Brief Definitive Report

P-Selectin Glycoprotein Ligand-1 (PSGL-1) on T Helper 1 but Not on T Helper 2 Cells Binds to P-Selectin and Supports Migration into Inflamed Skin

By Eric Borges,* Wolfgang Tietz,† Martin Steegmaier,* Thomas Moll,* Rupert Hallmann,§ Alf Hamann,‡ and Dietmar Vestweber*

From the *Institut für Zellbiologie ZMBE, Universität Münster, Germany; †Abteilung für Immunologie, Medizinische Klinik, Universität Eppendorf, Hamburg, Germany;‡Abteilung für Immunologie, Abteilung für Zellbiologie, ZMBF, Universität Münster, Germany;§Institut für Exp. Medizin u. Bindegewebsforschung, Universität Erlangen-Nürnberg, Germany

Summary
We have shown recently that mouse Th1 cells but not Th2 cells are selectively recruited into inflamed sites of a delayed-type hypersensitivity (DTH) reaction of the skin. This migration was blocked by monoclonal antibodies (mAb) against P- and E-selectin. Here we show that Th1 cells bind to P-selectin via the P-selectin glycoprotein ligand-1 (PSGL-1). This is the only glycoprotein ligand that was detectable by affinity isolation with a P-selectin-Ig fusion protein. Blocking of Th1 cells to P-selectin, as analyzed by flow cytometry and in cell adhesion assays, was completely blocked by antibodies against PSGL-1. The same antibodies blocked partially the migration of Th1 cells into cutaneous DTH reactions. This blocking activity, in combination with that of a mAb against E-selectin, was additive. PSGL-1 on Th2 cells, although expressed at similar levels as on Th1 cells, did not support binding to P-selectin. Thus, the P-selectin–binding form of PSGL-1 distinguishes Th1 cells from Th2 cells. Furthermore, PSGL-1 is relevant for the entry of Th1 cells into inflamed areas of the skin. This is the first demonstration for the importance of PSGL-1 for mouse leukocyte recruitment in vivo.

The binding of circulating leukocytes to activated endothelium is initiated by transient interactions that are mediated by the selectins (1). It is well documented in various inflammation models that blocking of the two endothelial selectins, P- and E-selectin, inhibits the entry of neutrophils into inflamed tissue (2). Less is known about the role of these adhesion molecules for T lymphocyte recruitment in inflammation. Binding to E-selectin was shown for certain subsets of human CD4+ memory T lymphocytes (3, 4) and for a large percentage of bovine γδ T cells (5). A small percentage of CD4+ T cells from peripheral blood (6) and also chronically activated CD4+ T cells (7) was found to bind to P-selectin. Human CD4+ T cell clones were described to bind to E- and P-selectin in static (8) as well as flow adhesion assays (9), and T cell recruitment into inflamed skin was blocked with polyclonal antibodies against P-selectin in vivo in the rat (10).

Binding of activated T cell lines to P-selectin under static conditions was partially blocked in vitro by high concentrations of an anti-human P-selectin glycoprotein ligand-1 (PSGL-1) antiserum (9, 11). PSGL-1 was originally identified on human neutrophils by affinity isolation with P-selectin (12, 13) and cloned by expression cloning (14). It was found to be the major binding site for P-selectin on human leukocytes (11, 15). Rolling of human leukocytes perfused into rat postcapillary venules was demonstrated to be blocked by a mAb against human PSGL-1 (16).

Upon activation, T helper lymphocytes polarize into Th1 and Th2 subsets, which are characterized by distinct profiles of secreted cytokines (17, 18). Th1 cells are involved in cell-mediated inflammatory reactions. Their cytokines activate cytotoxic and inflammatory functions and Th1 cells induce delayed-type hypersensitivity (DTH) reactions. Th2 cytokines support antibody production, particularly IgE responses, and in combination with their stimulatory effects on eosinophil proliferation and function, Th2 cytokines are commonly found in association with strong antibody and allergic responses. Although it is well established that Th1 cells predominate in DTH reactions, it was always unclear whether their presence is mainly due to polarized differentiation at these sites or could also be based on preferential immigration of Th1 versus Th2 cells. We have shown recently that mouse Th1 cells indeed migrate into cutaneous DTH reactions much better than Th2 cells do, and we could demonstrate that this migration is blocked by mAb against P- and E-selectin (19).

In this study, we have examined which molecules on the surface of Th1 cells would function as ligands for P-selectin during migration into cutaneous DTH reactions in the mouse. We could define the PSGL-1 as the exclusive P-selectin ligand on Th1 cells by affinity isolation experiments, FACS® analysis, and cell adhesion assays. Th2 cells carried similar amounts of PSGL-1; however, this form of PSGL-1 was unable to bind to P-selectin. Antibodies against PSGL-1...
could partially block the migration of T\textsubscript{H}1 cells into cutaneous DTH reactions and showed additive effects with a mAb against E-selectin.

