL-17F bind to the same receptors, the binding affinities of IL-17A and IL-17F experiments for H&E and Masson Trichrome staining (40x). Bars, 100 μm. (C) Quantification of collagen content using Sirius Red by colorimetric assay in the lungs of individual TCR-β−/−δ−/− (B−/−δ−/−) mice reconstituted with CD4+, CD8+, or sterile PBS by tail vein injection followed by repeated intranasal treatment with B. subtilis (Bsub) or sterile PBS for four consecutive weeks. The mean is shown as a solid line. Data were compiled from five individual mice from at least two separate experiments.
likely determines where they exert their functional capabilities (Gaffen, 2009). IL-17RA mRNA was shown to be highly expressed in lymphoid tissues such as the thymus, spleen, and lymph nodes, whereas IL-17RC mRNA was expressed in nonhematopoietic tissues such as the colon, small intestine, and lung (Ishigame et al., 2009), with IL-22 receptor expression reported on bronchial epithelial cells (Aujla et al., 2008). Collectively, these data suggest that IL-17A–expressing V66/V13+ T cells most likely exert their function on hematopoietic tissue to promote elimination of the microorganism, whereas the population of C3D4+ T cells expressing CXCR3 is shown in the top right quadrant of the density plot. Data represent at least five individual mice per experiment from two separate experiments. (B) Number of CXCR3-expressing C4+ T cells in the lung of WT C57BL/6 (wt), B6.AhR+/+(AhR), and B6.TCR6−/− mice treated with a combination of recombinant IL-17A and B. subtilis or vehicle for four consecutive weeks. Data were compiled from at least five individual mice per experiment from two separate experiments. (C) Levels of CXCL9, 10, and 11 in the lung of WT C57BL/6 (●), B6.Vγ6+ (●), and B6.TCR6−/− (▲) mice treated with B. subtilis for four consecutive weeks compared with PBS control. The asterisks denote a statistically significant (P < 0.05) difference in lung chemokine concentration at each time point between B6.TCR6−/− and B6.Vγ6+ mice. Data were compiled from at least five individual mice at each time point from two separate experiments (mean ± SD). (D) Levels of CXCL9 in the lung of WT C57BL/6, TCR−/−, and AhR+/− mice treated with a combination of recombinant IL-17A and B. subtilis, B. subtilis, or PBS for four consecutive weeks. The mean is shown as a solid line. Data were compiled from five individual mice from two separate experiments. (E) Levels of CXCL9 in the lung of B6.Vγ6+ mice treated with a combination of the AhR inhibitor, CH-223191, and B. subtilis, B. subtilis, or PBS for four consecutive weeks. The mean is shown as a solid line. Data were compiled from five individual mice from two separate experiments.

and IL-17F for IL-17RA and IL-17RC may be different (Hymowitz et al., 2001; Kuestner et al., 2007; Wright et al., 2008). In mice, IL–17RA binds both IL–17A and IL–17F, but IL–17RC interacts with only IL–17F (Kuestner et al., 2007). Therefore, tissue distribution of the receptors for these cytokines receptor expression reported on bronchial epithelial cells (Aujla et al., 2008). Collectively, these data suggest that IL–17A–expressing Vγ6/Vδ1+ T cells most likely exert their function on hematopoietic tissue to promote elimination of the microorganism, whereas the population of Vγ6/Vδ1+ T cells.
that express IL-22 and IL-17F may promote epithelial integrity in the lung through IL-22 receptor or IL-17R-C signaling.

