Mice Transgenic for BAFF Develop Lymphocytic Disorders Along with Autoimmune Manifestations

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Summary

The cause of many autoimmune and inflammatory diseases is unresolved, although dysregulated production of tumor necrosis factor (TNF) family members appears to be important in many cases. BAFF, a new member of the TNF family, binds to B cells and costimulates their growth in vitro. Mice transgenic for BAFF have vastly increased numbers of mature B and effector T cells, and develop autoimmune-like manifestations such as the presence of high levels of rheumatoid factors, circulating immune complexes, anti–DNA autoantibodies, and immunoglobulin deposition in the kidneys. This phenotype is reminiscent of certain human autoimmune disorders and suggests that dysregulation of BAFF expression may be a critical element in the chain of events leading to autoimmunity.

Key words: autoantibodies • tumor necrosis factor–related • B cell • rheumatoid factors • transgenic

Self-reactive B cells constantly emerge during lymphopoiesis. Fortunately, the immune system has several powerful means of eliminating or neutralizing autoreactive B cells, both centrally as they emerge from the stem cell pool and peripherally as mature B cells undergo somatic mutation (1, 2). Despite the diversity of systems that allow discrimination between self and nonself, failure to eliminate autoreactive lymphocytes occurs in many cases. The Fas/FasL system is probably the most striking example of a pathway directly involved in the elimination of some subsets of autoreactive B and T lymphocytes by apoptosis. Mice lacking functional expression of Fas (lpr) or FasL (gld) develop severe autoimmune lymphoproliferative disorders, leading to tissue destruction (13). Another member of the TNF superfamily, CD30L, seems to have a role of protection against autoimmunity, as signaling through its receptor was shown to protect against autoimmune diabetes mediated by CD8 T cells (14). More often, a reverse situation is seen by which abnormal levels rather than absence of expression directly contributes to the pathophysiology of autoimmune diseases (12). TNF is the most extensively studied member of this family, and its participation in several autoimmune disorders such as rheumatoid arthritis, inflammatory bowel disease, and autoimmune encephalomyelitis is now well established (15). Participation or abnormal expression in autoimmune disease has been described for other ligands of this family such as OX40 (16), CD27 (17), and lymphotoxin (18).

Early studies on the CD40/CD40L pathway revealed specific roles in B cell biology (19) promoting B cell activation, proliferation, survival, isotype switching, germinal center formation, and generation of memory B cells (20). It was, therefore, not surprising that interrupting this pathway inhibited disease in rodent models of rheumatoid arthritis and systemic lupus nephritis, in which B cell tolerance is severely altered (21, 22). Recent studies have shown that the role of the CD40/CD40L pathway in disease is not solely directed towards B cell regulation, but extends to many other cell types (20).

We have recently identified a new TNF-like ligand, BAFF, a new member of the TNF family, binds to B cells and costimulates their growth in vitro. Mice transgenic for BAFF have vastly increased numbers of mature B and effector T cells, and develop autoimmune-like manifestations such as the presence of high levels of rheumatoid factors, circulating immune complexes, anti–DNA autoantibodies, and immunoglobulin deposition in the kidneys. This phenotype is reminiscent of certain human autoimmune disorders and suggests that dysregulation of BAFF expression may be a critical element in the chain of events leading to autoimmunity.
called BAFF (for B cell activating factor belonging to the TNF family), which is expressed by dendritic cells and possibly T cells and binds primarily to B cells (23). BAFF is secreted from transfected cells and has the potential to act as a soluble mediator. BAFF was also independently identified as TALL-1 (24), as THAN K, which regulated apoptosis, NF-κB and c-Jun NH₂-terminal kinase in a human myelocytic cell line (25), and, more recently, as BlyS, a factor that, when administrated to normal mice, disrupted splenic B and T cell zones and resulted in elevated serum IgM concentrations (26). Similar to CD40L, BAFF promotes B cell proliferation in the presence of an anti–IgM antibody in vitro (23, 26). We have generated transgenic mice overexpressing BAFF under the control of a liver-specific promoter. These mice have excessive numbers of mature B cells, spontaneous germinal center reactions, secrete autoantibodies, high plasma cell numbers in secondary lymphoid organs, elevated numbers of effector T cells, and Ig deposition in the kidney.

