Endogenous ecotropic murine leukemia viruses (MuLV) found in low virus strains of mice differ from those found in high virus strains in both their genetic location and stability. Many investigators (1-7) have demonstrated that high virus strains contain multiple ecotropic proviruses, some of which are not allelic among substrains. However, low virus strains of similar genetic origin appear to contain a single ecotropic provirus that is allelic among divergent substrains (8, 9, Jenkins et al., personal communication).

Recently (10), we demonstrated that low virus strains of mice also contain genes that enhance the induction or expression of ecotropic MuLV in certain F1 hybrid mice. This increase in MuLV induction, as compared to the response of either parental strain, occurs both in iododeoxyuridine (IUdR)-treated tissue cultures and spontaneously in spleen cells (J. McCubrey and R. Risser, manuscript submitted for publication). We determined that BALB/c and C57BL/6 (B6) strains each contain one genetic element that, in combination, enhance virus induction, and we denoted the BALB/c locus as \textit{Inc-1} and the B6 locus as \textit{Inb-1}. The enhanced virus induction phenotype is dominant, and the interaction for virus expression is seen in all possible combinations, even when the F1 hybrid is heterozygous at the \textit{Fv-1} locus, a locus known to regulate MuLV replication.

To relate these induction genes to known proviruses, we determined their position in the mouse genome. Ihle and co-workers (11) and Kozak and Rowe (12) mapped the ecotropic virus-inducing gene of BALB/c mice to chromosome 5, 24 centimorgans (cM) from phosphoglucomutase (\textit{Pgmu-1}). Ihle and colleagues (11) also determined that the virus-inducing gene of BALB/c and C3H/He mice are allelic, whereas those of C3H/He and B6 are not (8). Here, we demonstrate that \textit{Inc-1} is allelic among mouse strains A/J, BALB/c, C3H/He, and SEC and maps to chromosome 5 close to \textit{Cv-1}. \textit{Inb-1} is allelic between strains B6 and C57BR and maps to chromosome 8, presumably near \textit{By-1}, the endogenous ecotropic virus-inducing gene of C57BL/10 mice (6).
Allelism Tests for Inc-1 in BALB/c-related Mice

<table>
<thead>
<tr>
<th>Cross</th>
<th>Number of embryos with Inc-1*, Inc-1* phenotype</th>
<th>Number of embryos with Inc-1*, Inc-1* phenotype</th>
<th>95% confidence interval</th>
</tr>
</thead>
<tbody>
<tr>
<td>(BALB/c × A/J) × B6</td>
<td>52/52</td>
<td>0/52</td>
<td>Lower limit to Upper limit</td>
</tr>
<tr>
<td>(BALB/c × C3H/HeJ) × B6</td>
<td>54/54</td>
<td>0/54</td>
<td>6.9 cM to 0 cM</td>
</tr>
<tr>
<td>(BALB/c × SEC) × B6</td>
<td>43/43</td>
<td>0/43</td>
<td>8 cM to 0 cM</td>
</tr>
</tbody>
</table>

* Statistical analysis of allelism and linkage studies. Presented in Tables I-IV are the error intervals for the allelism tests and the SE for the linkage tests. The confidence intervals for the allelism tests are calculated from the formula:

\[
\text{limit}_{\text{upper}} = \frac{[2NP + t^2] + \sqrt{(2NP + t^2)^2 - 4NP^2(N + t^2)}}{2(N + t^2)},
\]

where N is the number of mice examined, P is the observed proportion in a binomial distribution, and t is the confidence level figure. The lower limit is the same as the above formula, but the quantity under the square root is subtracted rather than added. Because no recombinants were observed in the allelism tests, the confidence intervals are merely a function of the desired confidence level and the number of mice examined. The lower limit is a statistical estimate of the furthest possible distance between the two genes under study.

The SE in the linkage tests is calculated by the formula:

\[
\text{SE} = \frac{P(1-P)}{N}^{1/2},
\]

where P is the proportion of A in a binomial population and N is the sample size.

