Brief Definitive Report

Plasmodium falciparum Produces Prostaglandins that are Pyrogenic, Somnogenic, and Immunosuppressive Substances in Humans

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Summary

Plasmodium falciparum causes the most severe form of human malaria, which kills ~1.5–2.7 million people every year, but the molecular mechanisms underlying the clinical symptoms and the host–parasite interaction remain unclear. We show here that P. falciparum produces prostaglandins (PGs) D₂, E₂, and F₂α. After incubation with 1 mM arachidonic acid (AA), cell homogenates of P. falciparum produced PGs as determined by enzyme immunoassay and gas chromatography–selected ion monitoring. PG production in the parasite homogenate was not affected by the nonsteroidal antiinflammatory drugs aspirin and indomethacin, and was partially heat resistant, whereas PG biosynthesis by mammalian cyclooxygenase was completely inhibited by these chemicals and by heat treatment. Addition of AA to the parasite cell culture markedly increased the ability of the parasite cell homogenate to produce PGs and of parasitized red blood cells to accumulate PGs in the culture medium. PGD₂ and PGE₂ accumulated in the culture medium at the stages of trophozoites and schizonts more actively than at the ring stage. These findings are the first evidence of the direct involvement of a malaria parasite in the generation of substances that are pyrogenic and injurious to the host defenses. We will discuss a possible contribution of the parasite-produced PGs to pathogenesis and host–parasite interaction of P. falciparum.

Key words: Plasmodium falciparum • prostaglandin • malaria pathogenesis • tumor necrosis factor α • pyrogen

Malaria symptoms in an acute infection are general inflammatory responses, including periodic fever with shivering, headache, body pains, sleepiness, and loss of appetite, that are caused by the proliferating malaria parasite cells at the erythrocytic stage. During acute blood stage malaria, suppression of both T and B cell–mediated immune responses is a common occurrence (1–3), although the corresponding mechanism is not known, suggesting that malaria parasites have certain strategies for evasion of host defenses to establish the infection. Several papers have reported enhancement of the TNF-α level in malaria patient sera infected with Plasmodium falciparum and Plasmodium vivax, suggesting that TNF-α acts as an endogenous pyrogenic substance (4–6).

Prostaglandins (PGs) are metabolites of arachidonic acid (AA) via the cyclooxygenase pathway in mammals. They play versatile biological roles in maintaining the homeostasis of cell, tissue, and body. Among PGs, PGE₂ is known to be a pyrogenic substance (7) as well as an immunosuppressor (8), and PGD₂ is a somnogenic substance (9). They mediate inflammation and initiate physiological responses that are similar to the symptoms observed during malaria. For many years, it has been thought that malaria symptoms are mostly, if not solely, mediated by PGs produced by host cells. We addressed the question whether or not P. falciparum produces PGs. Here we show the evidence that P. falciparum produces PGs in a way distinguishable from the mammalian system.
Materials and Methods

Culture and Preparation of Parasite Cells. P. falciparum FCR 3 and Honduras-1 strains were cultured as previously described (10) and modified by Sugiyama et al. (11) in a 5% O2 and 5% CO2 atmosphere with 3% (vol/vol) of type O RBCs in complete medium that consists of incomplete medium and 10% (vol/vol) heat inactivated type O human serum; incomplete medium is RPMI 1640 supplemented with 25 mM Heps, 25 mM NaHCO3, 0.36 mM hypoxanthine, 3.4 mM glutamine, 10 μg/ml gentamycin, 100 U/ml penicillin, and 100 μg/ml streptomycin. Type O blood was freshy withdrawn into a tube containing citrate phosphate dextrose as anticoagulant. RBCs were washed three times with incomplete medium and cells were collected by centrifugation at 3,000 rpm for 15 min. White blood cells were not detected (<0.01%) in the prepared RBCs by visual microscopic inspection. The parasite cell culture was synchronized by sorbitol treatment as previously described (11). Infected RBCs were isolated by Percoll (Amersham Pharmacia Biotech Inc., Uppsala, Sweden) density centrifugation as described by Tosta et al. (12) with minor modifications. The cell suspension was overlaid on 3 vol of 63% (vol/vol) Percoll in PBS. After centrifugation at 1,800 rpm for 10 min at 25°C, the interphase that contained mainly schizonts and residual trophozoites was collected. The resulting cell fraction was diluted with 2 vol of PBS and centrifuged. The parasite cells were prepared by lysing the infected RBC membrane with 0.075% saponin (11). The resulting parasite cell preparation contained a small number of RBCs (0.5–0.9% of parasite cells), but white blood cells were not detectable (<0.01%). Infected RBCs and isolated parasite cells were suspended in 1.5 vol of PBS and frozen before homogeneate preparation.