**Materials and Methods**

Generation of BAFF Transgenic Mice. A PCR fragment encoding full-length murine BAFF was generated by reverse transcription-PCR using previously described sequence information (23). First strand cDNA was synthesized from mouse lung polyA+ (Clontech) using oligo d(T) according to the manufacturer’s protocol (GIBCO BRL). The PCR reaction contained 1× pfu buffer (Stratagene Inc.), 0.2 mM dNTPs, 10% DMSO, 1.25 μM primers, 5 units pfu enzyme (Stratagene Inc.), and the following primers with NotI restriction sites 5′-TAAGAATGCGGCAGCGAATG-TGATGAGTCTGCAAA-3′ and 5′-TAAGAATGCGGCAGCGAATG-GGATCAGCTGCAAA-3′. The template was amplified for 30 cycles at 94°C for 1 min, 54°C for 2 min, and 72°C for 3 min, followed by a 10-min extension at 72°C. This sequence corresponds to nucleotides 214-1171 of the GenBank file AF119383. The PCR fragment was digested with NotI and cloned into a modified pCEP4 vector (Invitrogen Corp.). The resulting vector was then digested with XbaI to remove BAFF plus the SV40 polyA addition site sequence. This fragment was cloned into a pUC-based vector in which the promoter, a 1-kb blunt BglII–NotI fragment containing the human ApoE enhancer and AAT (alpha antitrypsin) previously purified from the plasmid clone 5408 (a gift from Dr. Katherine Parker Ponder, Washington University, St. Louis, MO) was further inserted at the EcoRV site. An EcoRV/BglII fragment was purified from the final vector and used for the generation of transgenic mice. The injected offspring of C57BL/6 female × DBA/2 male F1 (BDF1) mice were backcrossed onto C57BL/6 mice. Techniques of microinjection and generation of transgenic mice have been previously described (27). Animal experiments were approved by the Institutional Animal Care and Use Committee.

A natalialy Methods. Serum samples were subjected to reducing SDS-PAGE analysis using a linear 12.5% gel. MOPC–21 mouse IgG1 standard antibody was obtained from Pharmingen. Total RNA from mouse liver was prepared and processed for Northern blot analysis using an isolation kit from Promega Corp. according to the manufacturer’s guidelines. BAFF transgene-specific mRNA was detected using a probe spanning the SV40 polyA tail of the transgene construct and obtained by digestion of the modified pCEP4 vector with XbaI and BamHI. The probe recognizes a 1.8–2.0-kd band corresponding to mRNA from the BAFF transgene. PCR analysis of total DNA from BAFF transgenic (Tg) mice used 12.5 pM of the primers 5′-GCAGTTTCCACACCGAT-GTCTC-3′ and 5′-GTCTCGTGGCGGAATACTG-3′ in a reaction containing 1× Taq polymerase buffer (Stratagene Inc.), 0.2 mM dNTPs, 10% DMSO, and 5 U Taq polymerase (Stratagene Inc.). 719 bp of the transgene was amplified for 35 cycles at 94°C for 30 s, 54°C for 1 min, and 72°C for 1.5 min, followed by a 10-min extension at 72°C.

The presence of proteins in mouse urine was measured using Mablitext 10 SG reagent strips for urinalysis (Bayer Corp., Diagnostics Division).

Cellular Analysis. Differential white blood cell counts of fresh EDTA-anticoagulated whole blood were performed with an Abbott Cell Dye 3500 apparatus. For FACS® analysis (Becton Dickinson & Co.), fluorocytin–FITC, Cychrome™–, and phycoerythrin (PE)–labeled rat anti–mouse antibodies: anti–B220, anti–CD4, anti–CD8, anti–CD43, anti–IgM, anti–CD5, anti–CD25, anti–CD24, anti–CD38, anti–CD21, anti–CD44, anti–M HC class II, anti–i–selectin, and hamster anti–Bcl-2 control hamster Ig kit were purchased from PharMingen. Production of recombinant E. coli, as well as mammalian cell-derived mouse Flag-tagged BAFF, was performed as previously described for human BAFF (23). All antibodies were used according to the manufacturer’s specifications. PBL were isolated by density gradient centrifugation of EDTA-treated mouse blood over lymphocyte M (Cedarlane). FACS® analysis of spleen, bone marrow, and mesenteric lymph nodes was performed as described previously (28).

