Essential Role of Macrophage Colony-Stimulating Factor in the Osteoclast Differentiation Supported by Stromal Cells

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Summary

Severe deficiency of osteoclasts, monocytes, and peritoneal macrophages in osteopetrotic (op/op) mutant mice is caused by the absence of functional macrophage colony-stimulating factor (M-CSF). To clarify the role of M-CSF in the osteoclast differentiation, we established a clonal stromal cell line OP6L7 capable of supporting hemopoiesis from newborn op/op mouse calvaria. Although very few macrophages appeared in the cocultures of bone marrow cells and OP6L7 cells, a 50-fold larger number of macrophages was detected in the day 7 cocultures when purified recombinant human M-CSF (rhM-CSF) was exogenously supplied. Tartrate-resistant acid phosphatase (TRACP; a marker enzyme of osteoclasts)-positive cells appeared only when bone marrow cells were cultured in contact with OP6L7 cells and both rhM-CSF and 1α, 25 (OH)2D3 were added. The TRACP-positive cells became multinucleated with increasing time in culture and expressed the c-fms/M-CSF receptor. These results indicate that both contact with stromal cells and M-CSF are requisite for osteoclast differentiation under physiological conditions.

Macrophage colony-stimulating factor (M-CSF) has been identified in vitro by its ability to support the clonal growth and differentiation of precursor cells of monocytic-macrophage lineage. Mice homozygous for the recessive osteopetrosis mutation (op/op), having a severe deficiency of osteoclasts, monocytes, and peritoneal macrophages (1-3), have recently been found to be defective in the production of functional M-CSF (4-6) as a result of a one base-pair insertion within the coding region of the M-CSF gene on chromosome 3 (4). Wiktor-Jedrzejczak et al. (5) reported that the macrophage deficiency in the peritoneal cavity of the mutants is partially corrected by implantation of diffusion chambers containing M-CSF-secreting Ir929 cells. Felix et al. (7) and Kodama et al. (8) have demonstrated that the deficiencies of osteoclasts and monocytes can be completely cured by daily injections of purified recombinant human M-CSF (rhM-CSF). These findings indicate that M-CSF is essential for the differentiation not only of monocyte/macrophages but also of osteoclasts under physiological conditions.

However, the precise role of M-CSF in osteoclast differentiation has not yet been made clear. Kurihara et al. (9) reported that M-CSF, unlike interleukin 3 and granulocyte/macrophage colony-stimulating factor, cannot support in vitro osteoclast formation. Udagawa et al. (10) developed another in vitro culture system for osteoclast differentiation, one in which spleen cells are cocultured with stromal cell line MC3T3-G2/PA6 or ST2. MC3T3-G2/PA6 cells support differentiation of osteoclasts (10), as well as proliferation of hemopoietic stem cells defined as colony-forming units in spleen (CFU-S) (11), only through short range cell-to-cell interaction; whereas the stromal cells per se produce M-CSF (Sudo, T., personal communication).

In this study, we established a clonal stromal cell line OP6L7 from newborn op/op mouse calvaria capable of supporting hemopoiesis but not differentiation of macrophages and osteoclasts, and demonstrated that both contact with stromal cells and M-CSF are requisite for the osteoclast differentiation.

Materials and Methods

Mice. F2 hybrid mice of +/+ , op/+ , and op/op genotypes were raised in our laboratory from breeding pairs of B6C3F1-α/α, op/+ mice obtained from The Jackson Laboratory (Bar Harbor, ME). Female C57BL/6CrSlc mice were purchased from Shizuoka Laboratory Animal Center (Hamamatsu, Japan).

Establishment of Cell Lines. Calvariae were aseptically removed from a litter of nine newborn F2 mice and minced in 35-mm plastic dishes (Sumitomo Bakelite, Tokyo, Japan). The bone fragments were suspended in medium consisting of α-MEM (Irvine Scientific, Santa Ana, CA), 20% fetal bovine serum (FBS; Irvine Scientific), and 0.08% type I collagen (Nitta Gelatin, Osaka, Japan). After the collagen had gelled, the cultures were incubated at 37°C in 5% CO2 for 4 d. The cells from two mice (3 and 6) showing...
an osteopetrotic phenotype, i.e., an excessive amount of bone trabeculae and very few osteoclasts in their femurs, were subcultured on 3T3 schedule and supplemented with 10% FBS (12, 13). Three of 15 cell lines established could support hemopoiesis for more than 2 wk. One clone, designated OP6L7, was used for the following experiments.