**Materials and Methods**

Cells. The mouse neutrophilic cell line 32Dcl3 was cultured as described (20). T\textsubscript{H}1 and T\textsubscript{H}2 cells were generated from lymph node lymphocytes of SPF-reared BALB/c mice. CD4\textsuperscript{+} T cells were derived by panning of isolated lymphocytes with mAb against CD8 (53-672), CD25 (PC/6), FcR receptor II/III (2.4G2), M ac-1 (M 1/70), and I-A\textsuperscript{d} (17/227). Of the resulting cells, 98–99% were positive for CD4 staining. These cells (10\textsuperscript{6}/well) were incubated either in the presence of IL-12 (1,000 U/ml) and IFN-\gamma (200 U/ml) (for generation of T\textsubscript{H}1 cells) or in the presence of IL-2 (50 U/ml) and IL-4 (10 ng/ml) (for the generation of T\textsubscript{H}2 cells) in 24-well plates coated with mAb 145-2C11 against CD3. After 2 d, cells were transferred to noncoated plates without changing medium and cultured for another 3 or 4 d as described (21). Under these conditions, the differentiated effector cells have returned to a resting state (21). Purity of T\textsubscript{H}1/T\textsubscript{H}2 subsets were verified for cytokine production by intracellular FACS\textsuperscript{©} analysis (22). The two generated T cell populations contained routinely >50% cells of the desired phenotype and were contaminated by <1% cells of the opposite phenotype.

In Vivo Studies. T\textsubscript{H}1 cells harvested 6 d after initial activation were subjected to density gradient centrifugation (17.1% isotonic Nycodenz; Nycodan, Oslo, Norway) to remove dead cells. 10\textsuperscript{6} cells were subjected to density gradient centrifugation (17.1% isotonic Nycodenz; Nycodan, Oslo, Norway) to remove dead cells. 10\textsuperscript{6} were incubated with 25 \textmu g/ml selectin-IgG chimera or 10 \mu g/ml affinity-purified rabbit antibodies in HBSS (Biochrom, Berlin, Germany) with 3% FCS, and stained with PE-labeled F(ab'), donkey anti-human IgG (Dianova, Hamburg, Germany) or FITC labeled goat anti-rabbit IgG (Sigma). Analysis was performed on a Becton Dickinson FACSCalibur\textsuperscript{®} and CellQuest\textsuperscript{®} analysis software.

Antibodies and Selectin-IgG Chimera. The mAb UZ4 (rat IgM) against mouse E-selectin (23) and the mAb R B40.34 (rat IgG 1) against mouse P-selectin (24) were produced in and purified from protein-free hybridoma medium (Gibco/BRL, Karlsruhe, Germany). In analogy to Y ang el al. (25), we raised the rabbit anti-mouse PSGL-1 antiserum 124 against a peptide covering amino acids 42–60 of the sequence of murine PSGL-1. This sequence starts after the cleavage site of the propeptide, and the analogous region of human PSGL-1 is known to be necessary for P-selectin binding (26) and to harbor the binding epitope of the adhesion blocking mAb PL-1 (27). Specific antibodies were purified by affinity isolation on the BSA-conjugated PSGL-1 peptide linked to CN Br-Sepharose as described (28). The FACS\textsuperscript{©} signal which was obtained with these antibodies on 32D cl3 cells was abolished after pretreatment of the cells with O-sialoglycoprotease (treatment done as described in 29).

For the preparation of Fab-fragments, 2.5 mg of affinity-purified anti-PSGL-1 antibodies were incubated in 500 \mu l of 20 mM sodium phosphate, pH 7.0, 10 mM EDTA, 10 mM cysteine with 100 \mu l of washed papain Sepharose beads (Pierce Chemical Co., Rockford, IL) for 4 h at 37\textdegree C. The supernatant was dialyzed against PBS, and Fc fragments and undigested antibodies were removed with protein A-Sepharose (Pharmacia LKB, Piscataway, NJ). The P-selectin-IgG fusion protein was produced as described (28).

**Immunoprecipitation.** T\textsubscript{H}1 and T\textsubscript{H}2 cells harvested 5 d after initial activation were subjected to density gradient centrifugation, washed three times in PBS, and surface biotinylated in 1 ml of PBS containing 0.5 mg/ml Sulfo-NHS-biotin (Pierce) for 30 min on ice. The reaction was blocked by incubating the cells for 10 min in 1 ml of DMEM (Gibco/BR L) without FCS on ice. Cells were washed with PBS twice and subjected to immunoprecipitations performed as described (29). Immunoprecipitated proteins were separated by electrophoreses on 6% SD-PAGE and transferred to nitrocellulose (Schleicher & Schuell, Keene, N.H.). Filters were analyzed for biotinylated proteins with peroxidase-conjugated streptavidin (Dianova, Hamburg, Germany) and the ECL-system (Amersham Corp., Arlington Heights, IL).

Cell Adhesion Assay. Adhesion assays in rotating 96-well microtiter plates coated with selectin-Ig fusion proteins were performed as described (29), except for evaluating the number of bound cells, which was done by computer-aided image analysis using the NIH Image 1.55 software. For each value, three wells were analyzed, counting three randomly chosen areas per well.