Because recent studies show that AhR signaling is important for IL-22 expression by αβ− and γδ− expressing T cells (Veldhoen et al., 2008; Martin et al., 2009), we analyzed and found substantial reductions in IL-22 levels in the lung of B. subtilis−treated AhRd/d mice as well as reduced IL-22 expression by Vγ6/Vδ1+ γδ T cells, whereas IL-17A and IL-17F expression was unaffected. With the reduction in IL-22 levels in the lung of B. subtilis−treated AhRd/d mice, lung fibrosis was accelerated. The corroboration of these findings using an anti–IL-22 mAb suggests that IL-22 plays a critical role in regulating collagen deposition in the lung. We recently showed that in the absence of IL-17A receptor signaling, IL-17ra−/− mice had delayed clearance of B. subtilis and increased lung inflammation and fibrosis (Simonian et al., 2009a). Conversely, despite reduced IL-22 expression, AhRd/d mice chronically exposed to B. subtilis had no defects in bacterial clearance (Fig. S4), likely because of an unaffected IL-17A response. Although synergy between IL-17A and IL-22 cannot be excluded, these data support the concept that IL-17A−expressing Vγ6/Vδ1+ γδ T cells promote microbial elimination, whereas the IL-22−expressing population limits lung inflammation and fibrosis. In the bleomycin model of lung inflammation and fibrosis, however, IL-17A is necessary for lung fibrosis either through IL-1β (Wilson et al., 2010) or complex interactions that regulate IL-22 expression and function (Sonnenberg et al., 2010). In addition, IL-17A has been shown to be pathogenic in another model of HP induced by repeated exposure to Saccharopolyspora rectivirgula (Joshi et al., 2009; Simonian et al., 2009b). Therefore, whether IL-17A and IL-22 protect or promote lung inflammation and fibrosis is likely dependent on the inhaled agent.

IL-22 has emerged as an important cytokine in mucosal immunity (Aujla and Kolls, 2009). The mechanisms by which IL-22 protects epithelial structures in the lung, gut, and other organ systems from damage induced by numerous environmental insults are under intense investigation. Emerging data suggest that IL-22 may provide mucosal protection by inducing antimicrobial peptides from epithelial cells in the lung, gut, and skin as well as maintaining epithelial integrity either by preventing injury or accelerating epithelial repair after a variety of insults (Liang et al., 2006; Zheng et al., 2007, 2008; Aujla et al., 2008). In a mouse model of host defense against Gram-negative bacterial infection, WT mice given anti–IL-22 antibody before lung infection with Klebsiella pneumoniae had greater dissemination of infection and increased mortality than mice given a control antibody (Aujla et al., 2008). The combination of IL-17A and IL-22 significantly increased expression of an antimicrobial protein, lipocalin-2, in mouse tracheal epithelial cells, suggesting that IL-22 and IL-17A act synergistically, although IL-22 was shown to be more important for microbial elimination (Aujla et al., 2008). IL-22 was also found to induce clonogenic potential in human bronchial epithelial cells as well as enhanced recovery of epithelial resistance after epithelial injury (Aujla et al., 2008). In the B. subtilis model, however, we found increased numbers of activated CD4+ T cells in the lung of mice deficient in IL-22−expressing Vγ6/Vδ1+ γδ T cells and accelerated lung fibrosis. Upon treatment with recombinant IL-22, AhRd/d and TCR−δ−/− mice not only had reduced collagen deposition but also a diminished CD4+ T cell alveolitis. Conversely, transgenic Vγ6/Vδ1+ mice treated with the AhR inhibitor CH-23191 had decreased levels of IL-22 and increased numbers of CD4+ T cells in the lung that resulted in loss of protection against B. subtilis−induced lung fibrosis. In addition, neutralization of IL-22 by treatment with an anti–IL-22 mAb resulted in elevated numbers of CD4+ T cells and acceleration of lung fibrosis. Adoptive transfer of CD4+ T cells into mice deficient in both αβ and γδ T cells resulted in collagen deposition in the lung, further implicating CD4+ T lymphocytes in the pathogenesis of B. subtilis−induced lung fibrosis.