Cell Culture. All cell culture techniques have been described (10). Briefly, 2.5 × 10⁵ secondary embryo cells or tail biopsy fibroblast cells derived from segregating crosses were plated and induced for 48 h with 20 μg/ml of IUdR. 10⁵ SC-1 cells were then added to each culture; cultures were continued for 11 d without trypsinization and then developed in the ultraviolet-XC plaque test. BALB/c, B6, and (BALB/c × B6)F₁ cells show median titers of 0.2 (N = 44), 0.5 (N = 40), or 8 (N = 55) syncytia per 2.5 × 10⁵ induced cells, respectively, in this in situ virus-induction assay (10). Cultures prepared from hybrids within a family of related mouse strains showed no increase in the frequency of cells that produce MuLV, i.e., cells from (BALB/c × A)F₁, (BALB/c × SEC)F₁, and (BALB/c × C3H/He)F₁ mice all gave the same response as the parental cells, or 0-1 syncytia per 2.5 × 10⁵ treated cells. Therefore, we sought to determine by classical genetic techniques whether A/J, C3H/He, and SEC strains have a locus allelic to Inc-1 of BALB/c (10). BALB/c mice were mated to A mice, and this F₁ was mated to B6. Similar tests were performed

Results

Allelism Tests of Inc-1 in BALB/c-related Mice. Previously, we demonstrated that A/J, BALB/c, C3H/He, and SEC mice each carry a gene that in combination with a gene of B6 or C57BR increases the frequency of cells that produce ecotropic MuLV after IUdR induction. Thus, BALB/c or B6 cells show a median titer of ≤0.5 MuLV-induced syncytia in situ per 2.5 × 10⁵ IUdR-treated cells, whereas (BALB/c × B6)F₁ cells yielded a median titer of eight syncytia in situ per 2.5 × 10⁵ treated cells (10). Cultures prepared from hybrids within a family of related mouse strains showed no increase in the frequency of cells that produce MuLV, i.e., cells from (BALB/c × A)F₁, (BALB/c × SEC)F₁, and (BALB/c × C3H/He)F₁ mice all gave the same response as the parental cells, or 0-1 syncytia per 2.5 × 10⁵ treated cells.

Therefore, we sought to determine by classical genetic techniques whether A/J, C3H/He, and SEC strains have a locus allelic to Inc-1 of BALB/c (10). BALB/c mice were mated to A mice, and this F₁ was mated to B6. Similar tests were performed
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TABLE II
Allelism Test for Inb-I in B6 and C57BR

<table>
<thead>
<tr>
<th>Cross</th>
<th>Number of embryos with Inc-I&quot; phenotype</th>
<th>Number of embryos with Inc-I&quot; phenotype</th>
<th>95% confidence interval Lower limit to Upper limit</th>
</tr>
</thead>
<tbody>
<tr>
<td>(B6 x C57BR) x BALB/c</td>
<td>70/70</td>
<td>0/70</td>
<td>5.2 cM to 0 cM</td>
</tr>
</tbody>
</table>

DETERMINATION OF LINKAGE TO KNOWN VIRAL LOCI

![Diagram of linkage between known viral loci](image)

FIG. 1. Location of Co-1 and Be-1 on chromosomes 5 and 8, respectively, and relevant genetic markers.

TABLE III
Linkage of Inc-1 with Pgm-1 and Gus-1 (Chromosome 5)

<table>
<thead>
<tr>
<th></th>
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<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>(BALB/c x SWR) x B6</td>
<td>Inc-I&quot; Pgm-I&quot; Gus-I&quot;</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td>20</td>
</tr>
<tr>
<td></td>
<td>Parental (nonrecombinant)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>15</td>
</tr>
<tr>
<td></td>
<td>Recombinant (Inc-I-Pgm-I)</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>+</td>
<td></td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>Recombinant (Pgm-I-Gus)</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td></td>
<td>6</td>
</tr>
<tr>
<td></td>
<td>Recombinant (Pgm-I-Inv)</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>Double recombinant</td>
<td>-</td>
<td>+</td>
<td>+</td>
<td>-</td>
<td></td>
<td>1</td>
</tr>
<tr>
<td></td>
<td>Number of mice</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
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</tbody>
</table>