Preparation and Incubation of Parasite Cell Homogenates. A suspension of 106 parasite cells in 0.3 ml of PBS was mixed with 5.7 ml of 100 mM sodium phosphate, pH 7.0, containing a protease inhibitor cocktail (Complete; Boehringer Mannheim, Mannheim, Germany) and 600 mg of acid-washed glass beads (425–600 μm; Sigma Chemical Co., St. Louis, Mo.) in a glass tube. The content of the tube was vigorously vortexed for 2 min and kept in an ice bath for 2 min. The cycle was repeated five times to disrupt the cells. The resulting suspension was used as the cell homogenates. The reaction mixture (500 μl) contained 100 mM sodium phosphate (pH 7.0), 2 mM hematin, 5 mM tryptophan, 1 mM AA, and 300 μl of cell homogenates. The mixture was incubated at 37°C for 30 min and the reaction was stopped by addition of 100 μl of 1 M HCl and 6 vol of cold ethyl acetate.

Extraction and Quantification of PGs. [3H]-PGD2, [3H]-PGE2, and [3H]-PGF2a (60 Bq for each per assay; DuPont-NEN, Boston, MA) were added to each sample as tracer to determine the recovery of the following purification procedures. PGs produced in the homogenates (300 μl) and those in the culture medium (8 ml) were extracted three times with 6 vol of cold ethyl acetate and separated by TLC as previously described (13). PGD2, PGE2, and PGF2a were quantified by enzyme immunoassay (EIA) by use of the respective EIA kits (Cayman Chemical Co., Ann Arbor, MI).

Gas chromatography–mass spectrometry analysis. Gas chromatography–selected ion monitoring (GC-SIM) analyses were run on a double focusing mass spectrometer (M-808; Hitachi, Tokyo, Japan) equipped with a Van den Berg’s solventless injector and a fused silica capillary column (Uitra1; 25 m length, 0.32 mm internal diameter 280°C column temperature; Hewlett Packard, Palo Alto, CA). The parasite cells and the culture medium were prepared from the FCR 3 strain grown in the medium containing 33 μM AA. The synchronized 150-ml culture at trophozoite and schizont stages was centrifuged at 1,800 g for 5 min at room temperature and washed with incomplete medium. Fresh complete medium containing 33 μM AA was added to the culture. After another 2-h incubation, the culture medium was taken and PGs were extracted. Then PGs in culture medium and PGs produced in the reaction by the parasite cell homogenates were fractionated by HPLC. The PGs obtained in the eluate were converted into their corresponding methyl ester–dimethylisopropylsilyl (M-E-DMiPS) ether or M-E-methoxime (M-O)-DMiPS ether derivatives according to the method described previously, before applying on GC-SIM (14).

Results

Cell homogenates of RBCs infected with the FCR 3 (Gambia) strain of P. falciparum and of the isolated parasite cells by saponin treatment produced significant amounts of PGD2, PGE2, and PGF2a after incubation with 1 mM AA at 37°C for 30 min (Fig. 1). PGD2 production in both homogenates is highest among three PGs. The amount of PGF2a produced by homogenate of isolated parasite cells was significantly more than that of infected RBCs. PG production in the parasite homogenates required exogenous addition of AA, i.e., no significant amounts of PGs were detected after incubation without AA or without incubation (Fig. 1). PG production was not observed in the homogenates of uninfected RBCs or with complete medium (Fig. 1).