Th17 cytokines induce numerous chemokines and growth factors to promote cellular recruitment to the site of injury (for review see Alcorn et al., 2010). In the absence of γδ T cells and the resultant diminution in IL-22 expression in our model, the lungs of B. subtilis−treated TCR−δ−/− mice contain a significantly greater number of CD4+CXCR3+ T cells that express an activated effector phenotype as compared with WT C57BL/6 mice. The diminished CD4+CXCR3+ T cell inflammatory response in WT C57BL/6, TCR−δ−/−, and AhRd/d mice as a result of exogenous rIL-22 administration suggests that the mechanism by which IL-22 decreases CD4+ T cell recruitment to lung in response to B. subtilis is through decreased expression of CXCR3 ligands. Consistent with this observation, levels of the CXCR3 ligand CXCL9 were decreased in the lung of both AhRd/d and TCR−δ−/− mice in response to rIL-22. Because the IL-22 receptor and expression of CXCL9 have been demonstrated on lung epithelial cells (Aujla et al., 2008), rIL-22 may decrease expression of CXCL9 by lung epithelial cells that limit the recruitment of CD4+CXCR3+ T cells to the lung in response to repeated exposure to B. subtilis.

The fact that lung fibrosis was not completely abrogated in AhRd/d and TCR−δ−/− mice treated with rIL-22 raises the possibility that either higher levels of IL-22 in lung are required or other factors expressed by γδ T cells are necessary to further reduce collagen deposition in the lung. Therefore, IL-17A, IL-17F, or other factors expressed by Vγ6/Vδ1+ γδ T cells may still act synergistically with IL-22 to reduce CD4+ T cell recruitment to the lung and, thus, lung fibrosis. However, the presence of an exaggerated B. subtilis−induced fibrotic response in AhRd/d and TCR−δ−/− mice, despite high levels of IL-17A and IL-17F in the lung, strongly suggests that IL-22 is the more critical Th17 cytokine for protecting against lung fibrosis induced by chronic exposure to B. subtilis.

Although cessation of antigen exposure often improves lung inflammation, irreversible PF still develops in ~41% of HP subjects and is associated with increased morbidity and mortality (Vourlekis et al., 2004). Current therapeutic options for HP patients are limited to corticosteroids which nonspecifically suppress T cell function. The ability of rIL-22 to regulate γδ T cells in lung inflammation and fibrosis | Simonian et al.
PF in mice raises the possibility that immunotherapy directed toward increasing IL-22 levels in the lung may be beneficial in diseases characterized by chronic lung inflammation.

MATERIALS AND METHODS

Treatment of mice. 8-wk-old C57BL/6, TCR-δ−/−, TCR-Bγ−/−δ−/−, and AhR−/− mice (The Jackson Laboratory) and transgenic Vy6+ (Sim et al., 1995; Roark et al., 2004) and Vy4−/−δ−/− (Simonian et al., 2006) mice were treated with 30 µl (5 million CFU) B. subtilis (American Type Culture Collection) or sterile PBS on three consecutive days each week for up to four consecutive weeks by nasal inhalation. All mice have been backcrossed onto a C57BL/6 background for at least 10 generations. B. subtilis was prepared as previously described (Simonian et al., 2006). A lumines amebocyte assay (Sigma-Aldrich) confirmed that the B. subtilis preparation and sterile PBS contained <20 µg of endotoxin/ml. For all experiments using 100 ng or 100 pg of mouse IL-22 (R&D Systems) or 10 mM or 10 µM of the AhR inhibitor CH-233191 (EMD), mice were given IL-22 or CH-233191 by nasal inhalation in combination with B. subtilis for three consecutive days each week for four consecutive weeks. IL-22 was dissolved in PBS with 1% BSA, whereas CH-23191 is dissolved in DMSO. Therefore, PBS with 1% BSA and DMSO were used as vehicle controls. For experiments involving anti–IL-22 mAb, mice were treated with B. subtilis in combination with 1 µg of either anti–IL-22 or isotype control mAb (R&D Systems) for 3 d a week for four consecutive weeks. On the other 4d of the week, mice were treated with either anti–IL-22 or isotype control mAb suspended in sterile PBS. Mice were lightly anesthetized with isoflurane to allow inhalation of either B. subtilis or sterile PBS. These studies were approved by the Animal Care and Use Committee at the University of Colorado Denver.

