HETEROGENEOUS LOBULAR DISTRIBUTION OF
HEPATOCYTES EXPRESSING ACUTE-PHASE GENES
DURING THE ACUTE INFLAMMATORY REACTION

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Although a zonal distribution of hepatocytes expressing various metabolic functions in liver lobule has been described (1), it is still debated whether functional heterogeneity in the lobule also occurs with respect to plasma protein synthesis, an important function of hepatocytes. Acute-phase reactants are a group of secretory proteins with dramatically altered rates of synthesis during the acute-phase response of the liver (2). In this article we analyze the hepatic cellular distribution of the mRNA and protein products of three acute-phase genes, α2-macroglobulin (α2M), α1-inhibitor 3 (α1I3), and α1-antitrypsin (α1PI). α2M is a major acute-phase reactant in rats with drastically increased hepatic mRNA concentrations after turpentine injection (3). Concomitantly, the steady-state levels of α1I3 mRNA are decreased (4). α1PI is only a moderately reacting acute-phase protein in the rat (5). Here we investigate specifically whether hepatocytes expressing these three genes show a specialized location in normal rat livers and whether their topographic distribution is altered during the acute inflammatory reaction (AIR).

Materials and Methods

Animals. Adult male Sprague-Dawley rats (180–230 g) (Charles River Breeding Laboratories, St. Aubin-les-Elbeuf, France) were used. An AIR was induced by a subcutaneous injection of turpentine (0.5 ml per 100 g body weight) (Prolabo, Paris, France). The animals were killed at 0, 4, 8, 12, 18, 24, 48, and 72 h after induction of the AIR. Three rats per time point were used for in situ hybridization (ISH) and immunoperoxidase experiments.

Molecular Probes. The α2M (pRLA2M/29J) cDNA probe and the α1I3 (pRLA12/23J) cDNA probe were prepared as described (3, 4). The α1PI (pliv 3) cDNA probe (6) was provided by Dr. M. Weiss (Institut Pasteur, Paris, France).

In Situ Hybridization. Liver fixation and preparation of paraffin sections were made as previously detailed (7). The sections were fixed in 4% paraformaldehyde and hybridized as previously reported (7). The probes were labeled by nick translation to a specific activity of 1–2 × 10⁸ cpm/μg DNA. Each section was hybridized with 10³–10⁶ cpm of the radioactive...
probe at 42°C for 24 h. A quantitative analysis was made by optical grain counts, according to the morphometric method previously described (7). For each time point, at least two different slides from two different animals were evaluated. RNase controls were performed by incubation with RNase A (Sigma Chemical Co., St. Louis, MO) before hybridization with a specific probe. pBR controls were performed by hybridizing tissue sections with a 35S-labeled pBR 325 fragment.

**Immunoperoxidase Staining.** Liver sections adjacent to those used for ISH were processed for detection of α2M, α1I3, and α1PI with an indirect immunoperoxidase method using specific antisera (5, 8, 9). For control reactions, sections were incubated with peroxidase-labeled anti-rabbit Ig or directly placed in the diamino-benzidine medium.

**Results**

**Analysis by ISH.** In normal liver α2M, α1I3, and α1PI mRNA were detected in all hepatocytes. The signal for α1I3 mRNA was high, while α2M (Fig. 1 a) and α1PI mRNA appeared less abundant. Furthermore, while the signal for α1PI mRNA was homogeneously distributed within the liver lobule, the distribution of the signal for

![Figures 1 and 2.](image)