Detection of Total Mouse Ig and Rhuematoit Fators in Mouse Sera by ELISA Assays. ELISA plates (Corning Glass Works) were coated overnight at 4°C with a solution of 10 μg/ml goat anti–mouse Ig (Southern Biotechnology Associates, Inc.) in 50 mM sodium bicarbonate buffer, pH 9.6. Plates were washed three times with PBS/0.1% Tween and blocked overnight with 1% gelatin in PBS. 100 μl/well of serum serial dilutions or standard dilutions was added to the plates for 30 min at 37°C. Mouse Ig was detected using 100 μl/well of a 1:μg/ml solution of an alkaline phosphatase (AP)–labeled goat anti–mouse Ig (Southern Biotechnology Associates) for 30 min at 37°C. A 1:μg/ml solution of mouse Ig purchased from Southern Biotechnology Associates, Inc. instead of goat anti–mouse Ig, and detection of mouse Ig was performed as described above. Detection of mouse isotypes in the R F assay was done using AP-labeled goat anti–mouse IgA, IgM, and IgG2a, IgG2b, IgG1, and IgG3, as well as purified mouse IgA, IgM, IgG2a, IgG2b, IgG1, and IgG3 for standard curves (Southern Biotechnology Associates, Inc.). All statistical comparisons were performed by analysis of variance.

Detection of Circulating Immune Complexes and Precipitation of Cryoglobulins in Mouse Sera. The assay was performed as previ-
An ELISA assay for murine BAFF is not available; however, we found that 2% serum from BAFF-Tg mice, but not from control mice, blocked the binding of mammalian cell-derived mouse soluble Flag-tagged BAFF to BJAB cells. Moreover, 5% serum from BAFF-Tg mice but not from control mice increased the proliferation of human B cells from PBL in the presence of anti-μ (data not shown). These data suggest that soluble BAFF is present in the blood of BAFF-Tg mice. In all BAFF-Tg mice examined histologically, the livers showed no abnormalities, indicating that local overexpression of BAFF did not induce any immunological or pathological events (data not shown).

Expansion of the Peripheral Blood B Cell Compartment in BAFF-Tg Mice. The transgenic mouse population was found to have more lymphocytes in the blood when compared with control negative littersmates, reaching values as high as 13,000 lymphocytes/μl of blood (Fig. 1 A). In contrast, the number of granulocytes per microliter of blood in both BAFF-Tg and control mice remained within normal limits (Fig. 1 A). The elevated lymphocyte levels resulted from an expanded B cell subset, since FACS® analysis using anti-CD4 and -B220 antibodies, of peripheral blood cells (PBL) from 18 BAFF-Tg mice issued from six different founders showed increased B/T ratios (Fig. 1, B and C). Likewise, combining the number of lymphocytes per microliter of blood with the percentage of circulating T cells, calculation of absolute numbers of CD4 circulating T cells revealed a 50% reduction of this T cell subset in BAFF-Tg mice when compared with control mice, and the same observation was made for the CD8 T cell subset (data not shown). All peripheral blood B cells from BAFF-Tg mice had increased MHC class II and Bcl-2 expression when compared with B cells from control mice (Fig. 1, D and E, respectively), indicating some level of B cell activation in PBL of BAFF-Tg mice. T cells in the blood of BAFF-Tg mice did not express the early activation markers CD69 or CD25; however, 40–56% of CD4 or CD8 T cells were activated effector T cells with a CD44ဖျက်ဆွေးနွဲ့ − soluble mAb 205, which is specific for both ss- and dsDNA (31).

Immunohistochemistry. Frozen sections of spleen and lymph nodes were subjected to immunohistochemical analysis as previously described (32). Biotin-labeled antibodies rat anti-B220, anti-CD11c, and anti-syndecan-1, as well as unlabeled rat anti-CD4, anti-CD8α, and anti-CD8β were purchased from Phar-Mingen. Biotin-labeled peanut agglutinin was obtained from Mingen. Biotin-labeled peanut agglutinin was obtained from Vector Laboratories, Inc. Horseradish peroxidase (HRP)–labeled mouse anti-rat Ig and HRP-streptavidin were purchased from Jackson Immunoresearch Laboratories, Inc., and AP-labeled streptavidin from Southern Biotechnology Associates, Inc. In the case of immunohistochemistry on kidney tissue to detect Ig deposition, paraffin sections were used, dewaxed, and blocked using diluted horse serum from Vector Laboratories, Inc., followed by staining with HRP goat anti-mouse Ig from Jackson Immunoresearch Laboratories, Inc. Detection was performed as previously described for frozen sections (32).