**Results and Discussion**

We have recently shown that T\textsubscript{H}1 cells but not T\textsubscript{H}2 cells efficiently enter sites of acute inflammation in the skin (19). This migration was blocked by a combination of antibodies...
against P- and E-selectin (19). Based on these findings, we have now examined which molecules on Th1 cells would function as ligands for P-selectin in this process. We first analyzed which glycoproteins on the surface of Th1 cells bind to P-selectin. To this end, equal numbers of Th1 and Th2 cells were surface biotinylated and subjected to affinity isolation experiments with a P-selectin–Ig fusion protein. As shown in Fig. 1 (lane 5), two proteins of 230 and 130 kD were isolated with P-selectin–Ig that could be reprecipitated with affinity-purified antibodies of the rabbit antiserum 124 against an amino-terminal peptide of mouse PSGL-1 (lane 6). The 230-kD form is the not completely reduced dimeric form of the 130-kD PSGL-1. No selectin ligands could be isolated with the P-selectin probe from Th2 cells (lane 2), although the PSGL-1 protein was detectable with antibodies (lane 3) at similar levels as on Th1 cells (lanes 3 and 4). Interestingly, the 130-kD monomeric form of PSGL-1 on Th2 cells was slightly smaller than the one on Th1 cells (compare lane 3 with 5). Whereas antibodies against PSGL-1 precipitated a broader band from Th1 cells (lane 4), including a slightly smaller molecular species of the size as in Th2 cells, P-selectin–Ig only precipitated the higher molecular weight species. We conclude that only Th1 cells express a modified form of PSGL-1 that is capable of binding to P-selectin. Furthermore, PSGL-1 was the only glycoprotein ligand on Th1 cells that could be detected by this technique.

In line with the immunoprecipitation results, a comparison of Th1 and Th2 cells by FACS analysis revealed that both cell types expressed similar levels of PSGL-1 (Fig. 2, A and B). However, 45% of the Th1 cells, but none of the Th2 cells, could be stained with P-selectin–Ig (Fig. 2, C and D). Staining with P-selectin–Ig was completely blocked by preincubating the cells with nonimmune rabbit IgG (A and B) or with human IgG (C and D). E shows the staining of Th1 cells with P-selectin–IgG after preincubation of the cells either with nonimmune rabbit IgG (A and B) or with affinity-purified rabbit anti-PSGL-1 antibodies (bold line). P-selectin–Ig was detected with PE-conjugated F(ab’)2, donkey anti-human IgG, and rabbit antibodies were detected with FITC-conjugated goat anti-rabbit IgG.
PSGL-1 Requires Certain Posttranslational Modifications for P-Selectin Binding

PSGL-1 is the major receptor of P-selectin on Th1 cells and is involved in the migration of Th1 cells into inflamed skin. Our data demonstrate that PSGL-1 is the major P-selectin ligand on Th1 cells and that it is relevant for the entry of these cells into inflamed skin. The fact that anti-PSGL-1 Fab fragments inhibited Th1 cell migration less efficiently than the anti-P-selectin mAb is probably due to the lower blocking efficiency of the Fab fragments as compared with complete antibodies. In vitro adhesion assays demonstrated that 50 μg/ml anti-PSGL-1 Fab inhibited adhesion of mouse 32Dcl3 neutrophilic cells to P-selectin-Ig by only 35%, while 20 μg/ml complete anti-PSGL-1 antibodies blocked adhesion by 70% (not shown).

Although the cell surface expression level of PSGL-1 on Th1 and Th2 cells is similar, only PSGL-1 on Th1 cells is able to mediate binding to P-selectin. It is well known that PSGL-1 requires certain posttranslational modifications for binding to P-selectin, such as fucosylation (14), tyrosine-sulfation (30–32), and branched carbohydrate side chains generated by the core-2 enzyme (33–35). It is likely that one or several of these modifications are upregulated in the course of the differentiation process that leads to the acquisition of the Th1 cell phenotype. Indeed, careful analysis of the apparent molecular weight of PSGL-1 on Th1 and Th2 cells revealed that the P-selectin-binding form of PSGL-1 on Th1 cells has a slightly larger apparent molecular weight.
than the nonfunctional form on Th2 cells (Fig. 1, lanes 3 and 5). Besides the different repertoire of cytokines, the functional form of PSGL-1 is a cell surface molecule that distinguishes Th1 cells from Th2 cells. The expression of active PSGL-1 is relevant for the migration of Th1 cells into inflamed skin.

This work was supported by grants from the Deutsche Forschungsgemeinschaft to D. Vestweber, R. Hallmann, and A. Hamann and from the Meldred Scheel Stiftung to A. Hamann.

Address correspondence to Dietmar Vestweber, Institute of Cell Biology, ZMBE, Technologiehief, Mendelstr. 11, D-48149 Munster, Germany.

Received for publication 18 October 1996 and in revised form 24 November 1996.

References