(Fig. 1, left) Localization of α2M gene products. (a) Localization of mRNA in normal liver. Silver grains are more numerous in PP and ML hepatocytes than in PV hepatocytes. (b) Localization of α2M mRNA 18 h after AIR induction. The hybridization signal is dramatically increased over PP and ML hepatocytes. (c) Immunoperoxidase localization of α2M 18 h after AIR induction. A strong staining is visible in PP and ML hepatocytes, while PV hepatocytes are negative. PT, portal tract; CLV, centrilobular vein. Bars: (a and b) 50 μm, (c) 100 μm. (Fig. 2, right) Localization of α1PI gene products. (a) Localization of mRNA in normal liver. All hepatocytes are labeled with a homogeneous distribution in the lobule. (b) Immunoperoxidase localization of α1PI in normal liver. All hepatocytes in the lobule are reactive, with a more intense staining in PV hepatocytes. (c) Immunoperoxidase localization of α1PI 24 h after AIR induction. The staining intensity is increased, and is now homogeneously distributed across the lobule. PT: portal tract. CLV: centrilobular vein. Bars: (a) 50 μm, (b and c) 100 μm.
\(\alpha_1\beta_3\) and \(\alpha_2\beta_3\) was uneven over the three zones of the lobule. Silver grains corresponding to \(\alpha_2\beta_3\) mRNA (Fig. 1a) and \(\alpha_1\beta_3\) mRNA were more numerous in the periportal (PP) and mediolobular (ML) area than in the perivenous (PV) areas. 4 h after induction of the AIR, several sparsely distributed PP hepatocytes displaying a dramatic increase in the signal level for \(\alpha_2\beta_3\) mRNA were observed. At later times the number of hepatocytes with a high signal level per cell increased progressively. At the maximum of the AIR (18 h) nearly all hepatocytes of the PP and ML areas were intensely labeled, whereas the signal level in the PV areas remained mostly unchanged (Fig. 1b). By contrast, as early as 4 h after induction of the AIR, \(\alpha_1\beta_3\) signal levels were decreased in the PV and ML areas and were still low at 24 and 72 h. Finally, the signal for \(\alpha_1\beta_1\) mRNA increased moderately over all hepatocytes between 4 and 18 h after induction of the AIR, and the distribution of the signal within the lobule remained homogeneous at all times tested (Fig. 2a).

Optical counts revealed an increase in the number of \(\alpha_2\beta_3\) mRNA in PP hepatocytes during the AIR of about threefold and fourfold for the nuclear and cytoplasmic counts, respectively (Fig. 3a). In PV areas the nuclear signal level remained unchanged at all times tested, with moderate fluctuations in the cytoplasmic counts during the AIR. With regard to \(\alpha_1\beta_3\) expression, a decrease of nuclear and cytoplasmic counts was recorded in PP hepatocytes 4 h after induction of the AIR, reaching a 6-fold decrease of the nuclear counts and a 3.4-fold decrease of the cytoplasmic counts.

![Figure 3](image-url)

**Figure 3.** Quantitation of the signal level over PP and PV hepatocytes during an AIR. Liver sections were hybridized with an \(\alpha_2\beta_3\) (a), \(\alpha_1\beta_3\) (b), or \(\alpha_1\beta_1\) (c) probe, and the average number of nuclear and cytoplasmic silver grains per PP (open symbols) or PV (dark symbols) hepatocytes was recorded, as indicated in Materials and Methods. The values are the mean of three separate experiments. (O, □) Nuclear counts. (Δ, ▲) Cytoplasmic counts.
counts by 48 h (Fig. 3 b). Only slight fluctuations of the nuclear and cytoplasmic counts were visible over PV hepatocytes. For α1PI, the increase of the signal was small (~2-fold for the nuclear and cytoplasmic counts) and of the same magnitude in the two lobular zones (Fig. 3 c).

**Immunoperoxidase Staining.** In normal liver a weak α2M immunostaining was detected exclusively in PP and ML hepatocytes. All hepatocytes stained for α1I3 in the normal liver, except for a narrow ring of PV hepatocytes. All hepatocytes stained for α1PI, with a more intense staining in PV hepatocytes (Fig. 2 b). During the AIR, the staining intensity for α2M increased progressively, and by 18–24 h almost all hepatocytes in the PP and ML zones were strongly stained (Fig. 1 c). Concomitantly, the intensity of staining for α1I3 decreased progressively in PP and ML areas. PV hepatocytes remained negative for both proteins during the AIR. The lobular distribution of α1PI+ hepatocytes changed towards a homogeneous pattern between 12 and 48 h after induction of the AIR (Fig. 2 c).

**Discussion**

Our study demonstrate that the location of a hepatocyte within the lobule influences the level at which it expresses the three acute-phase genes studied via pre- and/or posttranslational mechanisms, depending on the gene.