Results

BAFF-Tg Mice. Full-length murine BAFF was expressed in transgenic mice using the liver-specific alpha-1 antitrypsin promoter with the APO E enhancer. The full-length version was chosen with the expectation that BAFF would be cleaved and act systemically or, if retained in a membrane-bound form, that local liver-specific abnormalities would be observed, possibly providing functional clues. We obtained 13 founder mice positive for the BAFF transgene. BAFF overexpression in the liver of transgenic mice was confirmed by Northern blot analysis (data not shown). An ELISA assay for murine BAFF is not available; however, we found that 2% serum from BAFF-Tg mice, but not from control mice, blocked the binding of mammalian cell-derived mouse soluble Flag-tagged BAFF to BJAB cells. Moreover, 5% serum from BAFF-Tg mice but not from control mice increased the proliferation of human B cells from PBL in the presence of anti-μ (data not shown). These data suggest that soluble BAFF is present in the blood of BAFF-Tg mice. In all BAFF-Tg mice examined histologically, the livers showed no abnormalities, indicating that local overexpression of BAFF did not induce any immunological or pathological events (data not shown).

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in all seven mice analyzed. These data indicate that over-expression of BAFF is affecting the mature B cell compartment in the periphery, but not progenitor B cells in the BM.

We calculated the total number of B and T cells in the spleen, BM, and MLN (Table I). The total number of B cells was at least sevenfold higher in the spleen and MLN of BAFF-Tg mice. The total number of T cells was also increased twofold in the spleen and MLN of these mice (Table I). Total numbers of B and T cells in the BM in BAFF-Tg mice were similar to that of control mice (Table I). The population of CD5⁺ B1 cells in the spleen, BM, and peritoneal lavages of BAFF-Tg mice was similar to that of control mice and only marginally increased in MLN (data not shown).

Analysis by FACS® of B cell subpopulations in the spleen revealed an increased proportion of marginal zone (MZ) B cells in BAFF-Tg mice when compared with control mice (Table II). The population of follicular B cells remained equivalent in both BAFF Tg and control mice, whereas the fraction of newly formed B cells was slightly decreased in BAFF-Tg mice (Table II). This result was also confirmed.
on B220+ splenic B cells using anti-CD38 vs. anti-CD24 antibodies or anti-IgM vs. anti-IgD antibodies and analyzing the CD38 hi/CD24+ and IgM hi/IgD lo MZ B cell population, respectively, as previously described (33) (data not shown). Immunohistochemical analysis using an anti-mouse IgM antibody revealed the expansion of the IgM-bright MZ B cell area in the spleen of BAFF-Tg mice when compared with control mice (data not shown). All BAFF-Tg
Figure 2. Increased proportion of B cells in the spleen, MLN, but not in the bone marrow of BAFF-Tg mice. (A) FACS® staining for mature B cells using both anti-IgM-FITC and anti–B220-PE, in spleen (top), bone marrow (middle), and MLN (bottom). Percentages of B220⁺/IgM⁺ mature B cells are indicated. (B) FACS® staining for pre–B cells (B220⁺/CD43⁻) and pro–B cells (B220⁺/CD43⁺) in the bone marrow using anti–CD43-FITC, anti–B220-CyChrome™, and anti–IgM-PE simultaneously. Shown are cells gated on the IgM negative population. Percentages of pre–B cells (B220⁺/CD43⁻) and pro–B cells (B220⁺/CD43⁺) are indicated. For all panels (A and B), one control mouse (left) and two BAFF-Tg mice (right) are shown and results are representative of seven animals analyzed for each group.
B220⁺ splenic B cells also expressed higher levels of Bcl-2 (data not shown) and also MHC class II (Table II) compared with splenic B cells from control mice, indicating that splenic B cells as well as peripheral blood B cells are in an activated state. At equal cell concentration, splenocytes isolated from BAFF-Tg mice survived longer in culture medium when compared with control splenocytes and the thymidine incorporation after 6 d of culture relative to the incorporation on day 0 was only decreased 20% in cultures with BAFF-Tg-derived splenocytes vs. 60% reduction with control splenocytes (data not shown). In vivo 5-bromo-2'-deoxyuridine (BrdU) incorporation for 4 d did not reveal any higher BrdU intake in BAFF Tg-splenic B cells when compared with control mice (data not shown).