Coculture of Bone Marrow Cells with OP6L7 Cells. Bone marrow cells from 6–10-wk-old C57BL/6CrSlc mice were inoculated at 10^5 cells/dish onto confluent OP6L7 cell layers established in collagen-coated 35-mm dishes and cultured with 1.5 ml of α-MEM supplemented with 20% horse serum (HyClone Laboratories, Logan, UT) and 10^-8 M dexamethasone (DEX) in the absence or presence of 0.5 μg/ml of purified rhM-CSF (gift of Morinaga Milk Industry, Kanagawa, Japan) for 7 d. Cells were harvested with treatment with 0.1% collagenase (Nitta Gelatin). Total number of the cells was determined using a hemocytometer. For differential cell counts, cytospun preparations were stained with May-Grünwald Giemsa solution or treated with F4/80 mAbs specific for macrophages (14) (American Type Culture Collection, Rockville, MD) and subsequently with the reagents of a Vectastain ABC-Go kit (Vector Laboratories, Inc., Burlingame, CA). CFU-S assay was performed as described previously (15).

Osteoclast Formation. Bone marrow cells and OP6L7 cells were cocultured as described above in 1.5 ml of α-MEM supplemented with 10% FBS, 10^-6 M 1α, 25 (OH)2D3 (gift of Tokyo Institute for Bio-Medical Research, Tokyo, Japan), and 10^-8 M DEX in the absence or presence of 0.5 μg/ml rhM-CSF, and the medium was changed twice a week. To examine the necessity of cell-to-cell contact with stromal cells for osteoclast differentiation, we inoculated bone marrow cells onto OP6L7 cell layers covered with 1 ml of collagen gel and cultured them as described above. At indicated times, the cultures were fixed and stained for tartrate-resistant acid phosphatase (TRACP) activity as described previously (10) or immunocytochemically stained with monoclonal anti-cfms/M-CSF receptor (Ab-1) antibody (Oncogene Science, Manhasset, NY). To confirm the osteoclastic activity of the multinucleated cells formed in the cocultures, we seeded bone marrow cells onto OP6L7 cell layers established on ivory slices (provided by Dr. M. Kumegawa, Meikai University School of Dentistry, Sakado, Japan), which were placed in 35-mm dishes and cultured as described above for 21 d. The slices were fixed and trypsinized to remove attached cells. After conductive staining of the slices, their secondary electron images were examined with a Hitachi S-450 scanning electron microscope (Hitachi, Tokyo, Japan).

Results and Discussion

By coculturing bone marrow cells with the stromal cell lines established from newborn op/op mouse calvariae, we found that very few macrophages appeared within the cocultures and that three of these cell lines could support hemopoiesis for more than 4 wk and differentiate into adipocytes in response to glucocorticoid, similarly to MC3T3-G2/PA6 cells (11, 13). Then we isolated a clone, designated OP6L7, from one of these three lines, the OP6L line.

When bone marrow cells were cocultured with OP6L7 cells for 7 d, the number of CFU-S increased about three-fold, since 10^5 normal bone marrow cells contained 23 ± 2.0 CFU-S (Table 1). Very few F4/80-positive macrophages were detected in the cocultures. When rhM-CSF (0.5 μg/ml) was added to the cocultures, a 50-fold larger number of macrophages appeared, while the number of neutrophils and blasts increased only 2.4- and 1.4-fold, respectively. The CFU-S number did not change significantly, showing that rhM-CSF does not affect CFU-S growth in our coculture system. These results demonstrate that OP6L7 cells can support the proliferation of hemopoietic stem cells and their differentiation into neutrophils, but not into macrophages, because of functional M-CSF.

To explore the conditions required for the osteoclast differenti-

### Table 1. Effect of rhM-CSF on the Hemopoiesis Supported by OP6L7 Cells

<table>
<thead>
<tr>
<th>Addition</th>
<th>Macrophages</th>
<th>Neutrophils</th>
<th>Blasts</th>
</tr>
</thead>
<tbody>
<tr>
<td>None</td>
<td>2.0 × 10^9</td>
<td>2.0 × 10^9</td>
<td>5.2 × 10^9</td>
</tr>
<tr>
<td>M-CSF</td>
<td>3.5 × 10^9</td>
<td>4.8 × 10^9</td>
<td>5.2 × 10^9</td>
</tr>
</tbody>
</table>

* Bone marrow cells were inoculated onto confluent OP6L7 cell layers at 10^5 cells/dish and cocultured in the absence or presence of 0.5 μg/ml rhM-CSF for 7 d.
We wish to thank Dr. M. Kumegawa for the generous gift of ivory slices, Morinaga Milk Industry for purified rhM-CSF, and Teijin Institute for Bio-Medical Research for 1α, 25(OH)₂D₃. We also thank Ms. M. Sato for technical assistance.

This work was supported in part by grants from the Ministry of Education, Science, and Culture of Japan.

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Received for publication 1 February 1991.

References


2. Marks, S.C. Jr. 1982. Morphological evidence of reduced bone...