In normal liver, we observed that PP and ML hepatocytes contained much more α1I3 and α2M mRNA and protein than PV hepatocytes, although a low level of expression of the two genes was also detected in these cells by ISH. These observations suggest that as yet unknown factors, linked to the location of hepatocytes within PP and ML microenvironments, influence the basal transcription rates and/or mRNA stabilities of the α2M and α1I3 genes. During the AIR, the α2M gene was stimulated early in PP hepatocytes with a later recruitment of ML hepatocytes, suggesting that the signals for α2M induction spread from the PP to the PV zones. These observations, in agreement with earlier reports on the morphologic kinetics of α2M and C-reactive protein syntheses in the liver (10, 11), are consistent with the interpretation that the AIR mediators spread from the portal vein through the entire lobule. Similarly the decrease in α1I3 mRNA during the AIR was shown to take place in the PP and ML zones. The pattern of lobular expression of the α1PI gene in normal and stimulated livers differed significantly from that of the α2M and α1I3 genes. Staining for the α1PI protein was more intense in PV hepatocytes, as previously reported (5), whereas, the repartition of α1PI mRNA was homogeneous across the lobule. This observation is compatible with the interpretation that some mechanism(s) affecting translational or posttranslational processing, stability, and/or secretion of α1PI specifically operate in PV hepatocytes, leading to a more important accumulation of the α1PI protein within these cells. We found no difference in the extent of α1PI mRNA induction between PP and PV hepatocytes on the one hand, and between nuclear and cytoplasmic α1PI RNA products on the other. The present data, together with our previous observations of lobular heterogeneity in the level of expression of the albumin and α-fetoprotein genes (12), suggest that functional lobular heterogeneity is a concept applicable not only to enzymatic or metabolic functions (1), but to plasma protein synthesis as well.

The molecular mechanisms responsible for differences in the expression of plasma protein genes in different areas of a liver lobule are presently unknown. While it
is clear that hormones released by macrophages participate in the regulation of acute-phase genes (2), it is not yet clear whether hormones are also responsible for the uneven expression of these genes across the liver lobule in unstimulated livers. Hepatocytes of the different lobular areas differ in at least two aspects: (a) They are surrounded by different microenvironments as a result of the unidirectional blood flow, which leads to decreasing concentrations of soluble mediators from the portal triads towards the centrilobular vein (13). (b) The concentrations of hormone receptors on the surface of hepatocytes may differ in hepatocytes located in the different lobular areas. In the future, it will be crucial to establish the in vivo distribution of receptors or other cellular components involved in the control of gene expression in hepatocytes across a liver lobule.

Summary

Functional heterogeneity in the lobule with regard to plasma protein synthesis is still debated. Therefore, we have localized in liver sections from normal rats and from rats with turpentine-induced AIR the mRNA and protein products of three genes with different alterations in their hepatic expression during an AIR: \( \alpha_2 \)M and \( \alpha_1 \)PI, two positively reacting acute-phase genes, and \( \alpha_1 \)I3, a negative acute-phase reactant. In normal liver, all hepatocytes expressed \( \alpha_2 \)M and \( \alpha_1 \)I3 mRNA, but a preferential expression of \( \alpha_2 \)M and \( \alpha_1 \)I3 mRNA and protein in the PP and ML zones was observed. During an AIR, the level of \( \alpha_2 \)M mRNA increased fourfold in the cytoplasm of PP and ML hepatocytes, while the level of cytoplasmic \( \alpha_1 \)I3 mRNA was decreased about fourfold in the same zones, with parallel variations in the expression of the corresponding proteins. In contrast, no significant modulation of the RNA and protein concentrations of both genes was detected in PV areas. \( \alpha_1 \)PI mRNA was expressed at the same levels in the three lobular zones in normal liver, but staining for the \( \alpha_1 \)PI protein was more intense in the PV zones. During the acute-phase response \( \alpha_1 \)PI mRNA levels were increased twofold in all three lobular zones, and \( \alpha_1 \)PI staining became homogeneous within the lobule. These results demonstrate that the location of a hepatocyte with the liver lobule can influence the expression of the three genes under study both at pre- and post-translational levels, in basal conditions, as well as during modulation of their expression during the inflammatory reaction.

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