### Table I. Total Number of T and B Cells in the Spleen, Mesenteric Lymph Nodes, and Bone Marrow

<table>
<thead>
<tr>
<th>Mouse number</th>
<th>Spleen (×10⁷)</th>
<th>BM * (×10⁷)</th>
<th>M LN (×10⁷)</th>
<th>Spleen (×10⁷)</th>
<th>BM * (×10⁷)</th>
<th>M LN (×10⁷)</th>
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<tbody>
<tr>
<td>BAFF-Tg mice</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>816-8-34</td>
<td>18</td>
<td>4</td>
<td>1.4</td>
<td>3.5</td>
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<td>802-43</td>
<td>34</td>
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<td>5.6</td>
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<tr>
<td>823-3-23</td>
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<td>3.1</td>
<td>3</td>
<td>3.8</td>
<td>0.46</td>
<td>3.6</td>
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<tr>
<td>802-41</td>
<td>46</td>
<td>3</td>
<td>5</td>
<td>5.3</td>
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<tr>
<td>816-1-29</td>
<td>23</td>
<td>1.4</td>
<td>1.2</td>
<td>4</td>
<td>0.07</td>
<td>0.3</td>
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<tr>
<td>Mean ± SD</td>
<td>27.4 ± 11</td>
<td>2.6 ± 1.04</td>
<td>3.2 ± 1.8</td>
<td>4 ± 0.6</td>
<td>0.28 ± 0.14</td>
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<td>1.5</td>
<td>0.23</td>
<td>0.3</td>
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<tr>
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<td>0.17</td>
<td>1.9</td>
<td>0.21</td>
<td>0.32</td>
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<tr>
<td>Mean ± SD</td>
<td>4.7 ± 1.5</td>
<td>2.2 ± 0.8</td>
<td>0.19 ± 0.08</td>
<td>2.2 ± 0.6</td>
<td>0.16 ± 0.05</td>
<td>0.31 ± 0.02</td>
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<td>Controls/BAFF-Tg</td>
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<tr>
<td>P values</td>
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<td>P &gt; 0.1</td>
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<td>P &lt; 0.01</td>
<td>P &gt; 0.1</td>
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*Bone marrow was extracted from two femurs. P values were obtained using ANOVA.

### Table II. Increased MHC Class II Expression on B Cells and Enlarged Proportion of MZ B Cells in the Spleen of BAFF-Tg Mice

<table>
<thead>
<tr>
<th>Levels of MHC class II expression on B220⁺ B cells (MFI)</th>
<th>Percentage of follicular B cells*</th>
<th>Percentage of MZ B cells†</th>
<th>Percentage of newly formed B cells‡</th>
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<tbody>
<tr>
<td>Control mice</td>
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<tr>
<td>816-1-10</td>
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<tr>
<td>816-1-1</td>
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<td>40</td>
<td>23</td>
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Splenocytes were analyzed by FACS® and gated on the B220⁺ population. A representative experiment is shown. MFI, mean of fluorescence intensity.

*B220⁺/IgMlo/CD21int.
†B220⁺/IgMhi/CD21hi.
‡B220⁺/IgMlo/CD21lo.
teriolar lymphocyte sheath (or T cell area) in BAFF-Tg mice (Fig. 4 B). Interestingly, few germinal centers were observed in nonimmunized control littermates (and is typical of this colony in general) and those present were small (Fig. 4 C), whereas BAFF-Tg mice possessed numerous and large germinal centers in the absence of immunization (Fig. 4 D). Staining with anti-CD11c for dendritic cells in the T cell zone and the marginal zone of control mice (Fig. 4 E) was considerably reduced in BAFF-Tg mice (Fig. 4 F). Syndecan-1-positive plasma cells were almost undetectable in the spleen from control littermates (Fig. 4 G), yet the red pulp of BAFF-Tg mice was strongly positive for syndecan-1 (Fig. 4 H). Very similar observations were made with MLN (Fig. 5). In the MLN of BAFF-Tg mice, the B cell areas were dramatically expanded (Fig. 5 B), in contrast to the normal appearance, where B cell follicles were easily recognizable at the periphery of the node under the capsule with a typical paracortical T cell zone (Fig. 5 A). The medulla of MLN from BAFF-Tg mice were filled with syndecan-1-positive plasma cells that presumably are plasma cells (Fig. 5 H). Therefore, analysis of secondary lymphoid organs in BAFF-Tg mice was consistent with the expanded B cell compartment seen by FACS® analysis and indicates multiple cellular abnormalities and intense immune activity.
BAFF-Tg Mice Have High Levels of Total Immunoglobulins, Rheumatoid Factors, and Circulating Immune Complexes in Their Serum.

The increased B cell compartment in BAFF-Tg mice suggested that the level of total Ig in the blood of these animals might also be increased. SDS-PAGE analysis of the serum showed that IgG levels were elevated in all BAFF-Tg mice, while the nontransgenic littermates displayed a normal pattern of serum proteins (Fig. 6 A). By comparison with an IgG1 standard antibody, the levels of IgG in a nontransgenic mouse were ±5–8 mg/ml, and these levels increased to at least 50 mg/ml in some BAFF-Tg mice (quantification was done with underloaded gels). In normal mice, the light chain band is smeared due to the polyclonal nature of the Ig and on this basis the elevated Ig levels in BAFF-Tg mice were also polyclonal in nature. IgM levels were visibly increased in these mice, albeit not as much as IgG, and this band is seen as a smear on top of a transferrin band in this gel.