To rule out the possibility that PG production is strain specific, PG production was examined in two parasite strains, FCR 3 and Honduras-1. Fig. 2 shows PG production was observed in both strains. In these parasite strains, the effect of exogenous addition of AA into culture medium was investigated. When parasite cells were grown in the medium supplemented with 33 μM AA for 48 h before harvest, PG production by the homogenates of FCR 3 increased from 200 to 700 pg/10^6 cells for PGD2, and from 100 to 550 pg/10^6 cells for PGF2a, whereas PGE2 production remained almost at the same level (120 pg/10^6 cells) (Fig. 2a). The Honduras-1 cell homogenates also produced PGD2 and PGF2a (120 and 80 pg/10^6 cells, respectively), and the PG production increased to 630 and 330 pg/10^6 cells, respectively, by adding 33 μM AA to the culture.

Accumulation of PGs in the culture medium of P. falciparum was observed in both strains and this accumulation was also increased by the addition of AA to culture medium.
Culture supernatant of FCR3 and Honduras-1 strains were grown in medium with or without the addition of 33 μM AA for 48 h before harvesting. After incubation of the parasite homogenates with 1 mM AA at 37°C for 30 min, PGs produced were measured by EIA. The values shown are from three independent experiments with SE. In a typical experiment, FCR3 culture contained 0.2, 2.5, and 2.5% of RBCs infected with rings, trophozoites, and schizonts, respectively, and that of Honduras-1 contained 0.8, 3.3, and 1.4% of RBCs with rings, trophozoites, and schizonts, respectively. Cells were collected by centrifugation at 1,800 g for 5 min and washed twice with incomplete medium. Fresh medium containing 33 μM AA was added to the culture, and the cultivation was continued for another 2 h. The culture supernatant (8 ml) was then collected by centrifugation at 1,800 g for 5 min at room temperature and stored at −80°C until PG quantification by EIA was performed. The Giemsa-stained parasite cells were counted microscopically. Open and filled circles and open squares represent ring, trophozoite, and schizont, respectively. White, light gray, and dark gray bars represent PGD₂, PGE₂, and PGF₂α, respectively.

Figure 3. Time course of PG release from parasite cells. A ring-rich parasite cell culture (120 ml) of P. falciparum FCR3 strain was grown in complete medium containing 33 μM AA for 48 h. This culture was then treated with sorbitol and incubated for 30 h followed by a second sorbitol treatment to obtain synchronized early rings with a life span of 4–6 h after invasion of fresh RBCs. The synchronized 120-ml culture, supplemented with additional AA, was divided into 10 plastic plates. The time course was started after the last sorbitol treatment. The culture in each plate was centrifuged at 1,800 g for 5 min at room temperature at the indicated times and washed twice with incomplete medium. Fresh complete medium containing 33 μM AA was added to the culture, and the cultivation was continued for another 2 h. The culture supernatant (8 ml) was then collected by centrifugation at 5,000 g and stored at −80°C until PG quantification by EIA was performed. The Giemsa-stained parasite cells were counted microscopically. Open and filled circles and open squares represent ring, trophozoite, and schizont, respectively. White, light gray, and dark gray bars represent PGD₂, PGE₂, and PGF₂α, respectively.
and by these chemicals (Table 1). These results indicate that the catalytic substance(s) for PG production in P. falciparum is clearly different from the well-known enzymes involved in the mammalian arachidonate cascade system.

When homogenates of P. falciparum cultured with 33 μM AA were incubated with various concentrations of AA, PG production increased in a concentration-dependent manner (Fig. 5). However, the concentration-dependent curves are kinetically distinctive among PGs. PGD₂ production almost linearly increased to 700 pg/10⁸ cells with 1 mM AA, giving an apparent Kₘ value for AA >1 mM, whereas PGE₂ and PGF₂α formation reached almost the maximum at ~0.25 mM AA (180 and 550 pg/10⁸ cells, respectively) and increased slightly with the higher concentrations, giving an apparent Kₘ value for AA of ~50 μM. Since the Kₘ values for AA of the human cyclooxygenase-1 and -2 are reportedly 5–6 μM (15, 16), the catalytic substance(s) for PG production in P. falciparum is clearly different from the mammalian enzyme.