Percent recombinant

Inc-I-Pgm-I = 16/70 = 23 cM ± 5.0 cM
Inc-I-Gus = 30/70 = 43 cM ± 5.9 cM
Pgm-I-Gus = 24/70 = 34 cM ± 3.6 cM

Linkage of Inc-1 with Pgm-1 (chromosome 5, 24 cM from Co-1)*

<table>
<thead>
<tr>
<th>Genetic cross:</th>
<th>Number of offspring with:</th>
<th>Pgm-I&quot;/Pgm-I&quot;</th>
<th>Pgm-I&quot;/Pgm-I&quot;</th>
</tr>
</thead>
<tbody>
<tr>
<td>(BALB/c x SWR) X B6</td>
<td>Inc-I&quot;</td>
<td>43</td>
<td>12</td>
</tr>
<tr>
<td></td>
<td>Number of recombinant mice</td>
<td>26/112 (23 cM ± 4 cM)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Total number of mice</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* Some of the mice could not be typed for Gus and Pgm-1, however, 42 additional mice were typed for Pgm-I and Inc-I, as presented in this subtable.

with C3H/He and SEC (in place of the A strain). If the genes were allelic, 100% of the mice should be positive for enhanced virus induction and resemble the (BALB/c X B6)F1. If the loci were not allelic and not closely linked, 75% of the mice should be positive for enhanced virus induction, and 25% should be parental-like. Intermediate values would indicate linkage but not allelism of the two elements.

The results presented in Table I clearly indicate that the Inc-I loci in BALB/c, A/J, C3H/He, and SEC are tightly linked. No recombinants were recovered in the
149 cultures examined, a result entirely consistent with the hypothesis that these genes are at identical locations in these strains.

Allelism of Inb-1 in B6 and C57BR Mice. Because B6 and C57BR mice both possess an In locus that will enhance virus induction in combination with a gene BALB/c and other members of the Inc-I family, we sought to determine whether these Inb loci are allelic in B6 and C57BR strains. B6 mice were mated to C57BR, and this F1 was crossed to BALB/c mice. For reasons similar to those presented above, 100% of the mice should be positive when these Inb-1 loci are allelic. The data presented in Table II indicate that B6 and C57BR are allelic for Inb-1.

Linkage of Inc-1 to Pgm-1 in BALB Mice. Kozak and Rowe (12) and Ihle and co-workers (11) have shown that the BALB/c ecotropic virus-inducing gene Cv-1 is linked to Pgm-1 on chromosome 5 (Fig. 1). Because it is quite possible that Inc-1 is linked to or identical with Cv-1, we examined the linkage of Inc-1 with Pgm-1 and Gus-1. Since BALB/c and B6 mice are Pgm-1 a, a third strain, SWR, (Pgm-1 b) had to be used to test for segregation of Pgm-1. The results of this experiment, presented in Table III, indicate that Inc-1 is ~24 cM from Pgm-1 and is loosely linked to Gus-1. Therefore, Inc-1 is on the centromeric side of Pgm-1 at approximately the position Kozak and Rowe (12) reported for Cv-1.

Discussion

Studies with high virus strains of mice indicate that the proviruses in these strains are located at diverse positions in the mouse genome (1, 5, 7). Our studies indicate that some genes involved in the induction of MuLV in low virus strains of a related pedigree are allelic and probably have not changed position in at least 60 yr of inbreeding. This result is significantly different from AKR proviruses that have been observed to reinsert in additional chromosomal locations during the construction of the NFS-Akv-1 congenic strain that is <10 yr old (3). Presumably, in low virus strains there is a lower probability of infection of germ-line cells and therefore of virus reintegration.

We determined that Inb-1 and Inc-1 are linked to markers known to be linked to
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Be-1 and C\textsubscript{v}-1, the respective ecotropic virus-inducing genes of B6 and BALB/c mice. Given that our Inc-1 linkage data is virtually identical with the map distances between the virus-inducing genes and biochemical markers observed by Kozak and Rowe (12) for the BALB/c virus-inducing gene C\textsubscript{v}-1, it seems reasonable to predict that Inb-1 will be found in the proximity of the C57BL virus-inducing gene Be-1 (6). It is reasonable to propose from our data that the genes governing induction phenotype (In loci) are tightly linked to MuLV structural elements, and this unit is conserved among inbred strains of related pedigree.