High serum Ig levels in BAFF-Tg mice were confirmed by ELISA (Fig. 6 B), and the high levels of Ig seen in these mice led us to suspect the presence of rheumatoid factors, or autoantibodies directed against antigenic determinants on the Fc domain of IgG (34). These antibodies could bind to the goat anti-mouse Ig used to coat the ELISA plates and give erroneously high values. ELISA plates were, therefore, coated with normal goat Ig and the binding of BAFF Tg Ig to normal goat Ig was measured. Fig. 6 C shows that sera from most BAFF-Tg mice contained Ig reacting with normal goat Ig, whereas only 2 of 19 control mice exhibited reactivity in the same assay. These RF were mainly of the IgM, IgA, and IgG2a isotypes (data not shown).

Presence of RF can be associated with the presence of high levels of circulating immune complexes (CIC) and cryoglobulin in the blood (34). To verify whether or not BAFF-Tg mice have abnormal serum levels of CIC, a C1q-based binding assay was used to detect CIC in the 21 BAFF-Tg mice analyzed above. Only five BAFF-Tg mice showed significantly high levels of CIC compared with the control mice; nonetheless, these mice corresponded to the animals having the highest total serum Ig and rheumatoid factor levels (Fig. 6 D). We also observed precipitate formation when sera from BAFF-Tg mice, but not control sera, were diluted 1/15 in water, indicating the presence of cryoglobulin in these mice (data not shown). Thus, in addition to B cell hyperplasia, BAFF-Tg mice display severe hypergobulinemia associated with the presence of RF and CIC.

Some BAFF-Tg Mice Have High Levels of Anti–ss and –dsDNA Autoantibodies, Ig Deposition in the Kidneys, and Proteinuria. Initially, we observed kidney abnormalities reminiscent of a lupus-like disease in two of our founder mice. The presence of anti–DNA autoantibodies has also been described in SLE patients or the SLE-like (SWR × NZB)F1 (SNF1) mouse (31). Anti–ssDNA autoantibody levels were detected in BAFF-Tg mice previously shown to have the highest level of total serum Ig (Fig. 7 A). We analyzed the serum of two BAFF-Tg mice negative for antibodies against ssDNA (697-5 and 816-1-1) and three transgenic mice secreting anti–ssDNA antibodies (820-14, 816-8-3, and 820-7) for the presence of anti–dsDNA antibodies in parallel with five control littermates. BAFF-Tg mice also secreted anti–dsDNA; however, the levels of secretion did not always correlate with that of anti–ssDNA antibodies, as serum from BAFF-Tg mouse 697-5, which did not contain detectable levels of anti–ssDNA antibodies, was clearly positive for the presence of anti–dsDNA antibodies (Fig. 7 B). Therefore, BAFF-Tg mice showing the most severe hyperglobulinemia secrete high levels of anti–DNA autoantibodies. Additionally, and also reminiscent of lupus-like nephritis, we detected immunoglobulin deposition in the kidney of six BAFF-Tg mice analyzed (Fig. 7 C), three of these mice did not secrete detectable levels of anti–DNA antibodies (data not shown). All BAFF-Tg mice have proteinuria (Table III). Sera from all BAFF-Tg mice, but not control mice, diluted 1/100, stained the nuclei of lymph node cells on tissue sections, indicating the presence of antinuclear antibodies in the serum of BAFF-Tg mice (data not shown).
Discussion

BAFF is a powerful cytokine affecting B cells, and has consequences for T cell and dendritic cell status. The nature of the expanded B cell subset in BAFF-Tg mice is still unclear, but seems to be restricted to mature B cells that have been activated. Overexpression of BAFF led to the emergence of autoimmune manifestations such as production of autoantibodies, proteinuria, Ig deposition in kidneys, and intense germinal center formation. Thus, BAFF ligand and its receptor on B cells form a novel immunoregulatory system.

Whether a ligand is secreted or membrane-bound has profound biological ramifications. These mice were designed to express high levels of BAFF in the liver and, while it cannot be excluded that low level of expression somewhere in the immune system accounts for this unusual biology, we view it more likely that BAFF is secreted from the liver and acts at a distant site. We have indirect evidence for the presence of BAFF in the serum of transgenic mice and, moreover, injection of recombinant BAFF in normal mice led to some of the effects described here (26). Well-defined secretion of a TNF family ligand with functional consequences in vivo has been observed only infrequently; e.g., TNF and lymphotoxin-α. BAFF, TWEAK, and APRIL are three relatively new ligands that possess canonical furin cleavage motifs in the stalk region and are readily secreted from transfected cell systems (23, 35, 36). Whether such secretion in vitro actually predicts for a soluble ligand system is not clear, yet this BAFF-Tg mouse would indicate that secretion can occur at least from the liver and, thus, soluble BAFF ligand is capable of mediating biological events in vivo. Alternatively, facile cleavage may represent a mechanism that ensures a very transient localized signaling event.