Table 1. Effects of Nonsteroidal Antiinflammatory Drugs and Heat on PG Production by P. falciparum FCR3

<table>
<thead>
<tr>
<th>Enzyme source</th>
<th>T Treatment</th>
<th>PGD₂ (pg/assay)</th>
<th>PGE₂ (pg/assay)</th>
<th>PGF₂α (pg/assay)</th>
</tr>
</thead>
<tbody>
<tr>
<td>P. f.</td>
<td>None</td>
<td>454</td>
<td>105</td>
<td>328</td>
</tr>
<tr>
<td>P. f.</td>
<td>Aspirin</td>
<td>420</td>
<td>92</td>
<td>292</td>
</tr>
<tr>
<td>P. f.</td>
<td>Indomethacin</td>
<td>408</td>
<td>100</td>
<td>279</td>
</tr>
<tr>
<td>P. f.</td>
<td>100°C, 10 min</td>
<td>272</td>
<td>79</td>
<td>213</td>
</tr>
<tr>
<td>P. f.</td>
<td>121°C, 90 min, 1.1 atm</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Sheep cox</td>
<td>None</td>
<td>930</td>
<td>1150</td>
<td>180</td>
</tr>
<tr>
<td>Sheep cox</td>
<td>Aspirin</td>
<td>48</td>
<td>53</td>
<td>12</td>
</tr>
<tr>
<td>Sheep cox</td>
<td>Indomethacin</td>
<td>30</td>
<td>46</td>
<td>15</td>
</tr>
<tr>
<td>Sheep cox</td>
<td>100°C, 10 min</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
</tbody>
</table>

The parasite cells (P. f.) were prepared from P. falciparum FCR3 strain grown in a medium containing 33 μM AA, as described in Materials and Methods. The cyclooxygenase preparation of sheep seminal vesicle (sheep cox) was obtained from Eldan-Tech Ltd. (Israel). The parasite homogenates (5 × 10⁸ cells) and sheep cyclooxygenase (300 ng protein) were incubated at 37°C for 30 min with 1 mM AA in the presence or absence of 3 mM aspirin or 42 μM indomethacin. Heat treatment was carried out at 100°C for 10 min and 121°C for 90 min before incubation with 1 mM AA. The amounts of PGs produced were measured by EIA. Detection limits were 25, 11, and 10 pg in each assay for PGD₂, PGE₂, and PGF₂α, respectively. ND, not detected.
The parasite homogenates were incubated at 37°C for 30 min with various concentrations of AA added exogenously. The amounts of PGs produced were quantified as described under Materials and Methods. Triangles, circles, and squares represent PGD2, PGE2, and PGF2α, respectively.

The malaria fever is coordinated with the schizont rupture associated with increased TNF-α level in the blood stream. It was shown that a glycosylphosphatidylinositol-like substance from malaria parasites stimulates the host TNF-α production (19). The malaria fever was, therefore, supposed to be caused by TNF-α after the induction of host PGE2 production. Our observation that parasite-produced PGs were accumulated in the culture medium at trophozoite and schizont stages raises a possibility that parasite-produced PGE2 and PGD2 directly contribute to fever and sleepiness as well.

Why does P. falciparum produce PGs? The most intriguing possibility is that the parasite-produced PGs modulates the host defense mechanism against malaria infection. For instance, an increased level of TNF-α was reported to suppress the parasite growth in vivo (20, 21). Lowering the host TNF-α production by parasite produced PGE2 would be beneficial to the parasite, since PGE2 regulates the level of TNF-α (22, 23). In the other parasitic organism, it was reported that microfiliae of Brugia malayi, intravascular nematode, generates and releases prostacyclin and PGE2 from arachidonic acid (24, 25). The reported PG synthesis is sensitive to indomethacin, unlike P. falciparum. These prostanooids may influence host immune and other cellular responses to ease the parasitism of microfiliae.

Further investigation into the contribution of parasite-produced PGs and catalysts in the clinical manifestation of this disease would provide a new means for controlling malaria.

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