Preparation of mononuclear cells from lung homogenates. Mice were sacrificed 24 h after the last treatment with B. subtilis as previously described (Simonian et al., 2006). The right lung was removed, snap-frozen in liquid nitrogen, and stored at −80°C for collagen quantification after sterile PBS was infused through the pulmonary vasculature by right heart puncture to remove any contaminating peripheral blood mononuclear cells. The left lung was digested with collagenase (Sigma-Aldrich) and DNase (Boehringer Mannheim). The collagenase-digested lungs were layered on top of Ficoll (Accurate Chemical & Scientific Corporation) and centrifuged at 1,200 rpm for 30 min at room temperature. The interface between the medium and Ficoll was removed and washed twice with RPMI 1640 containing 5% FBS.

Flow cytometry and immunofluorescence analysis. T cells isolated from the lungs of mice treated with either B. subtilis or sterile PBS were surface stained with mAbs directed against CD3 (PECy7; BD), CD4 (Alexa Fluor 405; BD), CD8 (FITC; BD), CXCR3 (APC; eBioscience), CD44 (APC; eBioscience), CD45RB (PE; eBioscience), CD27 (APC; eBioscience), CD26L (PECy5.5; Becton Dickinson), and CD69 (PECy5.5; eBioscience). For intracellular cytokine staining, total lung cells were cultured at 106 cells/ml in complete RPMI media containing 10 µg/ml brefeldin A (Sigma-Aldrich) at 37°C for 4 h. The cells were washed and stained with mAbs directed against CD3 (PerCP; BD), CD4 (FITC; BD), and CD8 (FITC; BD), followed by fixation in 1% paraformaldehyde overnight. γδ T cells that express the Vy6− TCR were identified using 17D1 mAb (Roark et al., 2004). Fixed cells were permeabilized in 0.5% saponin and stained with IL-17A (APC; eBioscience), IL-22 (PE; R&D Systems), and IL-17F (APC or PE; R&D Systems) mAbs and isotype controls for 30 min at 4°C. The lymphocyte population was identified using forward and 90° light scatter patterns, and fluorescence intensity was analyzed using a FACSAria cytometer (BD). The data files were analyzed using FlowJo software (Tree Star, Inc.).

Histology. Mice were sacrificed 24 h after their last exposure to B. subtilis or sterile PBS. The lungs were removed and infused with 10% formalin, embedded in paraffin, and stained with hematoxylin and eosin (H&E) and Mason Trichrome (Sigma-Aldrich) according to the manufacturer’s instructions.

Cytokine analysis and collagen quantification. Total lung homogenates were prepared by homogenizing whole lung samples in 500 µl of sterile PBS from each group of mice treated with either B. subtilis or sterile PBS for four consecutive weeks. 100 µl of supernatant from each lung homogenate was analyzed for cytokines by ELISA for IL-17A (eBioscience), IL-17F, IL-22, CXCL9, CXCL10, and CXCL11 (R&D Systems) according to the manufacturer’s instructions. The collagen content of the right lung was determined using Sirius Red staining as previously described (Simonian et al., 2006).

Total and differential cell counts. Total and differential cell counts were performed on collagenase-digested lung before purification of mononuclear cells over Ficoll as previously described (Simonian et al., 2006, 2009b). In brief, single-cell suspensions of total lung cells were transferred to a glass slide using a cytocentrifuge apparatus and stained with Wright and Giemsa per manufacturer’s instructions. Epithelial cells were not included in the total cell count and RBCs were removed by RBC lysis. Differential cell counts were performed by counting at least 400 cells under a high power field.