The in vitro analysis using recombinant soluble BAFF protein showed that BAFF costimulated B cell growth in conjunction with B cell receptor activation, yet by itself it did not stimulate proliferation of resting B cells (23). If BAFF is truly a soluble mediator, then this observation is similar to that made originally for IL-2 and T cell growth, and prompts the question of whether BAFF is a B cell growth factor. The present data do not allow one to distinguish between a costimulatory action (e.g., analogous to the activity of CD28) and a true B cell growth factor-like activity. Regardless of the mechanism, these data suggest that expansion of the B cell compartment in these mice is the result of BAFF-induced proliferative stimuli, yet negative results with 4 d in vivo BrdU incorporation and increased ex vivo survival of splenocytes raised the possibility of dilution of the sera giving an OD three times higher than that of background. The quantity of CIC is defined as the quantity of peroxidase-mouse antiperoxidase required to generate an OD equivalent to that obtained with the tested serum. The difference between control animals and BAFF-Tg mice was statistically significant (P < 0.001 in B and C, P < 0.003 in D).
that this observation stems from an increased output from the bone marrow and/or a decreased death rate.

The CD40 pathway clearly plays a major role in B cell regulation, inviting a comparison with the BAFF system. An increase in the size of the B cell population, enlarged spleens, lymph nodes, and Ig deposition in the kidney were also observed in CD40L-Tg mice (37). Several aspects clearly distinguish these two mice; for example, CD40L-Tg mice develop inflammatory bowel disease (IBD), which was not observed in BAFF-Tg mice, and the alterations in the organization of the secondary lymphoid organs are very different (37). In CD40L-Tg mice, but not BAFF-Tg mice, the organization of the thymus is altered, which presumably impairs proper T cell selection leading to IBD, as seen in other mouse models with thymic dysfunction (38). The difference between these two transgenic mice may

Figure 7. Presence of anti–ssDNA and –dsDNA autoantibodies in some BAFF-Tg mice, and Ig deposition in the kidneys. (A) Analysis by ELISA of anti–ssDNA autoantibodies in 19 control littersates (left) and 21 BAFF-Tg mice (black bars). (B) Analysis by ELISA of anti–ssDNA autoantibodies in five control littersates and the five animals showing levels of anti–ssDNA autoantibodies from A (black bars). (C) Paraffin sections of kidneys from a control mouse (left) and a BAFF-Tg mouse (right), stained with goat ant–mouse Ig HRP. Ig deposition is shown by a brown staining. These pictures are representative of six BAFF-Tg mice analyzed. Original magnifications.
be due in large part to the distinct distribution of these two ligands and their corresponding receptors. Transgene-expressed CD40L is a membrane-bound ligand primarily expressed on thymocytes and activated T cells, whereas transgenic liver-expressed BAFF has the characteristic of a soluble ligand and, therefore, can diffuse into multiple compartments. CD40 is expressed on a wide variety of cell types (20), whereas expression of BAFF receptor is restricted to B cells and possibly monocytes cells (23–26). Given the disparate phenotypes of the BAFF- and CD40L-Tg mice, it is fair to predict that BAFF and CD40L probably play distinct roles in normal animals as well. Our results on splenic architecture and elevated Ig levels in the serum of BAFF-Tg mice are consistent with those described in a recent study using short-term injection of soluble BAFF in normal mice (26) and tend to minimize possible developmental disturbances related to the expression of the BAFF transgene in our mice. However, chronic exposure of the transgenic mice to BAFF led to changes not paralleled in mice injected for 4 d with recombinant BAFF (26). In both cases, serum IgM levels were elevated, yet the BAFF-Tg mice exhibited vastly increased IgG and IgA levels. The effects of short-term injections were interpreted as possibly stemming from activation of T-independent B cell events, whereas the elevated IgG and IgA levels, the presence of non-IgM RF isotypes, and the extensive germinal center formation clearly indicate ongoing T cell-dependent B cell events.