However, even though we observed no recombinants in the allelism tests, a statistical analysis of the data indicates the genes of A/J and SEC that govern induction phenotype could be as far as 8 \text{ cM} from the BALB/c Inc-1 locus, a genetic distance that corresponds to \( \sim 5 \times 10^6 \) base pairs. Moreover, statistical analysis of the Pgm-1 linkage data indicates a comparable uncertainty in the linkage of Inc-1 to Pgm-1. From these results, the only valid conclusion is that Inc-1 is near the BALB/c ecotropic virus-inducing gene C\textsubscript{v}-1, and Inc-1 may or may not be identical to C\textsubscript{v}-1. Likewise, there is considerable uncertainty in the allelism of Inb-1 loci in C57BR and B6 mice, and we do not know for certain that Inb-1 is tightly linked to Be-1 because a flanking marker was not readily available in the Inb-1 linkage tests.

Nucleic acid hybridization studies by Horowitz (unpublished data) in this laboratory and by Jenkins and colleagues (personal communication), who compared the size of cellular-ecotropic proviral DNA junction fragments produced by digestion with restriction endonucleases, indicate that inbred strains that carry Inc-1 have a single ecotropic provirus with characteristic cellular flanking sequences, and inbred strain that carry Inb-1 have a single ecotropic provirus with different flanking cellular sequences than those of Inc-1 mice. Because the provirus-cell DNA junction fragments are the same among a family of related mouse strains, these results substantiate our hypothesis that proviruses and their controlling elements are allelic within low virus strains of mice of a common genealogical origin. Analysis of the segregation of proviral sequences with In loci in recombinant inbred strains and segregating generations should further resolve question of identity of proviral sequences and In loci.

The phenotypes associated with a particular ecotropic proviral locus, i.e., pattern of spontaneous and induced expression, and enhanced virus induction in combination with other proviruses, might be explained by two different hypotheses. The site of provirus integration might be important in provirus expression, as originally proposed by Cooper and Temin (13) and further substantiated by Jaenisch et al. (14). One interpretation of studies of Jaenisch and of our studies of virus induction and expression in low leukemic mice (10) is that low virus strains of mice have their proviruses integrated in positions that are restrictive for virus expression, whereas high virus strains have their proviruses in positions permissive for expression. An equally likely possibility is that minor changes within proviral sequences determine their unique biological properties.

Summary

Previously, we identified two genes, termed Inc-1 and Inb-1, that interact to enhance ecotropic murine leukemia virus induction in low virus strains of mice. Mice related to BALB/c in origin carry a locus termed Inc-1, whereas mice related to B6 carry an Inb-1 locus. Mice that carry both Inc-1 and Inb-1 yield 10- to 50-fold more virus-
producing cells than parental strains on induction with halogenated pyrimidines in vitro and demonstrate enhanced murine leukemia virus production in vivo. Here, we show that mice related to BALB/c in origin, i.e., A, C3H/He, and SEC, have an Inc-1 locus that is allelic with that of BALB/c. The C57BR mouse strain has an Inb-1 locus that is allelic with that of B6, located on chromosome 8, 30 cM from Es-1. We also show that the Inc-1 locus of BALB/c mice is located on chromosome 5, 24 cM from Pgm-1 and 43 cM from Gus. Kozak and Rowe (6, 8) and Ihle and co-workers (3) have shown that the ecotropic virus-inducing genes in BALB/c and B10 mice are located on chromosomes 5 and 8, respectively, with similar distances from the previously mentioned biochemical markers. Our data are consistent with two possibilities: Inc-1 and Inb-1 are part of the virus-inducing genes Cv-1 and By-l, respectively, or Inc-1 and Inb-1 are tightly linked regulatory genes.

We thank N. Korn for the excellent isoenzyme typing performed.

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References