Adoptive transfer experiments. CD4+ and CD8+ T cells were isolated from spleens of untreated C57BL/6 mice and purified by MACS (Miltenyi Biotec) using negative selection for either CD4+ or CD8+ T lymphocytes. Flow cytometry confirmed >95% purity for each T cell population before adoptive transfer of 5 × 107 cells/100 µl of sterile PBS by tail vein injection into TCR-δ−/−δ−/− mice. In brief, total splenocytes were obtained by pushing spleens through a 70-µm pore-size mesh (Falcon) with subsequent resuspension in RBC lysis buffer (eBioscience). Cells were resuspended in buffer and biotinylated antibody cocktail, mixed, and incubated at 4°C for 10 min. Anti-biotin microbeads were added per the manufacturer’s instructions, mixed, and incubated for 15 min at 4°C. The cells were washed and applied to the column per the manufacturer’s instructions. The effluent, which contained the unlabeled CD4+ or CD8+ T cells, was collected and passed over a second column to further enrich each T cell population. Cell counts for CD4+ and CD8+ T lymphocytes were determined by Coulter counter (Beckman Coulter).

Statistical analysis. A Mann–Whitney U analysis and one-way ANOVA with a Bonferroni’s multiple comparison test were used to determine whether there were significant differences between the treatment groups at each time point (Prism 4; GraphPad Software). P < 0.05 was considered statistically significant.

Online supplemental material. Fig. S1 demonstrates that IL-17A and IL-22 are not expressed by γδ T cells isolated from the spleen of homozygous transgenic Vy6+/V61 (B6.Vy6+δ−/) mice repeatedly exposed to B. subtilis. Fig. S2 shows that recombinant IL-22 does not decrease levels of CXCL10 in the lung. Fig. S3 demonstrates the absence of an effect of rIL-22 into TCR-δ−/−δ−/− mice. In brief, total splenocytes were obtained by pushing spleens through a 70-µm pore-size mesh (Falcon) with subsequent resuspension in RBC lysis buffer (eBioscience). Cells were resuspended in buffer and biotinylated antibody cocktail, mixed, and incubated at 4°C for 10 min. Anti-biotin microbeads were added per the manufacturer’s instructions, mixed, and incubated for 15 min at 4°C. The cells were washed and applied to the column per the manufacturer’s instructions. The effluent, which contained the unlabeled CD4+ or CD8+ T cells, was collected and passed over a second column to further enrich each T cell population. Cell counts for CD4+ and CD8+ T lymphocytes were determined by Coulter counter (Beckman Coulter).

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SUPPLEMENTAL MATERIAL

Simonian et al., http://www.jem.org/cgi/content/full/jem.20100061/DC1

Figure S1. Representative density plots of intracellular IL-22 and IL-17A expression in γδ T cells isolated from the spleen of homozygous transgenic Vγ6/Vδ1 (B6.Vγ6++) treated with B. subtilis for 4 wk. γδ T cells expressing IL-22 and IL-17A were selected from a bivariate dot plot of CD3+CD8+ cells from the lymphocyte gate. The percentage of γδ T cells expressing each cytokine is shown. Data represent at least five individual mice per experiment from at least three experiments.

Figure S2. Total cell and absolute lymphocyte counts performed on lung homogenates from WT C57BL/6 mice treated with a combination of recombinant IL-22 and B. subtilis or vehicle and B. subtilis for four consecutive weeks. Data were compiled from at least five mice per experiment from two separate experiments (mean ± SD).
Figure S3. Levels of CXCL10 in the lung of B6.TCRδ−/− mice treated with rIL-22 and B. subtilis for four consecutive weeks compared with PBS control. The mean is shown as a solid line. Data are compiled from five individual from at least two separate experiments.

Figure S4. Similar clearance of B. subtilis from the lung of AhRΔ/Δ and WT C57BL/6 mice. Individual AhRΔ/Δ and WT C57BL/6 mice were treated with B. subtilis for 4 wk. Colony counts from lung homogenates of individual mice were analyzed for bacterial persistence at different time points after last exposure to B. subtilis. Data are compiled from at least five individual mice from at least two separate experiments (mean ± SD).