Table III. Levels of Proteinuria* in BAFF-Tg Mice

<table>
<thead>
<tr>
<th>Control littermates</th>
<th>BAFF-Tg mice</th>
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<tbody>
<tr>
<td>816-1-20</td>
<td>-</td>
</tr>
<tr>
<td>802-51</td>
<td>+</td>
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<tr>
<td>823-2-3</td>
<td>+/-</td>
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<td>823-2-6</td>
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<td>816-1-24</td>
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<tr>
<td>823-14-19</td>
<td>+/-</td>
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<tr>
<td>816-6-4</td>
<td>+</td>
</tr>
</tbody>
</table>

*Proteinuria was measured using medical color strips dipped in mouse urine and is defined as follows: -, no proteinuria; +/, trace; +, 30 mg/dl; ++, 100 mg/dl; ++++, 300 mg/dl; ++++, >2,000 mg/dl.

Among the increased B cell populations in BAFF-Tg mice are emerging autoreactive B cells, secreting RF and anti-DNA autoantibodies. It is well known that tolerance to self antigens is never complete and autoreactive B cells, as well as low levels of rheumatoid factors, can be detected in normal individuals (3). These autoreactive B cells are referred to as natural autoreactive B cells. Therefore, one possibility in BAFF-Tg mice is that the emergence of a large number of autoreactive B cells may reflect the expansion of occasional natural autoreactive B cells in response to BAFF-proliferative stimuli. If this was the case, one would predict that only higher levels of IgM RF would be detected (34), yet high levels of IgA and IgG2a RF were observed indicating isotype switching in the RF-specific B cells. RF other than IgM are found in patients with autoimmune diseases such as rheumatoid arthritis (34). Therefore, the population of RF-specific autoreactive B cells in BAFF-Tg mice is not only expanded but also activated, indicating a dynamic antigen-specific process leading to autoimmune manifestations rather than passive expansion. It cannot be excluded that BAFF-Tg mice have larger numbers of nonnatural self-reactive B cells that have not been identified or possibly larger numbers of foreign antigen-specific activated B cells.

A number of studies have shown that the escape of autoreactive B cells from clonal deletion or functional inactivation (clonal anergy) alone is not enough to develop autoimmune disease (1, 41). Additional factors such as infection, cytokines, and costimulatory help from T cells are required. The presence of large germinal centers in secondary lymphoid organs of BAFF-Tg mice, higher total T cell numbers in the spleen and MLN, as well as the increased proportion of both CD4 and CD8 effector T cells in the periphery, and the quality of the RF isotypes strongly suggest the active participation of T cells in the immune reactions triggered in BAFF-Tg mice. Whether the enlarged population of activated effector T cells contains autoreactive T cells remains to be determined. One may question...
why BAFF-Tg mice do not show more severe pathological manifestations. Potential explanations include the expression of nonlethal BAFF levels in surviving BAFF-Tg founders, the absence of either pathogenic B cells or tissue-destructing antibodies as seen in some murine models of lupus and rheumatoid arthritis, respectively (6, 42) and, finally, the H-2b background of the BAFF-Tg mice, which may not favor the emergence of severe autoimmune manifestations.

If overexpression of BAFF indeed initiates an active autoimmune reaction in our transgenic mice, we need to question why tolerance to self has failed. Downregulation of Bcl-2 expression in autoreactive B cells has been shown to be one way to sensitize these cells to cell death signals (43). B cells in BAFF-Tg mice express higher levels of Bcl-2, indicating a possible protection against apoptotic signals, and also suggesting that BAFF, like CD40L, provides survival signals to B cells. One can speculate that this event coupled with a BAFF proliferative signal may explain the accumulation of autoreactive B cells in these mice. However, these changes alone are probably not sufficient to generate an autoreactive response and one potential answer may reside in the role of T cells. Increased numbers of effector T cells were directly observed in the periphery and there was an apparent reduction in the numbers of dendritic cells in the spleen of BAFF-Tg mice. Dendritic cells are believed to be essential for T cell tolerance to self (44), and their deficit in BAFF-Tg mice may promote the emergence of autoreactive T cells. Thus, we hypothesize that the role of BAFF overexpression in impairing self-tolerance may rely on two mechanisms: promoting enhanced survival and proliferation of activated autoreactive B cells and suppression of the protective effects of dendritic cells against the emergence of autoreactive T cells.

These experiments demonstrate that ectopic overexpression of BAFF was sufficient to initiate the expansion of the mature B cell compartment, resulting in lupus-like autoimmune manifestations. This transgenic mouse model potentially brings new insight into the etiology of autoimmune disorders, provides a novel framework for the investigation of autoreactivity, and potentially opens the door to new therapeutic strategies both for the treatment of some autoimmune disorders and the stimulation of humoral responses.

References